

Review Article

Biosynthesis and signalling functions of central and peripheral nervous system neurosteroids in health and disease

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Neurosteroids are steroid hormones synthesised *de novo* in the brain and peripheral nervous tissues. In contrast to adrenal steroid hormones that act on intracellular nuclear receptors, neurosteroids directly modulate plasma membrane ion channels and regulate intracellular signalling. This review provides an overview of the work that led to the discovery of neurosteroids, our current understanding of their intracellular biosynthetic machinery, and their roles in regulating the development and function of nervous tissue. Neurosteroids mediate signalling in the brain via multiple mechanisms. Here, we describe in detail their effects on GABA (inhibitory) and NMDA (excitatory) receptors, two signalling pathways of opposing function. Furthermore, emerging evidence points to altered neurosteroid function and signalling in neurological disease. This review focuses on neurodegenerative diseases associated with altered neurosteroid metabolism, mainly Niemann-Pick type C, multiple sclerosis and Alzheimer disease. Finally, we summarise the use of natural and synthetic neurosteroids as current and emerging therapeutics alongside their potential use as disease biomarkers.

Introduction

It is now accepted, from knowledge accrued over the past 30–40 years, that the brain is a site of *de novo* steroid biosynthesis [1,2]. This means that the brain is a steroidogenic organ akin to the peripheral steroidogenic tissues, namely the adrenal glands, the gonads (ovaries and testes) and also the placenta [3]. The so called ‘neurosteroids’ are generated by largely the same mechanisms, but are produced specifically in the central nervous system (CNS) and the peripheral nervous system (PNS), either endogenously or from circulating sterols [4,5]. The term ‘neurosteroids’, coined in 1981 [4,6], does not refer to a different class of steroids but is instead a catch-all term used to describe androstane and pregnane-derived neurosteroids [7,8] and some of their sulphated forms [8] (Figure 1). Peripheral steroid hormones, such as cortisol, estrogen and testosterone are also neuroactive. However, these mostly act upon intracellular nuclear hormone receptors [9], with only limited evidence of peripheral steroid hormone interaction with plasma membrane proteins [10], to mediate subsequent changes in function of both peripheral and CNS tissues [11]. Neurosteroids are different to peripheral steroid hormones, as several have no identified nuclear hormone receptor (e.g. allopregnanolone) and they generally act directly upon plasma membrane ion channels and on mechanisms of neurotransmitter homeostasis to directly modulate neuronal excitability [12]. This review will focus specifically on CNS neurosteroids, their synthesis and biological roles; the mechanisms of action of steroids in peripheral tissues are not covered here as detailed recent reviews exist [13–15].

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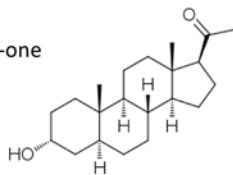
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IUPAC name

(3 α ,5 α)-3-hydroxypregnan-20-one

Common name(s)

5 α -pregnan-3 α -ol-20-one
Tetrahydroprogesterone
Allopregnanolone

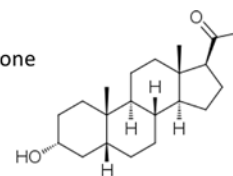


IUPAC name

(3 α ,5 β)-3-hydroxypregnan-20-one

Common name(s)

5 β -pregnan-3 α -ol-20-one
Pregnanolone

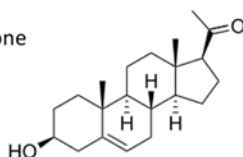


IUPAC name

3 β -3-hydroxypregn-5-en-20-one

Common name(s)

5-Pregnen-3 β -ol-20-one
Pregnenolone

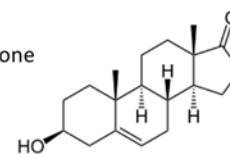


IUPAC name

(3 β)-3-hydroxyandrost-5-en-17-one

Common name(s)

Androst-5-ene-3 β -ol-17-one
Dehydroepiandrosterone

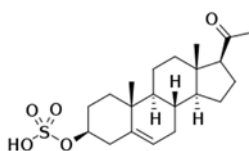


IUPAC name

(3 β)-20-Oxopregn-5-en-3-yl
hydrogen sulfate

Common name(s)

5-Pregnen-3 β -sulfate-20-one
Pregnenolone sulfate



IUPAC name

(3 β)-3-(Sulfooxy) androst-5-en-17-
one

Common name(s)

Androst-5-en-3 β -ol-17-one 3 β -
sulfate
Dehydroepiandrosterone sulfate

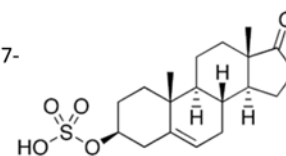


Figure 1. IUPAC and common nomenclature of certain pregnane, androstane and sulphated neurosteroids

The discovery of de novo brain 'neurosteroid' synthesis

The characterisation of the brain as a site of sterol synthesis was pioneered by the group of Baulieu. They identified that brain content of certain steroids and their sulfate esters, particularly dehydroepiandrosterone (DHEA), progesterone and to a lesser extent pregnenolone, did not drop in castrated rats that had also undergone adrenalectomy [4,6]. In contrast, the levels of testosterone and testosterone sulfate were quickly eliminated. When taken together with other findings that (1) the persistency of these brain sterols was unrelated to pools of circulating sterols, as seen by the rapid elimination of peripherally administered radioactive DHEA and pregnenolone in rats [16,17]; (2) the concentration of certain brain sterols is higher in the CNS than in peripheral fluids [17,18]; (3) brain pregnenolone levels are higher in newborn rats for several days despite low levels of sterol biosynthesis in the adrenal organs [4,16]; and (4) a circadian pattern of brain DHEA and pregnenolone exists separate to the periphery [19]. These data suggested that the brain could act as a *de novo* site of sterol synthesis. Subsequent work, detailed below, highlighted that neurosteroid biosynthesis was initially believed to be restricted to glia, which was later expanded to astrocytes, neurons of the hippocampus and cerebellum, and the pineal gland. Brain neurosteroid synthesis has been postulated to be necessary during times when circulating steroid hormone levels are low, e.g. occasionally during the female oestrous cycle, or to compensate for constitutively lower levels of progesterone in males [5]. However, neurosteroids are very lipophilic and are therefore less likely to be secreted into the circulation and more likely to be retained within the membranes of their synthetic cells, indicating that their actions are likely to be more local [20]. The 'local' biological requirement for neurosteroids in the brain is an emerging area, especially regarding signalling (discussed below). The importance of neurosteroid biosynthesis is further evidenced by the high degree of conservation of this process across many species including birds [21], amphibians [22] and fish [23]. This review will focus solely on their roles in mammals.

The neurosteroid biosynthetic pathway

Steroidogenesis in neurons and glia of the CNS and PNS begins following transport of cholesterol, either sourced from *de novo* biosynthesis at the endoplasmic reticulum (ER) or from endocytosed low-density lipoproteins (LDL), into the mitochondria [24,25] (Figure 2). Alternatively, some sterols found in the circulation (e.g. progesterone, DHEA) can cross the blood-brain barrier (reviewed in detail here [26]) and are utilised as precursor molecules in the neurosteroid biosynthetic pathway [27]. The precise contribution of these precursors to neurosteroid synthesis remains to be fully examined [26]. Within the cell, mitochondria are the essential site of neurosteroidogenesis, which involves multiple cytochrome P450 enzymes (Figure 2) [28]. Cholesterol transport from the outer to inner mitochondrial membrane, mediated via the action of the steroidogenic acute regulatory protein (StAR), exerts a

to 5α -dihydroprogesterone by steroid 5α -reductase, one of the first steroidogenic enzyme activities identified in brain tissue [40], in the ER [41], or to 5β -dihydroprogesterone by steroid 5β -reductase [42]. These steroids are then converted to $3\alpha,5\alpha$ -tetrahydroprogesterone (commonly referred to as allopregnanolone) by 3α -hydroxysteroid dehydrogenase (3α -HSD) [9] or $3\beta,5\alpha$ -tetrahydroprogesterone (referred to as pregnanolone) by 3β -HSD [43]. The 5α -reductase enzyme and 3α -HSD operate as a complex that is expressed widely across the brain [44].

The P450c17 enzyme can also convert progesterone into 17-hydroxyprogesterone, which, along with 17-hydroxypregnenolone, can be further converted by P450c17 into androstenedione and DHEA, respectively [45]. The enzyme 17β -hydroxysteroid dehydrogenase (17β -HSD) then converts DHEA and androstenedione into androstenediol and testosterone respectively [46]. The 3β -HSD enzyme can also convert DHEA and androstenediol into androstenedione and testosterone [47]. The cytochrome P450 aromatase enzyme (P450aro) then converts androstenedione and testosterone into estrone and estradiol respectively [48]. Finally, estrone can be converted into estradiol by 17β -HSD [45], whilst testosterone can be converted by 5α -reductase into 5α -dihydrotestosterone (DHT) and then by 3α -HSD into androstenediol [49]. Although not discussed in this review, for information on corticosteroids (Figure 2) and their role in the brain there are recent detailed commentaries [50,51].

Identification of neurons and glia of the CNS and PNS as sites of neurosteroid synthesis and function

At the tissue level, various brain areas have been shown to express StAR, TSPO and the P450scc enzyme [52–55], which exert strong control over the production of pregnenolone. Furthermore, the mRNA of 3β -HSD, responsible for progesterone production, has been detected in various brain areas [23,53,56]. In rat brain sections, glia were first considered as the primary site of neurosteroid synthesis owing to the presence, by immunocytochemistry, of P450scc in the white matter [57]. Furthermore, TSPO immunoreactivity has also been demonstrated in brain white matter [58]. TSPO expression was shown to increase in response to injury, suggesting a role of neurosteroids in brain repair [59,60]. The earlier discoveries of P450scc immunohistochemistry were subsequently supported by RT-PCR [61], with both data sets confirming biochemical observations that mitochondria from the myelin producing oligodendrocytes could convert 3H-mevalonate to pregnenolone [62]. Since these initial studies, the presence of 3β -HSD, 5α -reductase and 3α -HSD have been shown in mixed cultures of oligodendrocytes and astrocytes, pure type I astrocyte cultures, and regions of the brain corresponding to glia and oligodendrocytes [63–68]. These three enzymes sequentially convert pregnenolone into progesterone, 5α -dihydroprogesterone and finally allopregnanolone respectively (Figure 2). Glia are also the only confirmed site (*in vitro*) of DHEA synthesis from pregnenolone [69]. The ability of glial cells to generate neurosteroids may be related to their requirement during differentiation. Gago and colleagues reported in 2001 that the activity of 5α -reductase is 5 times higher in mature oligodendrocytes compared with glial progenitors leading to increased formation of the allopregnanolone precursor 5α -dihydroprogesterone [70]. The production of neurosteroids is not restricted to oligodendrocytes of the CNS. Myelinating PNS Schwann cells also express the relevant enzymes and synthesize neurosteroids (reviewed in [5]) as evidenced by the ~10 times higher levels of progesterone inside these cells than are found in plasma [71]. All myelinating cells are therefore capable of steroidogenesis, which is itself a critical metabolic component of the process of myelination [71]. Indeed, this was evidenced by a seminal study from 1995 using a mouse PNS axon lesion model in which it was shown that levels of pregnenolone and progesterone remain elevated during the process of remyelination [71]. Furthermore, disruption of progesterone biosynthesis or function by small molecule inhibitors results in defective remyelination [71]. The specific roles of neurosteroids in the function of oligodendrocytes and in the process of myelination have been reviewed in more detail elsewhere [72].

