

Research Article

Methylenetetrahydrofolate reductase C677T (Ala > Val, rs1801133 C > T) polymorphism decreases the susceptibility of hepatocellular carcinoma: a meta-analysis involving 12,628 subjects

Sheng Zhang^{1,*}, Jiakai Jiang^{1,*},  Weifeng Tang² and Longgen Liu³¹Department of General Surgery, Changzhou No. 3 People's Hospital, Changzhou, Jiangsu Province, China; ²Department of Cardiothoracic Surgery, Affiliated People's Hospital of Jiangsu University, Zhenjiang, Jiangsu Province, China; ³Department of Liver Disease, Changzhou No. 3 People's Hospital, Changzhou, Jiangsu Province, China**Correspondence:** Sheng Zhang (13601507172@163.com) or Longgen Liu (jsllg0519@163.com)

C677T (Ala > Val, rs1801133 C > T), a non-synonymous variant of *methylenetetrahydrofolate reductase* (*MTHFR*) gene, has been found to be associated with an impair enzyme activity of *MTHFR*. The relationship of *MTHFR* rs1801133 with hepatocellular carcinoma (HCC) has been extensively investigated. However, the findings were conflicting. Recently, more investigations have been conducted on the relationship of *MTHFR* rs1801133 with HCC. To obtain a more precise assessment on the effect of this non-synonymous variant to the development of HCC, a pooled-analysis was performed. This meta-analysis consisted of 19 independent case-control studies. By using the odds ratio (OR) combined with 95% confidence interval (CI), the relationship of *MTHFR* rs1801133 with HCC risk was determined. A total of 19 independent case-control studies were included. Finally, 6,102 HCC cases and 6,526 controls were recruited to examine the relationship of *MTHFR* rs1801133 with HCC risk. In recessive model (TT vs. CC/CT), the findings reached statistical significance (OR, 0.90; 95%CI, 0.82–0.98; $P = 0.016$). Subgroup analysis also found an association between *MTHFR* rs1801133 polymorphism and the decreased risk of HCC in hepatitis/virus related patients (recessive model: OR, 0.85; 95%CI, 0.72–0.99; $P = 0.035$, and allele model: OR, 0.90; 95%CI, 0.81–0.99; $P = 0.028$). Subgroup analyses indicated that extreme heterogeneity existed in Asian population, larger sample size investigation, hospital-based study and normal/healthy control subgroups. The shape of Begger's seemed symmetrical. Egger's linear regression test also confirmed these evaluations. Sensitivity analyses suggested that our findings were stable. In summary, our results highlight that *MTHFR* rs1801133 polymorphism decreases HCC susceptibility. The relationship warrants a further assessment.

*These authors have contributed equally to this work.

Received: 13 December 2019
Revised: 03 January 2020
Accepted: 13 January 2020Accepted Manuscript online:
03 February 2020
Version of Record published:
20 February 2020

Introduction

In 2018, global cancer statistics estimated that liver malignancy was the fifth most frequent type of cancer incidence among men and the eleven most frequent type among women, about 596,574 and 244,506 new cases diagnosed worldwide, respectively [1]. However, the fatality was the third most frequent type [1]. The etiology of liver cancer (LC) was not well-established. Hepatocellular carcinoma (HCC) is one of the most important primary LC, which comprised almost 80% of LC cases. Some major susceptibility factors (e.g. aflatoxin-contaminated food, superabundant drinking, tobacco consumption, chronic virus infection, higher body mass index and Type 2 diabetes) [2–6] may contribute to the development of HCC. Additionally, hereditary factor has also been suggested to affect the susceptibility for the occurrence of HCC.