Despite a large body of work on the role of glia in neurosteroid biosynthesis, considerably less was initially known about this process in neurons. Studies by Tsutsui and the group of Baulieu were the first to demonstrate the presence of neuronal neurosteroid biosynthesis machinery in the brain. First came the identification of P450scc in cerebellar Purkinje neurons by immunohistochemistry in quail brain [73]. This was followed almost immediately by the identification of 3β -HSD by *in situ* hybridization in neurons throughout the rat brain including in the hippocampus, cortex and cerebellum, with strongest expression confirmed by immunohistochemistry to neurons of the cerebellum [74]. Confirmation of the important role of cerebellar Purkinje neurons in brain neurosteroid production came from further immunostaining and RT-PCR studies demonstrating P450scc expression [55]. Cerebellar Purkinje neurons have one of the most recognised architectures of any neuron with extensively branched dendrites and large cell bodies forming the characteristic Purkinje cell layer that resides between the molecular and the granule cell layers. Purkinje neurons are therefore well equipped and well placed to receive thousands of excitatory inputs from parallel fibres and single climbing fibres of the cerebellum [75]. Remarkably, Purkinje cells, which are GABAergic and therefore inhibit neurotransmission elsewhere in the CNS, are the only neuron that carries outputs from the cerebellum to the cortex

[75]. Although traditionally associated with motor and balance function, the cerebellum also has roles in cognitive and affective behaviour [76]. Subsequently, neurons from various parts of the brain including the hippocampus, dentate gyrus and cerebellar granule layer, as well as Purkinje neurons, have been demonstrated to contain the necessary machinery for *de novo* synthesis of neurosteroids including; pregnenolone, pregnenolone sulfate, progesterone and progesterone metabolites (allopregnanolone) [52]. The expression of StAR, P450_{scc}, 3 β -HSD and 5 α -reductase have been measured in neurons from differentiation through development and into adulthood [52,55,56]. It is known that the requirement for neurosteroids in postnatal development is substantial, with high levels of progesterone and metabolites measured biochemically [55,56,77,78] alongside increasing expression of 3 β -HSD to produce progesterone [56]. Interestingly, neurons have been shown to contain higher amounts of 5 α -reductase activity compared to glia whilst type 1 astrocytes express more 3 α -HSD [66]. As described above, these enzymes form a complex to regulate the production of allopregnanolone, which is believed to be essential for the survival of Purkinje and granule neurons during cerebellar development [79,80]. This observation of differential expression of neurosteroidogenic genes in different cells of the CNS may provide a mechanism for the generation and release of neurosteroids in order to mediate their extracellular signalling effects [81], which are discussed in more detail below. Support for a specific role of neurosteroids in the organisation and development of cerebellar circuitry came from studies showing that progesterone could promote dendritogenesis in Purkinje cells *in vitro* in rat cerebellar slice cultures, and promote Purkinje spine density *in vivo* [77,78]. At the whole organism level, it has been shown that treatment with pregnenolone, and several other neurosteroids, including DHEA and their sulphated derivatives, enhanced task-associated learning during footshock avoidance tests in mice [82]. This is supported by the findings that infusion of pregnenolone sulfate into the nucleus basalis magnocellularis enhanced task associated memory in rats, whilst an infusion of tetrahydroprogesterone had the opposite effect [83]. These effects were explained by the impact of these neurosteroids on GABA_A receptor (GABA_AR) signalling (detailed below), with pregnenolone sulfate being a GABA_AR antagonist and tetrahydroprogesterone being a GABA_AR agonist at the concentrations used in the study.

Beyond neurons and glia, there is also evidence that the pineal gland is able to synthesise neurosteroids. The pineal gland is a small endocrine organ located in the epithalamus, between the cerebellum and the cortex, and is involved in the production of melatonin for the modulation of circadian and seasonal cycles [84]. The pineal gland has been shown to secrete neurosteroids in response to light in chicks, and during quail cerebellar development, potentially promoting Purkinje cell survival by suppressing apoptosis [80,85]. These observations are further supported by the findings that removal of the pineal gland leads to loss of Purkinje neurons during chick development [86]. However, despite the complementary *in vitro* and *in vivo* data, it is still unclear as to whether the CNS functions of neurosteroids are derived solely from synthesis within the CNS or whether there is a contribution of adrenal gland/gonad steroids.

Modulators of neurosteroid biosynthesis

Very little is currently known about whether neurosteroidogenesis is triggered or regulated by extracellular or other physiological stimuli. With respect to intracellular second messengers, rat glia and C6 glioma cells have been shown to respond to elevations in cyclic adenosine monophosphate (cAMP) and subsequent activation of protein kinase A (PKA) mediated signalling pathways by rapidly increasing the transcription of 5 α -reductase mRNA which in turn increases enzyme levels and activity [87,88]. In addition to elevation of 5 α -reductase, StAR and TP50 have both been shown to play a role in neurosteroidogenesis linked to the activation of glial cells [89]. Both are phosphorylated by PKA leading to increased influx of cholesterol into mitochondria via increased activity of StAR and alteration of the ligand binding affinity of TP50 [90]. The neurosteroid estradiol has also been shown, in female rat hypothalamic astrocytes, to bind to the metabotropic glutamate receptor 1 (mGluR1) and induce Ca²⁺ release mediated by inositol 1,4,5-trisphosphate [91], subsequent activation of Ca²⁺ dependent adenylyl cyclase, which in turn elevates cAMP and triggers PKA-mediated phosphorylation of TP50 and StAR [92]. One potential function of this signalling cascade is regulation of the luteinizing hormone surge, which triggers ovulation [93]. Circulating estradiol of the developing follicles triggers synthesis of progesterone, by elevating 3 β -HSD expression, in astrocytes of the female hypothalamus [91]. This in turn induces changes in gonadotropin neuron synaptic plasticity and enhanced secretion of luteinizing hormone [93], which can be blocked by inhibition of progesterone biosynthesis [91]. This example is therefore one demonstration of how peripheral steroid hormones integrate with neurosteroidogenesis and neurosteroid signalling.

Signalling functions of neurosteroids

Predating the knowledge that neurosteroids were synthesized within the brain, reports of brain targeted function of steroids has existed for some time. Steroid hormones have been shown to produce anti-seizure like properties, anticonvulsant, sedative and anaesthetic effects [94–96]. Acute effects of neurosteroids on the brain are typically not mediated by classical steroid hormone mechanisms as neurosteroids are not active upon these intracellular sites;

instead they mediate their effects on neuronal excitability via ion channels and neurotransmitter receptors, primarily GABA_A receptors [97]. Here, we will review the opposing effects of neurosteroids on GABA_A and NMDA receptors [97–99]. Neurosteroids also act on several other ion channel receptors, including glycine, AMPA and sigma receptors, which have recently been reviewed by Rebas et al [100].

GABA

GABA_A receptors (GABA_AR) are the major mediators of inhibitory neurotransmission in the CNS. They are heteropentamers, with abundant receptors comprising α , β and γ or δ subunits. There are a wide array of different GABA_AR configurations, which confer different electrophysiological and pharmacological properties, reviewed here [101]. The activity of GABA_AR is modulated by several classes of molecule, including neurosteroids, benzodiazepines, and barbiturates, which reduce neuronal excitability and mediate anxiolytic and sedative effects. Neurosteroids can be positive or negative modulators of GABA_AR activity. Potentiating neurosteroids, such as allopregnanolone, tetrahydrodeoxycorticosterone (THDOC) and androstanediol, are positive modulators of GABA_AR activity with two distinct modes of action. At high concentrations (>100 nM), neurosteroids can directly activate GABA_AR [102,103]. However, neurosteroids are also potent positive modulators of GABA_AR activity, acting via two different mechanisms depending on ambient GABA concentration. Low nanomolar neurosteroid concentrations extend the open time of GABA_AR exposed to a low concentration of GABA for a prolonged period, and thus enhance inhibitory neurotransmission [104]. This is physiologically relevant to extrasynaptic GABA_AR, which experience these conditions. In studies that mimic the short exposure to saturating levels of GABA relevant to synaptic GABA_AR, neurosteroids extend the delay of GABA_AR mediated current, with a delay to receptor recycling a proposed mechanism [105]. Inhibitory neurosteroids, which mostly comprise sulphated neurosteroids such as pregnenolone sulfate and DHEA sulfate, act as non-competitive GABA_AR antagonists [99,106,107].

Neurosteroids can influence a wide range of GABA_AR subtypes; this is supported by the identification of the proposed potentiating neurosteroid-binding site close to the membrane between subunits [108]. A homopentameric α subunit transmembrane domain construct has been crystallised with the potentiating neurosteroid THDOC, which revealed binding between the subunits. This binding site has been modelled in the most abundant $\alpha 1\beta 2\gamma 2$ arrangement, revealing that binding pockets capable of accommodating neurosteroids in the correct binding orientation are found between all the subunits [109]. This is different to a previously proposed neurosteroid binding site in the transmembrane domains of the α subunit [110], but is the only site supported by structural data. The promiscuity of neurosteroids with regard to GABA_AR subtypes is therefore not surprising. There are, however, differences in the extent to which different GABA_AR subtypes are modulated. At low ambient GABA levels, such as those typically experienced by extrasynaptic GABA_AR, neurosteroids mediate their potentiating effects mostly via GABA_AR that contain a δ subunit; these effects are almost abolished in neurons from mice lacking the δ subunit [111–113]. This difference is not thought to be due to neurosteroid binding by the δ subunit, but is rather to do with the mechanism of activation of δ containing GABA_AR by GABA [110]. GABA_AR containing δ subunits are located extrasynaptically, where they mediate tonic inhibitory currents. Although GABA binds to these receptors with high affinity, it only acts as a partial agonist. It is thought that neurosteroids increase the likelihood of channel opening following GABA binding, thus converting GABA to a full agonist [114]. As GABA already acts as a full agonist at γ containing GABA_AR, there is less scope for this type of potentiation.

The site of inhibitory neurosteroid binding to GABA_AR is unknown. However, it is thought to be outside the pore, and distinct from the binding sites of both potentiating neurosteroids and picrotoxin (GABA_AR open channel blocker) binding [115]. The evidence for this is that inhibition is not voltage-dependent, as would be expected for an open channel blocker, the inhibitory effects of pregnenolone sulfate and picrotoxin are additive, and, unlike potentiating neurosteroids, pregnenolone sulfate on the inner membrane leaflet cannot affect GABA_AR. Neurosteroid mediated GABA_AR inhibition is more effective at higher GABA concentrations, suggesting that the binding site is exposed by receptor activation [116]. This is supported by the greater inhibition of steady state, rather than peak, current through GABA_AR. Similarly to potentiating neurosteroids, inhibitory neurosteroids act across a wide range of GABA_AR configurations, but, unlike potentiating neurosteroids, are more effective at GABA_AR that contain γ subunits [117,118].

NMDA

NMDA receptors (NMDAR) are a class of ionotropic glutamate receptor, found pre-synaptically, post-synaptically and extra-synaptically, that mediate excitatory neurotransmission. They take the form of heterotetramers, made up of a dimer of dimers, with each dimer comprised of a GluN1 and GluN2 or GluN3 subunit. Variation within each subunit class further increases NMDA receptor diversity. Unlike other glutamate receptors, NMDA receptors require

co-stimulation with glycine or D-serine for activation [119]. As with GABA_AR, neurosteroids can both positively and negatively modulate receptor activity. The sulphated neurosteroid pregnanolone sulfate is a non-competitive NMDAR inhibitor, with the greatest inhibitory effects on receptors containing the GluN2C and GluN2D subunits [120,121]. This inhibitory effect is achieved via decreasing channel open probability and open time, and by increasing receptor desensitisation [122].

In contrast to GABA_AR, where sulphated neurosteroids act solely as inhibitors, pregnenolone sulfate has pleiotropic effects on NMDAR. Classically, it has been confirmed numerous times to potentiate NMDAR activity, including [98,120,123–126]. This potentiation is subtype-dependent, acting only on receptors that contain the GluN2A or GluN2B subunits, with the greatest effect on GluN2A-containing receptors [98,120,126]. Pregnenolone sulfate potentiates NMDA receptors by increasing channel open probability and slowing receptor deactivation [98,126]. The greater effect on GluN2A containing receptors is due to their faster native deactivation rate; the rate for GluN2A and GluN2B containing receptors is similar in the presence of pregnenolone sulfate [98]. Also in contrast to the situation with GABA_AR, the potentiating effect of pregnenolone sulfate on NMDAR is dependent on binding to inactive receptors. The potentiating effect of pregnenolone sulfate is much greater when cells are pre-treated than when it is co-applied with glutamate, suggesting that it cannot bind to activated receptors [127]. In addition to this potentiation, pregnenolone sulfate can directly activate GluN1/2A and GluN1/2B NMDAR [128], making its actions on these NMDAR analogous to the actions of potentiating neurosteroid modulation of GABA_AR. The final modulation of NMDAR by pregnenolone sulfate is as an inhibitor; it inhibits NMDAR that contain GluN2C and GluN2D subunits [98,120,127].

The mechanistic basis of neurosteroid modulation of NMDAR is unclear. Pregnenolone sulfate and pregnanolone sulfate bind at different sites, and exert their effect from the outer leaflet of the plasma membrane, suggesting that the site of synthesis and site of action are distinct [124]. More specifically, the M3-M4 extracellular loop of GluN2 subunits has been found to be vital for neurosteroid modulation of NMDAR [126,129]. Pregnanolone sulfate inhibition of NMDAR is voltage-independent, suggesting that it binds outside the pore region, but is usage-dependent, with binding only seen in the presence of receptor agonist [129]. Modelling studies suggest that pregnanolone sulfate binds extracellularly to resting receptors, and that agonist binding causes a conformational change, allowing pregnanolone sulfate to move into the pore region and block ion conductance [130]. More is known about pregnenolone sulfate binding. A steroid modulatory domain has been identified in the GluN2B subunit, comprising parts of the glutamate binding site, and the M4 transmembrane region. Models indicate that residues in this domain contribute to a hydrophobic binding pocket between subunits that could accommodate pregnenolone sulfate [131]. More recently, pregnenolone sulfate and pregnanolone sulfate have been shown to bind different sites in the extracellular amino terminus of GluN2D, and to the S1/S2 domain of GluN2B [132]. However, as these experiments used soluble fragments and saturating neurosteroid concentrations, the physiological relevance of this binding is uncertain.

Impact of neurosteroids in neurodegenerative disease

Diseases of steroid biosynthesis and transport

There are surprisingly few studies on how disrupted sterol biosynthesis or sterol transport associated to human diseases impact upon neurosteroid levels and their signalling functions. Sterols are either created *de novo* within cells via the action of 3-hydroxy-3-methyl-glutaryl coenzyme A (HMG CoA) reductase to generate mevalonate or are brought into the cell via LDL receptors [133]. With respect to the LDL receptor pathway, apolipoprotein E (ApoE), which binds with high affinity to the LDL receptor, is associated with Alzheimer disease (AD) and altered neurosteroid levels (described in detail below). At the lysosome, LDL receptors release their cargo and free cholesterol is liberated by the action of acid lipase [133]. To our knowledge, no studies have been reported determining whether neurosteroids are altered in lysosomal acid lipase deficiency, a lysosomal storage disease with accumulation of cholesteryl esters and triglycerides in cells and tissues [134]. Once cholesterol is released, it is mobilised within the lumen of lysosomes by the NPC2 protein, which delivers the cholesterol to the lysosomal transmembrane protein NPC1 [135]. This protein, or others such as LIMP2, may transfer the cholesterol out of lysosomes via ER-lysosome contact points [136]. Loss-of-function mutations in the genes that encode either NPC1 or NPC2 result in the neurodegenerative lysosomal multi-lipid storage disease Niemann-Pick type C (NPC) [135]. Lysosomal cholesterol storage in NPC disease was found to associate with age-dependent reductions in the activity of 3 α -HSD [79] and the biochemical levels of estradiol [92] in the brains of the NPC1 protein deficient *Npc1^{nih}* mouse model. The deficiency in estradiol secretion by the *Npc1^{-/-}* mouse glia was shown in co-culture experiments to be less supportive of the growth of wild-type neurons, with fewer and shorter neurites measured [92]. A more substantial role for neurosteroids in the pathogenesis of NPC disease was suggested in Griffin et al. whereby treatment with allopregnanolone, solubilised in cyclodextrin,

could compensate for functional absence of the synthesis of this neurosteroid. With either a single post-natal day 7 injection [79], or in a subsequent study by multiple injections [137], allopregnanolone solubilised in a 20% (w/v) solution of 2-hydroxypropyl- β -cyclodextrin (HP β CD) led to a dramatic increase in the life expectancy of the mouse model from 67 days to 124 days by single injection or even longer by multiple injections. However, subsequent studies by the groups of Dietschy and then Walkley identified that this effect was mostly mediated by the cyclodextrin carrier alone, with only a small additive effect when both HP β CD and allopregnanolone were tested together versus the HP β CD vehicle alone [138,139]. These findings, alongside many others, ultimately led to a clinical trial of HP β CD in NPC patients [140,141]. Furthermore, allopregnanolone administered in complex with other solubilising agents such as corn oil had no therapeutic effect on disease phenotypes or life expectancy [139], suggesting that the efficacious molecule in all NPC disease studies whereby allopregnanolone is combined with HP β CD is, in fact, the HP β CD carrier. In the case of estradiol, a subcutaneous injection of 2 or 20 mg/kg estradiol solubilised in corn oil at post-natal day 7 leads to a modest improvement in life expectancy alongside some functional improvement on the rota-rod test [142]. It therefore appears, for now, that the impact of deficient neurosteroid biosynthesis on NPC disease pathogenesis is relatively minimal [79,143]. Beyond the endosomal system, cholesterol is transported to lysosomes by StARD proteins, such as StARD3, and then into the inner mitochondrial membrane as outlined above. Diseases associated with STAR and P450scc have been recently reviewed in detail elsewhere [143], whilst diseases of the steroid hormone biosynthesis pathway, including mutations in the *CYP7B1* gene, encoding 25-hydroxycholesterol 7 α -hydroxylase, which cause hereditary spastic paraplegia [144], have also been reviewed [145].

Multiple sclerosis, neurosteroids and myelination

Multiple sclerosis (MS) is a grey and white matter demyelinating disease of the CNS, characterised by episodes of relapse and remission. The disease is common, with an estimated incidence of 1/320 in the U.S.A. [146]. At the cellular level, MS presents with oligodendrocyte dysfunction and loss, demyelination and axon injury, neurodegeneration and subsequent neuroinflammation [147]. Steroid hormones have been implicated in MS as a result of the findings that (1) disease relapse is less likely to occur during the final 3 months of pregnancy and more likely to relapse during the first 3 months after birth, which fits with elevated and then reduced levels of oestrogen and progesterone respectively [148] and (2) incidence and progression are sex-dependent [149]. In addition, the brains of MS patients and the MS mouse model (experimental autoimmune encephalomyelitis) present with reduced allopregnanolone and DHEA levels, alongside reduced expression of neurosteroid biosynthetic enzymes [147,150]. Furthermore, treating MS model mice with progesterone led to a correction in neurosteroid production [151,152], reduced neuroinflammation and reduced demyelination [151,153]. Although multiple therapeutic options exist for MS, neurosteroids appear to be another promising avenue for treating this disease.

Alzheimer disease

An estimated ~47 million people worldwide are living with dementia, of which Alzheimer disease (AD) is the most common cause (60–80%) [154]. Classically, the disease is characterised by extracellular plaques composed of amyloid beta (β -amyloid) peptide and intracellular neurofibrillary tangles of the tau protein [154]. In addition to rare genetic forms of AD, the only gene initially associated to increased risk of AD was *APOE4*, encoding an apolipoprotein lipid carrier associated with cholesterol transport [154]. More recently, GWAS studies have identified components of the sterol pathway as increasing the risk of developing AD, all of which point to a role for sterol homeostasis in AD pathophysiology [155]. With respect to neurosteroids, lower levels of allopregnanolone (and other neurosteroids) have been shown in AD patient plasma and serum [156,157]. Furthermore, lower levels of allopregnanolone have also been shown in post-mortem samples from the prefrontal and temporal cortex of male and female AD patients compared with controls. These changes were inversely correlated to the Braak disease stage [158,159] and positively correlated with the *APOE4* AD risk gene [159]. In contrast to the lower levels of allopregnanolone, higher levels of pregnenolone and DHEA have been measured in both cerebrospinal fluid and postmortem brain tissue of AD patients, which correlate with the Braak stage of pathology [158–162]. Both pregnenolone and DHEA have been shown to provide a protective effect against cellular [163–165] and *in vivo* [166] toxicity induced by the β -amyloid protein, suggesting a protective effect. However, as levels of DHEA have been shown to decrease naturally with age [167], and as β -amyloid addition to cells triggers an increase in levels of DHEA [161], it is currently unclear whether the increase in DHEA in AD samples is a compensatory mechanism or a direct component of the disease. As the sterol pathway has been implicated in the risk of AD [155] and as the β -amyloid protein can bind to sterols [168], it is tempting to speculate that neurosteroids may be directly implicated in AD pathogenesis.

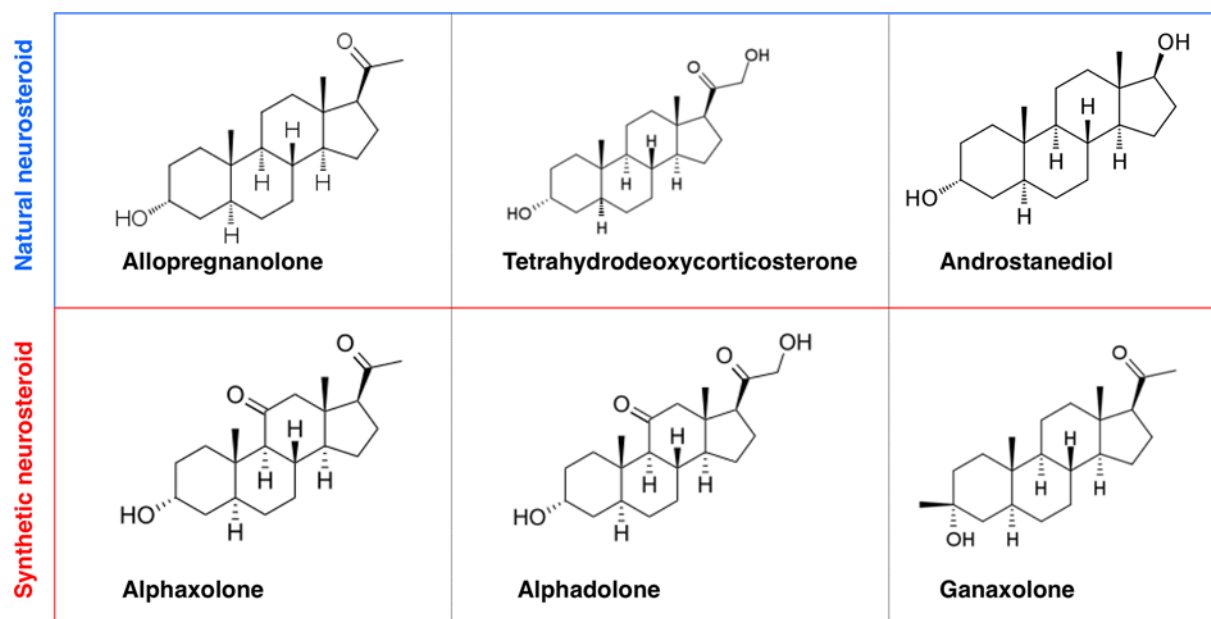


Figure 3. Synthetic neurosteroids used in therapeutic studies and their natural neurosteroid analogue

Neurosteroids as therapeutic molecules and biomarkers

Multiple synthetic analogues of neurosteroids exist (Figure 3), some of which have previously been in widespread therapeutic use since the 1970s, namely alphaxalone and alphadolone [169] dissolved in 20% solution of polyoxyethylated castor oil surfactant [170]. A combination of two neurosteroids were used as alphadolone was less potent than, but increased the solubility of, alphaxalone, which is a powerful but short acting anaesthetic with minimal side effects [170]. However, this synthetic neurosteroid mixture was removed from human clinical use in the 1980s owing to adverse anaphylactic effects incurred by the solubilising agent. However, it remains in veterinary use under the trade name of Saffan [169]. More recently, the synthetic neurosteroid ganaxolone, a GABA_AR modulator, has been in clinical trials for epilepsy, as it protects against seizures in animal models, with some evidence of efficacy in humans [171]. Several other neurosteroid analogues have been in clinical trials (e.g. for Fragile X syndrome and autism), although with mixed outcomes, reviewed here [169,172–174]. The implications of the use of various neurosteroids as therapeutic molecules in several human diseases and medical conditions have been described in detail elsewhere [169] and are therefore not covered in any further detail here. In conjunction with the development of neurosteroids as therapeutics, their use as disease biomarkers has also been suggested. This avenue of research appears to be supported by the findings that cerebrospinal fluid levels of pregnenolone and DHEA correlate with concentrations within the temporal cortex and that both are elevated in AD patient cohorts [160]. Further evidence from rodent experiments indicates that changes in serum pregnenolone levels correlate sufficiently with concentrations in the hippocampus [175]. However, plasma levels of DHEA and its sulphated form have been reported to be lower in AD patients [176]. Furthermore, any measurements may be clouded by the transfer of peripheral blood steroids into the brain [177], alongside the aforementioned potential for errors in measurement of some sulphated neurosteroids [8].

Conclusions and perspectives

Despite significant progress in our understanding of neurosteroids over the last 30+ years, much work remains to fully elucidate their roles in maintaining healthy function of the CNS and PNS. There is evident therapeutic potential for neurosteroids, as exemplified by the recent approval of brexanolone, an allopregnanolone formulation, for postpartum depression [178]. However, this potential remains largely under-developed. Difficulties associated with their nomenclature, measurement, reported effects and development as therapeutic molecules may be hindering the expansion, and evident potential, of this field.

Summary

- Neurosteroids are generated *de novo* in peripheral and central nervous system tissues.
- Neurosteroids, in contrast to peripherally generated adrenal steroid hormones, directly target extracellular receptors rather than mediate their effects via classical intracellular steroid hormone nuclear receptors.
- Neurosteroids regulate developmental, growth and signalling functions of the CNS and PNS.
- Neurosteroids have emerging roles in neurological disease and have potential to be utilised therapeutically.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

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Abbreviations

3 α -HSD, 3 α -hydroxysteroid dehydrogenase; 3 β -HSD, 3 β -hydroxysteroid dehydrogenase; 17 β -HSD, 17 β -hydroxysteroid dehydrogenase; AD, Alzheimer disease; AllotetrahydroDOC, allotetrahydrodeoxycorticosterone; ApoE, apolipoprotein E; cAMP, cyclic adenosine monophosphate; CNS, central nervous system; CRAC, cholesterol recognition/interaction amino acid consensus; DHEA, dehydroepiandrosterone; DHEAS, dehydroepiandrosterone sulphate; DHT, 5 α -dihydrotestosterone; ER, endoplasmic reticulum; GABA, γ -aminobutyric acid; GABA_AR, γ -aminobutyric acid receptors, type A; GWAS, genome wide association study; HMG CoA, 3-hydroxy-3-methyl-glutaryl coenzyme A; HP β CD, 2-hydroxypropyl- β -cyclodextrin; LDL, low density lipoprotein; LIMP, lysosomal integrated membrane protein; mGluR1, metabotropic glutamate receptor 1; MS, multiple sclerosis; NMDA, N-methyl-D-aspartic acid; NMDAR, NMDA receptor; NPC, Niemann-Pick type C; P450aro, cytochrome P450 aromatase; P450c11 β , sterol 11 β -hydroxylase; P450c17, cytochrome P450 17A1/17 α -hydroxylase; P450c21, steroid 21-hydroxylase; P450scc, cytochrome P450 side chain cleavage; PKA, protein kinase A; PNS, peripheral nervous system; PREGS, pregnenolone sulphate; RT-PCR, reverse transcriptase polymerase chain reaction; StAR, steroidogenic acute regulatory protein; StARD, steroidogenic acute regulatory domain; THDOC/TetrahydroDOC, tetrahydrodeoxycorticosterone; TSPO, translocator protein.

References

- 1 Tsutsui, K. (2019) Kobayashi award: Discovery of cerebellar and pineal neurosteroids and their biological actions on the growth and survival of Purkinje cells during development (review). *Gen. Comp. Endocrinol.* **284**, 113051, <https://doi.org/10.1016/j.ygcen.2018.10.014>
- 2 Baulieu, E.E., Robel, P. and Schumacher, M. (2001) Neurosteroids: beginning of the story. *Int. Rev. Neurobiol.* **46**, 1–32, [https://doi.org/10.1016/S0074-7742\(01\)46057-0](https://doi.org/10.1016/S0074-7742(01)46057-0)
- 3 Payne, A.H. and Hales, D.B. (2004) Overview of steroidogenic enzymes in the pathway from cholesterol to active steroid hormones. *Endocr. Rev.* **25**, 947–970, <https://doi.org/10.1210/er.2003-0030>
- 4 Baulieu, E.E. (1998) Neurosteroids: a novel function of the brain. *Psychoneuroendocrinology* **23**, 963–987, [https://doi.org/10.1016/S0306-4530\(98\)00071-7](https://doi.org/10.1016/S0306-4530(98)00071-7)

- 5 Baulieu, E. and Schumacher, M. (2000) Progesterone as a neuroactive neurosteroid, with special reference to the effect of progesterone on myelination. *Steroids* **65**, 605–612, [https://doi.org/10.1016/S0039-128X\(00\)00173-2](https://doi.org/10.1016/S0039-128X(00)00173-2)
- 6 Corpechot, C., Robel, P., Axelson, M., Sjovall, J. and Baulieu, E.E. (1981) Characterization and measurement of dehydroepiandrosterone sulfate in rat brain. *Proc. Natl. Acad. Sci. U.S.A.* **78**, 4704–4707, <https://doi.org/10.1073/pnas.78.8.4704>
- 7 Maninger, N., Wolkowitz, O.M., Reus, V.I., Epel, E.S. and Mellon, S.H. (2009) Neurobiological and neuropsychiatric effects of dehydroepiandrosterone (DHEA) and DHEA sulfate (DHEAS). *Front. Neuroendocrinol.* **30**, 65–91, <https://doi.org/10.1016/j.yfrne.2008.11.002>
- 8 Schumacher, M., Liere, P., Akwa, Y., Rajkowski, K., Griffiths, W., Bodin, K. et al. (2008) Pregnenolone sulfate in the brain: a controversial neurosteroid. *Neurochem. Int.* **52**, 522–540, <https://doi.org/10.1016/j.neuint.2007.08.022>
- 9 Penning, T.M., Jin, Y., Steckelbroeck, S., Lanisnik Rizner, T. and Lewis, M. (2004) Structure-function of human 3 alpha-hydroxysteroid dehydrogenases: genes and proteins. *Mol. Cell. Endocrinol.* **215**, 63–72, <https://doi.org/10.1016/j.mce.2003.11.006>
- 10 Doroudi, M., Schwartz, Z. and Boyan, B.D. (2015) Membrane-mediated actions of 1,25-dihydroxy vitamin D3: a review of the roles of phospholipase A2 activating protein and Ca(2+)/calmodulin-dependent protein kinase II. *J. Steroid Biochem. Mol. Biol.* **147**, 81–84, <https://doi.org/10.1016/j.jsbmb.2014.11.002>
- 11 Olivares, A.M., Moreno-Ramos, O.A. and Haider, N.B. (2015) Role of Nuclear Receptors in Central Nervous System Development and Associated Diseases. *J. Exp. Neurosci.* **9**, 93–121, <https://doi.org/10.4137/JEN.S25480>
- 12 Schverer, M., Lanfumey, L., Baulieu, E.E., Froger, N. and Villey, I. (2018) Neurosteroids: non-genomic pathways in neuroplasticity and involvement in neurological diseases. *Pharmacol. Ther.* **191**, 190–206, <https://doi.org/10.1016/j.pharmthera.2018.06.011>
- 13 Amenyogbe, E., Chen, G., Wang, Z., Lu, X., Lin, M. and Lin, A.Y. (2020) A Review on Sex Steroid Hormone Estrogen Receptors in Mammals and Fish. *Int. J. Endocrinol.* **2020**, 5386193, <https://doi.org/10.1155/2020/5386193>
- 14 Evans, R.M. and Mangelsdorf, D.J. (2014) Nuclear Receptors, RXR, and the Big Bang. *Cell* **157**, 255–266, <https://doi.org/10.1016/j.cell.2014.03.012>
- 15 Le Dily, F. and Beato, M. (2018) Signaling by Steroid Hormones in the 3D Nuclear Space. *Int. J. Mol. Sci.* **19**, <https://doi.org/10.3390/ijms19020306>
- 16 Robel, P. and Baulieu, E.E. (1985) Neuro-steroids: 3?-hydroxy-(5)-derivatives in the rodent brain. *Neurochem. Int.* **7**, 953–958, [https://doi.org/10.1016/0197-0186\(85\)90143-3](https://doi.org/10.1016/0197-0186(85)90143-3)
- 17 Corpechot, C., Synguelakis, M., Talha, S., Axelson, M., Sjovall, J., Vihko, R. et al. (1983) Pregnenolone and its sulfate ester in the rat brain. *Brain Res.* **270**, 119–125, [https://doi.org/10.1016/0006-8993\(83\)90797-7](https://doi.org/10.1016/0006-8993(83)90797-7)
- 18 Lacroix, C., Fiet, J., Benais, J.P., Gueux, B., Bonete, R., Villette, J.M. et al. (1987) Simultaneous radioimmunoassay of progesterone, androst-4-enedione, pregnenolone, dehydroepiandrosterone and 17-hydroxyprogesterone in specific regions of human brain. *J. Steroid. Biochem.* **28**, 317–325, [https://doi.org/10.1016/0022-4731\(87\)91025-9](https://doi.org/10.1016/0022-4731(87)91025-9)
- 19 Synguelakis, M., Halberg, F., Baulieu, E.E. and Robel, P. (1985) Circadian course of delta 5,3 beta-hydroxysteroids and glucocorticosteroids in the plasma and brain of rats. *C. R. Acad. Sci. III* **301**, 823–826
- 20 Chisari, M., Eisenman, L.N., Covey, D.F., Mennerick, S. and Zorumski, C.F. (2010) The sticky issue of neurosteroids and GABA(A) receptors. *Trends Neurosci.* **33**, 299–306, <https://doi.org/10.1016/j.tins.2010.03.005>
- 21 Tsutsui, K. and Yamazaki, T. (1995) Avian neurosteroids. I. Pregnenolone biosynthesis in the quail brain. *Brain Res.* **678**, 1–9, [https://doi.org/10.1016/0006-8993\(95\)00116-8](https://doi.org/10.1016/0006-8993(95)00116-8)
- 22 Mensah-Nyagan, A.G., Feuilloley, M., Dupont, E., Do-Rego, J.L., Le Boulenger, F., Pelletier, G. et al. (1994) Immunocytochemical localization and biological activity of 3 beta-hydroxysteroid dehydrogenase in the central nervous system of the frog. *J. Neurosci.* **14**, 7306–7318, <https://doi.org/10.1523/JNEUROSCI.14-12-07306.1994>
- 23 Sakamoto, H., Ukena, K. and Tsutsui, K. (2001) Activity and localization of 3beta-hydroxysteroid dehydrogenase/ Delta5-Delta4-isomerase in the zebrafish central nervous system. *J. Comp. Neurol.* **439**, 291–305, <https://doi.org/10.1002/cne.1351>
- 24 Platt, F.M., Wassif, C., Colaco, A., Dardis, A., Lloyd-Evans, E., Bembli, B. et al. (2014) Disorders of cholesterol metabolism and their unanticipated convergent mechanisms of disease. *Annu. Rev. Genomics Hum. Genet.* **15**, 173–194, <https://doi.org/10.1146/annurev-genom-091212-153412>
- 25 Miller, W.L. and Bose, H.S. (2011) Early steps in steroidogenesis: intracellular cholesterol trafficking. *J. Lipid Res.* **52**, 2111–2135, <https://doi.org/10.1194/jlr.R016675>
- 26 Grube, M., Hagen, P. and Jedlitschky, G. (2018) Neurosteroid Transport in the Brain: Role of ABC and SLC Transporters. *Front Pharmacol.* **9**, 354, <https://doi.org/10.3389/fphar.2018.00354>
- 27 Baulieu, E.E. (1997) Neurosteroids: of the nervous system, by the nervous system, for the nervous system. *Recent Prog. Horm. Res.* **52**, 1–32
- 28 Omura, T. (2006) Mitochondrial P450s. *Chem. Biol. Interact.* **163**, 86–93, <https://doi.org/10.1016/j.cbi.2006.06.008>
- 29 Lin, D., Sugawara, T., Strauss, 3rd, J.F., Clark, B.J., Stocco, D.M., Saenger, P. et al. (1995) Role of steroidogenic acute regulatory protein in adrenal and gonadal steroidogenesis. *Science* **267**, 1828–1831, <https://doi.org/10.1126/science.7892608>
- 30 Papadopoulos, V., Amri, H., Boujrad, N., Cascio, C., Culty, M., Garnier, M. et al. (1997) Peripheral benzodiazepine receptor in cholesterol transport and steroidogenesis. *Steroids* **62**, 21–28, [https://doi.org/10.1016/S0039-128X\(96\)00154-7](https://doi.org/10.1016/S0039-128X(96)00154-7)
- 31 Lacapere, J.J. and Papadopoulos, V. (2003) Peripheral-type benzodiazepine receptor: structure and function of a cholesterol-binding protein in steroid and bile acid biosynthesis. *Steroids* **68**, 569–585, [https://doi.org/10.1016/S0039-128X\(03\)00101-6](https://doi.org/10.1016/S0039-128X(03)00101-6)
- 32 Li, F., Liu, J., Zheng, Y., Garavito, R.M. and Ferguson-Miller, S. (2015) Protein structure. Crystal structures of translocator protein (TSPO) and mutant mimic of a human polymorphism. *Science* **347**, 555–558, <https://doi.org/10.1126/science.1260590>
- 33 Banati, R.B., Middleton, R.J., Chan, R., Hatty, C.R., Kam, W.W., Quin, C. et al. (2014) Positron emission tomography and functional characterization of a complete PBR/TSPO knockout. *Nat. Commun.* **5**, 5452, <https://doi.org/10.1038/ncomms6452>
- 34 Tuckey, R.C. and Cameron, K.J. (1993) Catalytic properties of cytochrome P-450sc purified from the human placenta: comparison to bovine cytochrome P-450sc. *Biochim. Biophys. Acta* **1163**, 185–194, [https://doi.org/10.1016/0167-4838\(93\)90180-Y](https://doi.org/10.1016/0167-4838(93)90180-Y)

- 35 Simard, J., Ricketts, M.L., Gingras, S., Soucy, P., Feltus, F.A. and Melner, M.H. (2005) Molecular biology of the 3beta-hydroxysteroid dehydrogenase/delta5-delta4 isomerase gene family. *Endocr. Rev.* **26**, 525–582, <https://doi.org/10.1210/er.2002-0050>
- 36 Thomas, J.L., Frieden, C., Nash, W.E. and Strickler, R.C. (1995) An NADH-induced conformational change that mediates the sequential 3 beta-hydroxysteroid dehydrogenase/isomerase activities is supported by affinity labeling and the time-dependent activation of isomerase. *J. Biol. Chem.* **270**, 21003–21008, <https://doi.org/10.1074/jbc.270.36.21003>
- 37 Prasad, M., Thomas, J.L., Whittal, R.M. and Bose, H.S. (2012) Mitochondrial 3beta-hydroxysteroid dehydrogenase enzyme activity requires reversible pH-dependent conformational change at the intermembrane space. *J. Biol. Chem.* **287**, 9534–9546, <https://doi.org/10.1074/jbc.M111.333278>
- 38 Ian Mason, J. (1993) The 3beta-hydroxysteroid dehydrogenase gene family of enzymes. *Trends Endocrinol. Metab.* **4**, 199–203, [https://doi.org/10.1016/1043-2760\(93\)90117-W](https://doi.org/10.1016/1043-2760(93)90117-W)
- 39 Lorence, M.C., Naville, D., Graham-Lorence, S.E., Mack, S.O., Murry, B.A., Trant, J.M. et al. (1991) 3 beta-hydroxysteroid dehydrogenase/delta 5—4-isomerase expression in rat and characterization of the testis isoform. *Mol. Cell. Endocrinol.* **80**, 21–31, [https://doi.org/10.1016/0303-7207\(91\)90139-J](https://doi.org/10.1016/0303-7207(91)90139-J)
- 40 Naftolin, F., Ryan, K.J. and Petro, Z. (1971) Aromatization of androstenedione by the diencephalon. *J. Clin. Endocrinol. Metab.* **33**, 368–370, <https://doi.org/10.1210/jcem-33-2-368>
- 41 Eicheler, W., Tuohimaa, P., Vilja, P., Adermann, K., Forssmann, W.G. and Aumuller, G. (1994) Immunocytochemical localization of human 5 alpha-reductase 2 with polyclonal antibodies in androgen target and non-target human tissues. *J. Histochem. Cytochem.* **42**, 667–675, <https://doi.org/10.1177/42.5.8157936>
- 42 Lisboa, B.P., Strassner, M., Wulff, C. and Hoffmann, U. (1974) 5β Reductase in the human fetal brain. *Acta Endocrinologica (Copenh.)* **75**, 156, <https://doi.org/10.1530/acta.0.077S156>
- 43 Dupont, E., Simard, J., Luu-The, V., Labrie, F. and Pelletier, G. (1994) Localization of 3 beta-hydroxysteroid dehydrogenase in rat brain as studied by in situ hybridization. *Mol. Cell. Neurosci.* **5**, 119–123, <https://doi.org/10.1006/mcne.1994.1014>
- 44 Steckelbroeck, S., Watzka, M., Reichelt, R., Hans, V.H., Stoffel-Wagner, B., Heidrich, D.D. et al. (2001) Characterization of the 5alpha-reductase-3alpha-hydroxysteroid dehydrogenase complex in the human brain. *J. Clin. Endocrinol. Metab.* **86**, 1324–1331, <https://doi.org/10.1210/jcem.86.3.7325>
- 45 Fevold, H.R., Lorence, M.C., McCarthy, J.L., Trant, J.M., Kagimoto, M., Waterman, M.R. et al. (1989) Rat P450(17 alpha) from testis: characterization of a full-length cDNA encoding a unique steroid hydroxylase capable of catalyzing both delta 4- and delta 5-steroid-17,20-lyase reactions. *Mol. Endocrinol.* **3**, 968–975, <https://doi.org/10.1210/mend-3-6-968>
- 46 Oshima, H., Fan, D.F. and Troen, P. (1975) Studies of the human testis. V. Properties of delta-5-3beta and 17beta-hydroxysteroid dehydrogenases in the biosynthesis of testosterone from dehydroepiandrosterone. *J. Clin. Endocrinol. Metab.* **40**, 573–581, <https://doi.org/10.1210/jcem-40-4-573>
- 47 Lachance, Y., Luu-The, V., Labrie, C., Simard, J., Dumont, M., de Launoit, Y. et al. (1990) Characterization of human 3 beta-hydroxysteroid dehydrogenase/delta 5-delta 4-isomerase gene and its expression in mammalian cells. *J. Biol. Chem.* **265**, 20469–20475
- 48 Osawa, Y., Higashiyama, T., Shimizu, Y. and Yarborough, C. (1993) Multiple functions of aromatase and the active site structure; aromatase is the placental estrogen 2-hydroxylase. *J. Steroid Biochem. Mol. Biol.* **44**, 469–480, [https://doi.org/10.1016/0960-0760\(93\)90252-R](https://doi.org/10.1016/0960-0760(93)90252-R)
- 49 Jin, Y. and Penning, T.M. (2001) Steroid 5alpha-reductases and 3alpha-hydroxysteroid dehydrogenases: key enzymes in androgen metabolism. *Best Pract. Res. Clin. Endocrinol. Metab.* **15**, 79–94, <https://doi.org/10.1053/beem.2001.0120>
- 50 Riedemann, T., Patchev, A.V., Cho, K. and Almeida, O.F. (2010) Corticosteroids: way upstream. *Mol. Brain* **3**, 2, <https://doi.org/10.1186/1756-6606-3-2>
- 51 Joels, M. (2018) Corticosteroids and the brain. *J. Endocrinol.* **238**, R121–R130, <https://doi.org/10.1530/JOE-18-0226>
- 52 Furukawa, A., Miyatake, A., Ohnishi, T. and Ichikawa, Y. (1998) Steroidogenic acute regulatory protein (STAR) transcripts constitutively expressed in the adult rat central nervous system: colocalization of STAR, cytochrome P-450SCC (CYP XIA1), and 3beta-hydroxysteroid dehydrogenase in the rat brain. *J. Neurochem.* **71**, 2231–2238, <https://doi.org/10.1046/j.1471-4159.1998.71062231.x>
- 53 Inai, Y., Nagai, K., Ukena, K., Oishi, T. and Tsutsui, K. (2003) Seasonal changes in neurosteroid concentrations in the amphibian brain and environmental factors regulating their changes. *Brain Res.* **959**, 214–225, [https://doi.org/10.1016/S0006-8993\(02\)03745-9](https://doi.org/10.1016/S0006-8993(02)03745-9)
- 54 Mensah-Nyagan, A.G., Do-Rego, J.L., Beaujean, D., Luu-The, V., Pelletier, G. and Vaudry, H. (1999) Neurosteroids: expression of steroidogenic enzymes and regulation of steroid biosynthesis in the central nervous system. *Pharmacol. Rev.* **51**, 63–81
- 55 Ukena, K., Usui, M., Kohchi, C. and Tsutsui, K. (1998) Cytochrome P450 side-chain cleavage enzyme in the cerebellar Purkinje neuron and its neonatal change in rats. *Endocrinology* **139**, 137–147, <https://doi.org/10.1210/endo.139.1.5672>
- 56 Ukena, K., Kohchi, C. and Tsutsui, K. (1999) Expression and activity of 3beta-hydroxysteroid dehydrogenase/delta5-delta4-isomerase in the rat Purkinje neuron during neonatal life. *Endocrinology* **140**, 805–813, <https://doi.org/10.1210/endo.140.2.6516>
- 57 Le Goascogne, C., Robel, P., Gouezou, M., Sananes, N., Baulieu, E.E. and Waterman, M. (1987) Neurosteroids: cytochrome P-450scc in rat brain. *Science* **237**, 1212–1215, <https://doi.org/10.1126/science.3306919>
- 58 Leicaj, M.L., Pasquini, L.A., Lima, A., Gonzalez Deniselle, M.C., Pasquini, J.M., De Nicola, A.F. et al. (2018) Changes in neurosteroidogenesis during demyelination and remyelination in cuprizone-treated mice. *J. Neuroendocrinol.* **30**, e12649, <https://doi.org/10.1111/jne.12649>
- 59 Bonsack, F.T., Alleyne, Jr, C.H. and Sukumari-Ramesh, S. (2016) Augmented expression of TSPO after intracerebral hemorrhage: a role in inflammation? *J. Neuroinflammation* **13**, 151, <https://doi.org/10.1186/s12974-016-0619-2>
- 60 Rupprecht, R., Papadopoulos, V., Rammes, G., Baghai, T.C., Fan, J., Akula, N. et al. (2010) Translocator protein (18 kDa) (TSPO) as a therapeutic target for neurological and psychiatric disorders. *Nat. Rev. Drug Discov.* **9**, 971–988, <https://doi.org/10.1038/nrd3295>
- 61 Mellon, S.H. and Deschepper, C.F. (1993) Neurosteroid biosynthesis: genes for adrenal steroidogenic enzymes are expressed in the brain. *Brain Res.* **629**, 283–292, [https://doi.org/10.1016/0006-8993\(93\)91332-M](https://doi.org/10.1016/0006-8993(93)91332-M)

- 62 Hu, Z.Y., Bourreau, E., Jung-Testas, I., Robel, P. and Baulieu, E.E. (1987) Neurosteroids: oligodendrocyte mitochondria convert cholesterol to pregnenolone. *Proc. Natl. Acad. Sci. U.S.A.* **84**, 8215–8219, <https://doi.org/10.1073/pnas.84.23.8215>
- 63 Khanna, M., Qin, K.N. and Cheng, K.C. (1995) Distribution of 3 alpha-hydroxysteroid dehydrogenase in rat brain and molecular cloning of multiple cDNAs encoding structurally related proteins in humans. *J. Steroid Biochem. Mol. Biol.* **53**, 41–46, [https://doi.org/10.1016/0960-0760\(95\)00019-V](https://doi.org/10.1016/0960-0760(95)00019-V)
- 64 Agis-Balboa, R.C., Pinna, G., Pibiri, F., Kadriu, B., Costa, E. and Guidotti, A. (2007) Down-regulation of neurosteroid biosynthesis in corticolimbic circuits mediates social isolation-induced behavior in mice. *Proc. Natl. Acad. Sci. U.S.A.* **104**, 18736–18741, <https://doi.org/10.1073/pnas.0709419104>
- 65 Melcangi, R.C., Celotti, F., Castano, P. and Martini, L. (1993) Differential localization of the 5 alpha-reductase and the 3 alpha-hydroxysteroid dehydrogenase in neuronal and glial cultures. *Endocrinology* **132**, 1252–1259, <https://doi.org/10.1210/endo.132.3.8440186>
- 66 Melcangi, R.C., Celotti, F. and Martini, L. (1994) Progesterone 5-alpha-reduction in neuronal and in different types of glial cell cultures: type 1 and 2 astrocytes and oligodendrocytes. *Brain Res.* **639**, 202–206, [https://doi.org/10.1016/0006-8993\(94\)91731-0](https://doi.org/10.1016/0006-8993(94)91731-0)
- 67 Akwa, Y., Sananes, N., Guezou, M., Robel, P., Baulieu, E.E. and Le Goascogne, C. (1993) Astrocytes and neurosteroids: metabolism of pregnenolone and dehydroepiandrosterone. Regulation by cell density. *J. Cell Biol.* **121**, 135–143, <https://doi.org/10.1083/jcb.121.1.135>
- 68 Kabbadj, K., el-Etr, M., Baulieu, E.E. and Robel, P. (1993) Pregnenolone metabolism in rodent embryonic neurons and astrocytes. *Glia* **7**, 170–175, <https://doi.org/10.1002/glia.440070206>
- 69 Cascio, C., Brown, R.C., Liu, Y., Han, Z., Hales, D.B. and Papadopoulos, V. (2000) Pathways of dehydroepiandrosterone formation in rat brain glia. *J. Steroid Biochem. Mol. Biol.* **75**, 177–186, [https://doi.org/10.1016/S0960-0760\(00\)00163-1](https://doi.org/10.1016/S0960-0760(00)00163-1)
- 70 Gago, N., Akwa, Y., Sananes, N., Guennoun, R., Baulieu, E.E., El-Etr, M. et al. (2001) Progesterone and the oligodendroglial lineage: stage-dependent biosynthesis and metabolism. *Glia* **36**, 295–308, <https://doi.org/10.1002/glia.1117>
- 71 Koenig, H.L., Schumacher, M., Ferzaz, B., Thi, A.N., Ressouches, A., Guennoun, R. et al. (1995) Progesterone synthesis and myelin formation by Schwann cells. *Science* **268**, 1500–1503, <https://doi.org/10.1126/science.7770777>
- 72 Schumacher, M., Hussain, R., Gago, N., Oudinet, J.P., Mattern, C. and Ghomari, A.M. (2012) Progesterone synthesis in the nervous system: implications for myelination and myelin repair. *Front Neurosci.* **6**, 10, <https://doi.org/10.3389/fnins.2012.00010>
- 73 Usui, M., Yamazaki, T., Kominami, S. and Tsutsui, K. (1995) Avian neurosteroids. II. Localization of a cytochrome P450_{sc}-like substance in the quail brain. *Brain Res.* **678**, 10–20, [https://doi.org/10.1016/0006-8993\(95\)00117-9](https://doi.org/10.1016/0006-8993(95)00117-9)
- 74 Guennoun, R., Fiddes, R.J., Guezou, M., Lombes, M. and Baulieu, E.E. (1995) A key enzyme in the biosynthesis of neurosteroids, 3 beta-hydroxysteroid dehydrogenase/delta 5-delta 4-isomerase (3 beta-HSD), is expressed in rat brain. *Brain Res. Mol. Brain Res.* **30**, 287–300, [https://doi.org/10.1016/0169-328X\(95\)00016-L](https://doi.org/10.1016/0169-328X(95)00016-L)
- 75 Hirano, T. (2018) Purkinje Neurons: Development, Morphology, and Function. *Cerebellum* **17**, 699–700, <https://doi.org/10.1007/s12311-018-0985-7>
- 76 Jackman, S.L., Chen, C.H., Offermann, H.L., Drew, I.R., Harrison, B.M., Bowman, A.M. et al. (2020) Cerebellar Purkinje cell activity modulates aggressive behavior. *Elife* **9**, <https://doi.org/10.7554/eLife.53229>
- 77 Sakamoto, H., Ukena, K. and Tsutsui, K. (2001) Effects of progesterone synthesized de novo in the developing Purkinje cell on its dendritic growth and synaptogenesis. *J. Neurosci.* **21**, 6221–6232, <https://doi.org/10.1523/JNEUROSCI.21-16-06221.2001>
- 78 Sakamoto, H., Ukena, K. and Tsutsui, K. (2002) Dendritic spine formation in response to progesterone synthesized de novo in the developing Purkinje cell in rats. *Neurosci. Lett.* **322**, 111–115, [https://doi.org/10.1016/S0304-3940\(02\)00077-0](https://doi.org/10.1016/S0304-3940(02)00077-0)
- 79 Griffin, L.D., Gong, W., Verot, L. and Mellon, S.H. (2004) Niemann-Pick type C disease involves disrupted neurosteroidogenesis and responds to allopregnanolone. *Nat. Med.* **10**, 704–711, <https://doi.org/10.1038/nm1073>
- 80 Haraguchi, S., Hara, S., Ubuka, T., Mita, M. and Tsutsui, K. (2012) Possible role of pineal allopregnanolone in Purkinje cell survival. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 21110–21115, <https://doi.org/10.1073/pnas.1210804109>
- 81 Agis-Balboa, R.C., Pinna, G., Zhubi, A., Maloku, E., Veldic, M., Costa, E. et al. (2006) Characterization of brain neurons that express enzymes mediating neurosteroid biosynthesis. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 14602–14607, <https://doi.org/10.1073/pnas.0606544103>
- 82 Flood, J.F., Morley, J.E. and Roberts, E. (1992) Memory-enhancing effects in male mice of pregnenolone and steroids metabolically derived from it. *Proc. Natl. Acad. Sci. U.S.A.* **89**, 1567–1571, <https://doi.org/10.1073/pnas.89.5.1567>
- 83 Mayo, W., Dellu, F., Robel, P., Cherkaoui, J., Le Moal, M., Baulieu, E.E. et al. (1993) Infusion of neurosteroids into the nucleus basalis magnocellularis affects cognitive processes in the rat. *Brain Res.* **607**, 324–328, [https://doi.org/10.1016/0006-8993\(93\)91524-V](https://doi.org/10.1016/0006-8993(93)91524-V)
- 84 Macchi, M.M. and Bruce, J.N. (2004) Human pineal physiology and functional significance of melatonin. *Front. Neuroendocrinol.* **25**, 177–195, <https://doi.org/10.1016/j.yfrne.2004.08.001>
- 85 Hatori, M., Hirota, T., Iitsuka, M., Kurabayashi, N., Haraguchi, S., Kokame, K. et al. (2011) Light-dependent and circadian clock-regulated activation of sterol regulatory element-binding protein, X-box-binding protein 1, and heat shock factor pathways. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 4864–4869, <https://doi.org/10.1073/pnas.1015959108>
- 86 Tunc, A.T., Turgut, M., Aslan, H., Sahin, B., Yurtseven, M.E. and Kaplan, S. (2006) Neonatal pinealectomy induces Purkinje cell loss in the cerebellum of the chick: a stereological study. *Brain Res.* **1067**, 95–102, <https://doi.org/10.1016/j.brainres.2005.10.011>
- 87 Melcangi, R.C., Celotti, F., Castano, P. and Martini, L. (1992) Intracellular signalling systems controlling the 5 alpha-reductase in glial cell cultures. *Brain Res.* **585**, 411–415, [https://doi.org/10.1016/0006-8993\(92\)91247-C](https://doi.org/10.1016/0006-8993(92)91247-C)
- 88 Morita, K., Arimochi, H. and Tsuruo, Y. (2004) Adrenergic activation of steroid 5alpha-reductase gene expression in rat C6 glioma cells: involvement of cyclic amp/protein kinase A-mediated signaling pathway. *J. Mol. Neurosci.* **22**, 205–212, <https://doi.org/10.1385/JMN:22:3:205>
- 89 Gatliff, J. and Campanella, M. (2016) TSP0: kaleidoscopic 18-kDa amid biochemical pharmacology, control and targeting of mitochondria. *Biochem. J.* **473**, 107–121, <https://doi.org/10.1042/BJ20150899>
- 90 Whalin, M.E., Boujrad, N., Papadopoulos, V. and Krueger, K.E. (1994) Studies on the phosphorylation of the 18 kDa mitochondrial benzodiazepine receptor protein. *J. Recept. Res.* **14**, 217–228, <https://doi.org/10.3109/10799899409066033>

- 91 Micevych, P., Soma, K.K. and Sinchak, K. (2008) Neuroprogesterone: key to estrogen positive feedback? *Brain Res. Rev.* **57**, 470–480, <https://doi.org/10.1016/j.brainresrev.2007.06.009>
- 92 Chen, C., Kuo, J., Wong, A. and Micevych, P. (2014) Estradiol modulates translocator protein (TSPO) and steroid acute regulatory protein (STAR) via protein kinase A (PKA) signaling in hypothalamic astrocytes. *Endocrinology* **155**, 2976–2985, <https://doi.org/10.1210/en.2013-1844>
- 93 Dhandapani, K.M., Mahesh, V.B. and Brann, D.W. (2003) Astrocytes and brain function: implications for reproduction. *Exp. Biol. Med. (Maywood)* **228**, 253–260, <https://doi.org/10.1177/153537020322800303>
- 94 Aird, R.B. and Gordan, G.S. (1951) Anticonvulsive properties of desoxycorticosterone. *J. Am. Med. Assoc.* **145**, 715–719, <https://doi.org/10.1001/jama.1951.02920280027006>
- 95 Gyermek, L. (1967) Pregnanolone: a highly potent, naturally occurring hypnotic-anesthetic agent. *Proc. Soc. Exp. Biol. Med.* **125**, 1058–1062, <https://doi.org/10.3181/00379727-125-32276>
- 96 Green, C.J., Halsey, M.J., Precious, S. and Wardley-Smith, B. (1978) Alphaxalone-alphadolone anaesthesia in laboratory animals. *Lab. Anim.* **12**, 85–89, <https://doi.org/10.1258/002367778780953206>
- 97 Majewska, M.D., Harrison, N.L., Schwartz, R.D., Barker, J.L. and Paul, S.M. (1986) Steroid hormone metabolites are barbiturate-like modulators of the GABA receptor. *Science* **232**, 1004–1007, <https://doi.org/10.1126/science.2422758>
- 98 Ceccon, M., Rumbaugh, G. and Vicini, S. (2001) Distinct effect of pregnenolone sulfate on NMDA receptor subtypes. *Neuropharmacology* **40**, 491–500, [https://doi.org/10.1016/S0028-3908\(00\)00197-0](https://doi.org/10.1016/S0028-3908(00)00197-0)
- 99 Majewska, M.D. and Schwartz, R.D. (1987) Pregnenolone-sulfate: an endogenous antagonist of the gamma-aminobutyric acid receptor complex in brain? *Brain Res.* **404**, 355–360, [https://doi.org/10.1016/0006-8993\(87\)91394-1](https://doi.org/10.1016/0006-8993(87)91394-1)
- 100 Rebas, E., Radzik, T., Boczek, T. and Zylinska, L. (2017) Calcium-engaged Mechanisms of Nongenomic Action of Neurosteroids. *Curr. Neuropharmacol.* **15**, 1174–1191, <https://doi.org/10.2174/1570159X15666170329091935>
- 101 Sieghart, W. (2006) Structure, pharmacology, and function of GABAA receptor subtypes. *Adv. Pharmacol.* **54**, 231–263, [https://doi.org/10.1016/S1054-3589\(06\)54010-4](https://doi.org/10.1016/S1054-3589(06)54010-4)
- 102 Callachan, H., Cottrell, G.A., Hather, N.Y., Lambert, J.J., Nooney, J.M. and Peters, J.A. (1987) Modulation of the GABAA receptor by progesterone metabolites. *Proc. R. Soc. Lond. B. Biol. Sci.* **231**, 359–369
- 103 Lambert, J.J., Peters, J.A., Sturgess, N.C. and Hales, T.G. (1990) Steroid modulation of the GABAA receptor complex: electrophysiological studies. *Ciba Found. Symp.* **153**, 56–71
- 104 Twyman, R.E. and Macdonald, R.L. (1992) Neurosteroid regulation of GABAA receptor single-channel kinetic properties of mouse spinal cord neurons in culture. *J. Physiol.* **456**, 215–245, <https://doi.org/10.1113/jphysiol.1992.sp019334>
- 105 Zhu, W.J. and Vicini, S. (1997) Neurosteroid prolongs GABAA channel deactivation by altering kinetics of desensitized states. *J. Neurosci.* **17**, 4022–4031, <https://doi.org/10.1523/JNEUROSCI.17-11-04022.1997>
- 106 Seljeset, S., Lavery, D. and Smart, T.G. (2015) Inhibitory neurosteroids and the GABAA receptor. *Adv. Pharmacol.* **72**, 165–187, <https://doi.org/10.1016/bs.apha.2014.10.006>
- 107 Spivak, C.E. (1994) Desensitization and noncompetitive blockade of GABAA receptors in ventral midbrain neurons by a neurosteroid dehydroepiandrosterone sulfate. *Synapse* **16**, 113–122, <https://doi.org/10.1002/syn.890160205>
- 108 Lavery, D., Thomas, P., Field, M., Andersen, O.J., Gold, M.G., Biggin, P.C. et al. (2017) Crystal structures of a GABAA-receptor chimera reveal new endogenous neurosteroid-binding sites. *Nat. Struct. Mol. Biol.* **24**, 977–985, <https://doi.org/10.1038/nsmb.3477>
- 109 Alvarez, L.D. and Pecci, A. (2018) Structure and dynamics of neurosteroid binding to the alpha1beta2gamma2 GABAA receptor. *J. Steroid Biochem. Mol. Biol.* **182**, 72–80, <https://doi.org/10.1016/j.jsbmb.2018.04.012>
- 110 Hosie, A.M., Clarke, L., da Silva, H. and Smart, T.G. (2009) Conserved site for neurosteroid modulation of GABA A receptors. *Neuropharmacology* **56**, 149–154, <https://doi.org/10.1016/j.neuropharm.2008.07.050>
- 111 Carver, C.M. and Reddy, D.S. (2016) Neurosteroid Structure-Activity Relationships for Functional Activation of Extrasynaptic deltaGABA(A) Receptors. *J. Pharmacol. Exp. Ther.* **357**, 188–204, <https://doi.org/10.1124/jpet.115.229302>
- 112 Spigelman, I., Li, Z., Liang, J., Cagetti, E., Samzadeh, S., Mihalek, R.M. et al. (2003) Reduced inhibition and sensitivity to neurosteroids in hippocampus of mice lacking the GABA(A) receptor delta subunit. *J. Neurophysiol.* **90**, 903–910, <https://doi.org/10.1152/jn.01022.2002>
- 113 Stell, B.M., Brickley, S.G., Tang, C.Y., Farrant, M. and Mody, I. (2003) Neuroactive steroids reduce neuronal excitability by selectively enhancing tonic inhibition mediated by delta subunit-containing GABAA receptors. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 14439–14444, <https://doi.org/10.1073/pnas.2435457100>
- 114 Bianchi, M.T. and Macdonald, R.L. (2003) Neurosteroids shift partial agonist activation of GABA(A) receptor channels from low- to high-efficacy gating patterns. *J. Neurosci.* **23**, 10934–10943, <https://doi.org/10.1523/JNEUROSCI.23-34-10934.2003>
- 115 Seljeset, S., Bright, D.P., Thomas, P. and Smart, T.G. (2018) Probing GABAA receptors with inhibitory neurosteroids. *Neuropharmacology* **136**, 23–36, <https://doi.org/10.1016/j.neuropharm.2018.02.008>
- 116 Eisenman, L.N., He, Y., Fields, C., Zorumski, C.F. and Mennerick, S. (2003) Activation-dependent properties of pregnenolone sulfate inhibition of GABAA receptor-mediated current. *J. Physiol.* **550**, 679–691, <https://doi.org/10.1113/jphysiol.2003.043810>
- 117 Brown, N., Kerby, J., Bonnert, T.P., Whiting, P.J. and Wafford, K.A. (2002) Pharmacological characterization of a novel cell line expressing human alpha(4)beta(3)delta GABA(A) receptors. *Br. J. Pharmacol.* **136**, 965–974, <https://doi.org/10.1038/sj.bjp.0704795>
- 118 Zhu, W.J., Wang, J.F., Krueger, K.E. and Vicini, S. (1996) Delta subunit inhibits neurosteroid modulation of GABAA receptors. *J. Neurosci.* **16**, 6648–6656, <https://doi.org/10.1523/JNEUROSCI.16-21-06648.1996>
- 119 Vieira, M., Yong, X.L.H., Roche, K.W. and Anggono, V. (2020) Regulation of NMDA glutamate receptor functions by the GluN2 subunits. *J. Neurochem.* **154**, 121–143, <https://doi.org/10.1111/jnc.14970>

- 120 Malayev, A., Gibbs, T.T. and Farb, D.H. (2002) Inhibition of the NMDA response by pregnenolone sulphate reveals subtype selective modulation of NMDA receptors by sulphated steroids. *Br. J. Pharmacol.* **135**, 901–909, <https://doi.org/10.1038/sj.bjp.0704543>
- 121 Park-Chung, M., Wu, F.S. and Farb, D.H. (1994) 3 alpha-Hydroxy-5 beta-pregnan-20-one sulfate: a negative modulator of the NMDA-induced current in cultured neurons. *Mol. Pharmacol.* **46**, 146–150
- 122 Kussius, C.L., Kaur, N. and Popescu, G.K. (2009) Pregnanolone sulfate promotes desensitization of activated NMDA receptors. *J. Neurosci.* **29**, 6819–6827, <https://doi.org/10.1523/JNEUROSCI.0281-09.2009>
- 123 Bowlby, M.R. (1993) Pregnenolone sulfate potentiation of N-methyl-D-aspartate receptor channels in hippocampal neurons. *Mol. Pharmacol.* **43**, 813–819
- 124 Park-Chung, M., Wu, F.S., Purdy, R.H., Malayev, A.A., Gibbs, T.T. and Farb, D.H. (1997) Distinct sites for inverse modulation of N-methyl-D-aspartate receptors by sulfated steroids. *Mol. Pharmacol.* **52**, 1113–1123, <https://doi.org/10.1124/mol.52.6.1113>
- 125 Wu, F.S., Gibbs, T.T. and Farb, D.H. (1991) Pregnenolone sulfate: a positive allosteric modulator at the N-methyl-D-aspartate receptor. *Mol. Pharmacol.* **40**, 333–336
- 126 Horak, M., Vlcek, K., Chodounska, H. and Vyklicky, Jr, L. (2006) Subtype-dependence of N-methyl-D-aspartate receptor modulation by pregnenolone sulfate. *Neuroscience* **137**, 93–102, <https://doi.org/10.1016/j.neuroscience.2005.08.058>
- 127 Horak, M., Vlcek, K., Petrovic, M., Chodounska, H. and Vyklicky, Jr, L. (2004) Molecular mechanism of pregnenolone sulfate action at NR1/NR2B receptors. *J. Neurosci.* **24**, 10318–10325, <https://doi.org/10.1523/JNEUROSCI.2099-04.2004>
- 128 Adamusova, E., Cais, O., Vyklicky, V., Kudova, E., Chodounska, H., Horak, M. et al. (2013) Pregnenolone sulfate activates NMDA receptor channels. *Physiol. Res.* **62**, 731–736
- 129 Petrovic, M., Sedlacek, M., Horak, M., Chodounska, H. and Vyklicky, Jr, L. (2005) 20-oxo-5beta-pregnan-3alpha-yl sulfate is a use-dependent NMDA receptor inhibitor. *J. Neurosci.* **25**, 8439–8450, <https://doi.org/10.1523/JNEUROSCI.1407-05.2005>
- 130 Vyklicky, V., Krausova, B., Cerny, J., Balik, A., Zapotocky, M., Novotny, M. et al. (2015) Block of NMDA receptor channels by endogenous neurosteroids: implications for the agonist induced conformational states of the channel vestibule. *Sci. Rep.* **5**, 10935, <https://doi.org/10.1038/srep10935>
- 131 Jang, M.K., Mierke, D.F., Russek, S.J. and Farb, D.H. (2004) A steroid modulatory domain on NR2B controls N-methyl-D-aspartate receptor proton sensitivity. *Proc. Natl. Acad. Sci. U.S.A.* **101**, 8198–8203, <https://doi.org/10.1073/pnas.0401838101>
- 132 Cameron, K., Bartle, E., Roark, R., Fanelli, D., Pham, M., Pollard, B. et al. (2012) Neurosteroid binding to the amino terminal and glutamate binding domains of ionotropic glutamate receptors. *Steroids* **77**, 774–779, <https://doi.org/10.1016/j.steroids.2012.03.011>
- 133 Ikonen, E. (2008) Cellular cholesterol trafficking and compartmentalization. *Nat. Rev. Mol. Cell Biol.* **9**, 125–138, <https://doi.org/10.1038/nrm2336>
- 134 Wilson, D.P. and Patni, N. (2000) *Lysosomal Acid Lipase Deficiency* (Feingold, K.R., Anawalt, B., Boyce, A., Chrousos, G., Dungan, K., Grossman, A. et al., eds), Endotext, South Dartmouth (MA)
- 135 Lloyd-Evans, E. and Platt, F.M. (2010) Lipids on trial: the search for the offending metabolite in Niemann-Pick type C disease. *Traffic* **11**, 419–428, <https://doi.org/10.1111/j.1600-0854.2010.01032.x>
- 136 Meng, Y., Heybrock, S., Neculai, D. and Saftig, P. (2020) Cholesterol Handling in Lysosomes and Beyond. *Trends Cell Biol.* **30**, 452–466, <https://doi.org/10.1016/j.tcb.2020.02.007>
- 137 Ahmad, I., Lope-Piedrafita, S., Bi, X., Hicks, C., Yao, Y., Yu, C. et al. (2005) Allopregnanolone treatment, both as a single injection or repetitively, delays demyelination and enhances survival of Niemann-Pick C mice. *J. Neurosci. Res.* **82**, 811–821, <https://doi.org/10.1002/jnr.20685>
- 138 Liu, B., Li, H., Repa, J.J., Turley, S.D. and Dietschy, J.M. (2008) Genetic variations and treatments that affect the lifespan of the NPC1 mouse. *J. Lipid Res.* **49**, 663–669, <https://doi.org/10.1194/jlr.M700525-JLR200>
- 139 Davidson, C.D., Ali, N.F., Micsenyi, M.C., Stephney, G., Renault, S., Dobrenis, K. et al. (2009) Chronic cyclodextrin treatment of murine Niemann-Pick C disease ameliorates neuronal cholesterol and glycosphingolipid storage and disease progression. *PLoS ONE* **4**, e6951, <https://doi.org/10.1371/journal.pone.0006951>
- 140 Ory, D.S., Ottinger, E.A., Farhat, N.Y., King, K.A., Jiang, X., Weissfeld, L. et al. (2017) Intrathecal 2-hydroxypropyl-beta-cyclodextrin decreases neurological disease progression in Niemann-Pick disease, type C1: a non-randomised, open-label, phase 1-2 trial. *Lancet* **390**, 1758–1768, [https://doi.org/10.1016/S0140-6736\(17\)31465-4](https://doi.org/10.1016/S0140-6736(17)31465-4)
- 141 Walkley, S.U., Davidson, C.D., Jacoby, J., Marella, P.D., Ottinger, E.A., Austin, C.P. et al. (2016) Fostering collaborative research for rare genetic disease: the example of niemann-pick type C disease. *Orphanet. J. Rare Dis.* **11**, 161, <https://doi.org/10.1186/s13023-016-0540-x>
- 142 Chen, G., Li, H.M., Chen, Y.R., Gu, X.S. and Duan, S. (2007) Decreased estradiol release from astrocytes contributes to the neurodegeneration in a mouse model of Niemann-Pick disease type C. *Glia* **55**, 1509–1518, <https://doi.org/10.1002/glia.20563>
- 143 Miller, W.L. (2017) Disorders in the initial steps of steroid hormone synthesis. *J. Steroid Biochem. Mol. Biol.* **165**, 18–37, <https://doi.org/10.1016/j.jsmb.2016.03.009>
- 144 Roos, P., Svenstrup, K., Danielsen, E.R., Thomsen, C. and Nielsen, J.E. (2014) CYP7B1: novel mutations and magnetic resonance spectroscopy abnormalities in hereditary spastic paraplegia type 5A. *Acta Neurol. Scand.* **129**, 330–334, <https://doi.org/10.1111/ane.12188>
- 145 Holst, J.P., Soldin, O.P., Guo, T. and Soldin, S.J. (2004) Steroid hormones: relevance and measurement in the clinical laboratory. *Clin. Lab. Med.* **24**, 105–118, <https://doi.org/10.1016/j.cll.2004.01.004>
- 146 Wallin, M.T., Culpepper, W.J., Campbell, J.D., Nelson, L.M., Langer-Gould, A., Marrie, R.A. et al. (2019) The prevalence of MS in the United States: A population-based estimate using health claims data. *Neurology* **92**, e1029–e1040, <https://doi.org/10.1212/WNL.0000000000007035>
- 147 Noorbakhsh, F., Ellestad, K.K., Maingat, F., Warren, K.G., Han, M.H., Steinman, L. et al. (2011) Impaired neurosteroid synthesis in multiple sclerosis. *Brain* **134**, 2703–2721, <https://doi.org/10.1093/brain/awr200>
- 148 Confavreux, C., Hutchinson, M., Hours, M.M., Cortinovis-Tournaire, P. and Moreau, T. (1998) Rate of pregnancy-related relapse in multiple sclerosis. Pregnancy in Multiple Sclerosis Group. *N. Engl. J. Med.* **339**, 285–291, <https://doi.org/10.1056/NEJM199807303390501>

- 149 Chao, M.J., Ramagopalan, S.V., Herrera, B.M., Orton, S.M., Handunnetthi, L., Lincoln, M.R. et al. (2011) MHC transmission: insights into gender bias in MS susceptibility. *Neurology* **76**, 242–246, <https://doi.org/10.1212/WNL.0b013e318207b060>
- 150 Boghoozian, R., McKenzie, B.A., Saito, L.B., Mehta, N., Branton, W.G., Lu, J. et al. (2017) Suppressed oligodendrocyte steroidogenesis in multiple sclerosis: Implications for regulation of neuroinflammation. *Glia* **65**, 1590–1606, <https://doi.org/10.1002/glia.23179>
- 151 Garay, L., Gonzalez Deniselle, M.C., Lima, A., Roig, P. and De Nicola, A.F. (2007) Effects of progesterone in the spinal cord of a mouse model of multiple sclerosis. *J. Steroid Biochem. Mol. Biol.* **107**, 228–237, <https://doi.org/10.1016/j.jsbmb.2007.03.040>
- 152 Garay, L., Gonzalez Gigueaux, P., Guennoun, R., Schumacher, M., Gonzalez Deniselle, M.C. and De Nicola, A.F. (2017) Progesterone treatment modulates mRNA OF neurosteroidogenic enzymes in a murine model of multiple sclerosis. *J. Steroid Biochem. Mol. Biol.* **165**, 421–429, <https://doi.org/10.1016/j.jsbmb.2016.09.001>
- 153 Yates, M.A., Li, Y., Chlebeck, P., Proctor, T., Vandenberg, A.A. and Offner, H. (2010) Progesterone treatment reduces disease severity and increases IL-10 in experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* **220**, 136–139, <https://doi.org/10.1016/j.jneuroim.2010.01.013>
- 154 Lloyd-Evans, E. and Haslett, L.J. (2016) The lysosomal storage disease continuum with ageing-related neurodegenerative disease. *Ageing Res. Rev.* **32**, 104–121, <https://doi.org/10.1016/j.arr.2016.07.005>
- 155 Jones, L., Holmans, P.A., Hamshere, M.L., Harold, D., Moskvina, V., Ivanov, D. et al. (2010) Genetic evidence implicates the immune system and cholesterol metabolism in the aetiology of Alzheimer's disease. *PLoS ONE* **5**, e13950, <https://doi.org/10.1371/journal.pone.0013950>
- 156 Bernardi, F., Lanzone, A., Cento, R.M., Spada, R.S., Pezzani, I., Genazzani, A.D. et al. (2000) Allopregnanolone and dehydroepiandrosterone response to corticotropin-releasing factor in patients suffering from Alzheimer's disease and vascular dementia. *Eur. J. Endocrinol.* **142**, 466–471, <https://doi.org/10.1530/eje.0.1420466>
- 157 Smith, C.D., Wekstein, D.R., Markesbery, W.R. and Frye, C.A. (2006) 3alpha,5alpha-THP: a potential plasma neurosteroid biomarker in Alzheimer's disease and perhaps non-Alzheimer's dementia. *Psychopharmacology (Berl.)* **186**, 481–485, <https://doi.org/10.1007/s00213-005-0186-1>
- 158 Marx, C.E., Trost, W.T., Shampine, L.J., Stevens, R.D., Hulette, C.M., Steffens, D.C. et al. (2006) The neurosteroid allopregnanolone is reduced in prefrontal cortex in Alzheimer's disease. *Biol. Psychiatry* **60**, 1287–1294, <https://doi.org/10.1016/j.biopsych.2006.06.017>
- 159 Naylor, J.C., Kilts, J.D., Hulette, C.M., Steffens, D.C., Blazer, D.G., Ervin, J.F. et al. (2010) Allopregnanolone levels are reduced in temporal cortex in patients with Alzheimer's disease compared to cognitively intact control subjects. *Biochim. Biophys. Acta* **1801**, 951–959, <https://doi.org/10.1016/j.bbali.2010.05.006>
- 160 Naylor, J.C., Hulette, C.M., Steffens, D.C., Shampine, L.J., Ervin, J.F., Payne, V.M. et al. (2008) Cerebrospinal fluid dehydroepiandrosterone levels are correlated with brain dehydroepiandrosterone levels, elevated in Alzheimer's disease, and related to neuropathological disease stage. *J. Clin. Endocrinol. Metab.* **93**, 3173–3178, <https://doi.org/10.1210/jc.2007-1229>
- 161 Brown, R.C., Han, Z., Cascio, C. and Papadopoulos, V. (2003) Oxidative stress-mediated DHEA formation in Alzheimer's disease pathology. *Neurobiol. Aging* **24**, 57–65, [https://doi.org/10.1016/S0197-4580\(02\)00048-9](https://doi.org/10.1016/S0197-4580(02)00048-9)
- 162 Kim, S.B., Hill, M., Kwak, Y.T., Hampl, R., Jo, D.H. and Morfin, R. (2003) Neurosteroids: Cerebrospinal fluid levels for Alzheimer's disease and vascular dementia diagnostics. *J. Clin. Endocrinol. Metab.* **88**, 5199–5206, <https://doi.org/10.1210/jc.2003-030646>
- 163 Cardounel, A., Regelson, W. and Kalimi, M. (1999) Dehydroepiandrosterone protects hippocampal neurons against neurotoxin-induced cell death: mechanism of action. *Proc. Soc. Exp. Biol. Med.* **222**, 145–149, <https://doi.org/10.1046/j.1525-1373.1999.d01-124.x>
- 164 Gursoy, E., Cardounel, A. and Kalimi, M. (2001) Pregnenolone protects mouse hippocampal (HT-22) cells against glutamate and amyloid beta protein toxicity. *Neurochem. Res.* **26**, 15–21, <https://doi.org/10.1023/A:1007668213330>
- 165 Kato-Negishi, M. and Kawahara, M. (2008) Neurosteroids block the increase in intracellular calcium level induced by Alzheimer's beta-amyloid protein in long-term cultured rat hippocampal neurons. *Neuropsychiatr. Dis. Treat* **4**, 209–218, <https://doi.org/10.2147/ndt.s2059>
- 166 Maurice, T., Su, T.P. and Privat, A. (1998) Sigma1 (sigma 1) receptor agonists and neurosteroids attenuate B25-35-amyloid peptide-induced amnesia in mice through a common mechanism. *Neuroscience* **83**, 413–428, [https://doi.org/10.1016/S0306-4522\(97\)00405-3](https://doi.org/10.1016/S0306-4522(97)00405-3)
- 167 Migeon, C.J., Keller, A.R., Lawrence, B. and Shepard, II, T.H. (1957) Dehydroepiandrosterone and androsterone levels in human plasma: effect of age and sex; day-to-day and diurnal variations. *J. Clin. Endocrinol. Metab.* **17**, 1051–1062, <https://doi.org/10.1210/jcem-17-9-1051>
- 168 Harris, J.R. (2008) Cholesterol binding to amyloid-beta fibrils: a TEM study. *Micron* **39**, 1192–1196, <https://doi.org/10.1016/j.micron.2008.05.001>
- 169 Reddy, D.S. and Estes, W.A. (2016) Clinical Potential of Neurosteroids for CNS Disorders. *Trends Pharmacol. Sci.* **37**, 543–561, <https://doi.org/10.1016/j.tips.2016.04.003>
- 170 Kharasch, E.D. and Hollmann, M.W. (2015) Steroid Anesthesia Revisited: Again. *Anesth. Analg.* **120**, 983–984, <https://doi.org/10.1213/ANE.0000000000000667>
- 171 Nohria, V. and Giller, E. (2007) Ganaxolone. *Neurotherapeutics* **4**, 102–105, <https://doi.org/10.1016/j.nurt.2006.11.003>
- 172 Fung, L.K., Libove, R.A., Phillips, J., Haddad, F. and Hardan, A.Y. (2014) Brief report: an open-label study of the neurosteroid pregnenolone in adults with autism spectrum disorder. *J. Autism Dev. Disord.* **44**, 2971–2977, <https://doi.org/10.1007/s10803-014-2144-4>
- 173 Braat, S. and Kooy, R.F. (2015) Insights into GABAergic system deficits in fragile X syndrome lead to clinical trials. *Neuropharmacology* **88**, 48–54, <https://doi.org/10.1016/j.neuropharm.2014.06.028>
- 174 Ligsay, A., Van Dijk, A., Nguyen, D.V., Lozano, R., Chen, Y., Bickel, E.S. et al. (2017) A randomized double-blind, placebo-controlled trial of ganaxolone in children and adolescents with fragile X syndrome. *J. Neurodev. Disord.* **9**, 26, <https://doi.org/10.1186/s11689-017-9207-8>
- 175 Marx, C.E., Shampine, L.J., Duncan, G.E., VanDoren, M.J., Grobin, A.C., Massing, M.W. et al. (2006) Clozapine markedly elevates pregnenolone in rat hippocampus, cerebral cortex, and serum: candidate mechanism for superior efficacy? *Pharmacol. Biochem. Behav.* **84**, 598–608, <https://doi.org/10.1016/j.pbb.2006.07.026>
- 176 Aldred, S. and Mecocci, P. (2010) Decreased dehydroepiandrosterone (DHEA) and dehydroepiandrosterone sulfate (DHEAS) concentrations in plasma of Alzheimer's disease (AD) patients. *Arch. Gerontol. Geriatr.* **51**, e16–e18, <https://doi.org/10.1016/j.archger.2009.07.001>

- 177 Steckelbroeck, S., Jin, Y., Gopishetty, S., Oyesanmi, B. and Penning, T.M. (2004) Human cytosolic 3 α -hydroxysteroid dehydrogenases of the aldo-keto reductase superfamily display significant 3 β -hydroxysteroid dehydrogenase activity: implications for steroid hormone metabolism and action. *J. Biol. Chem.* **279**, 10784–10795, <https://doi.org/10.1074/jbc.M313308200>
- 178 Dacarett-Galeano, D.J. and Diao, X.Y. (2019) Brexanolone: A Novel Therapeutic in the Treatment of Postpartum Depression. *Am. J. Psychiatry* **15**, 2–4, <https://doi.org/10.1176/appi.ajp-rj.2019.150201>