Methylenetetrahydrofolate reductase (MTHFR) locates in 1p36.3, which maps from 11785723 to 11806103 (GRCh38; April, 2018). *MTHFR*, a key enzyme, plays a vital effect in folate metabolism by the role of catalyzing the 5,10-methylenetetrahydrofolate (5,10-methylene-THF) to 5-methyltetrahydrofolate (5-methylene-THF) irreversibly. In the conversion of homocysteine to methionine, 5-methylene-THF is a primary methyl donor [7]. *MTHFR* rs1801133 (C677T), a non-synonymous variant (Ala>Val), has been suggested to influence the activity of *MTHFR* enzyme [8]. The correlation of *MTHFR* rs1801133 polymorphism with malignancy has been extensively explored. This single-nucleotide polymorphism (SNP) was suggested to be associated with thyroid cancer [9], colorectal cancer [10,11], breast cancer [12], esophagogastric junction adenocarcinoma [13], non-small cell lung cancer [14], acute lymphoblastic leukemia [15,16], gastric cancer [17], renal cell carcinoma [18] and esophageal carcinoma [19], among others.

Recently, many case–control studies have been carried out to determine the relationship of *MTHFR* rs1801133 polymorphism with the development of HCC [19–29]. However, the observations were controversial. Several meta-analyses also got conflicting results. To shed light on this issue, we conducted an extensive pooled-analysis to determine the role of *MTHFR* rs1801133 polymorphism on the development of HCC.

Materials and methods

Study searching

Publications were obtained by searching the PubMed and EMBASE databases before October 19, 2019. The following strategy was used: (Methylenetetrahydrofolate reductase OR *MTHFR* OR rs1801133) AND (SNP OR polymorphism) AND (cancer OR carcinoma) and (hepatocellular OR liver). The references in reviews and meta-analyses were also retrieved to get data. In this pooled-analysis, there was no language limited.

Inclusion criteria

In our meta-analysis, the eligible criteria of the included publications were: (1) designed as a case–control study; (2) focusing on the relationship of the *MTHFR* rs1801133 polymorphism with HCC risk; (3) genotype data could be extracted and (4) publications were compatible with Hardy–Weinberg equilibrium (HWE) in controls.

Exclusion criteria

The criteria for exclusion were as following: (1) publications incompatible with HWE; (2) overlapping data; (3) not case–control study design and (4) only focusing on the relationship of *MTHFR* rs1801133 polymorphism with HCC survival.

Data extraction

The authors (S. Zhang and J. Jiang) extracted the following data: the surname of first author, publication year, populations studied, country where the investigation was carried out, ratio of sex, age, drinking, positive (%) of hepatitis B surface antigen (HBsAg), genotyping method, the number of participants and *MTHFR* rs1801133 genotype. If there was conflicting assessment, another reviewer (W. Tang) was invited. During this process, they made a vote to obtain the final decision.

Statistical methods

In the present study, the odds ratios (ORs) combined with 95% confidence intervals (CIs) were harnessed to compare the difference between HCC group and controls. *P* value (<0.05) was considered statistically significant. The present meta-analysis determined the correlation in four genetic models [e.g. dominant model (TT/CT vs. CC), homozygote model (TT vs. CC), allele model (T vs. C) and recessive model (TT vs. CC/CT)]. Using *I*² metric and *Q* statistic, the heterogeneity among the eligible case–control studies was evaluated. If *P* < 0.10 or *I*² > 50%, we defined that there was significant heterogeneity. Thus, the random-effect model was used [30,31]. Otherwise, there was no heterogeneity detected. A fixed-effect model was used to combine the data [32]. The Egger test and Begg's test were used to assess the bias of publication. If *P* < 0.10, we defined that there was a significant publication bias. By omitting a study one by one and analyzing the remainders, sensitivity analysis was performed to assess the stability of our findings. The distribution of the *MTHFR* rs1801133 genotype was used to calculate the *P* value of HWE by using an online software (<http://ihg.gsf.de/cgi-bin/hw/hwa1.pl>) in controls [33–35]. STATA 12.0 software (Stata Corp., College Station, Texas) was used to conduct the analysis. In this study, *P* value was two sided.

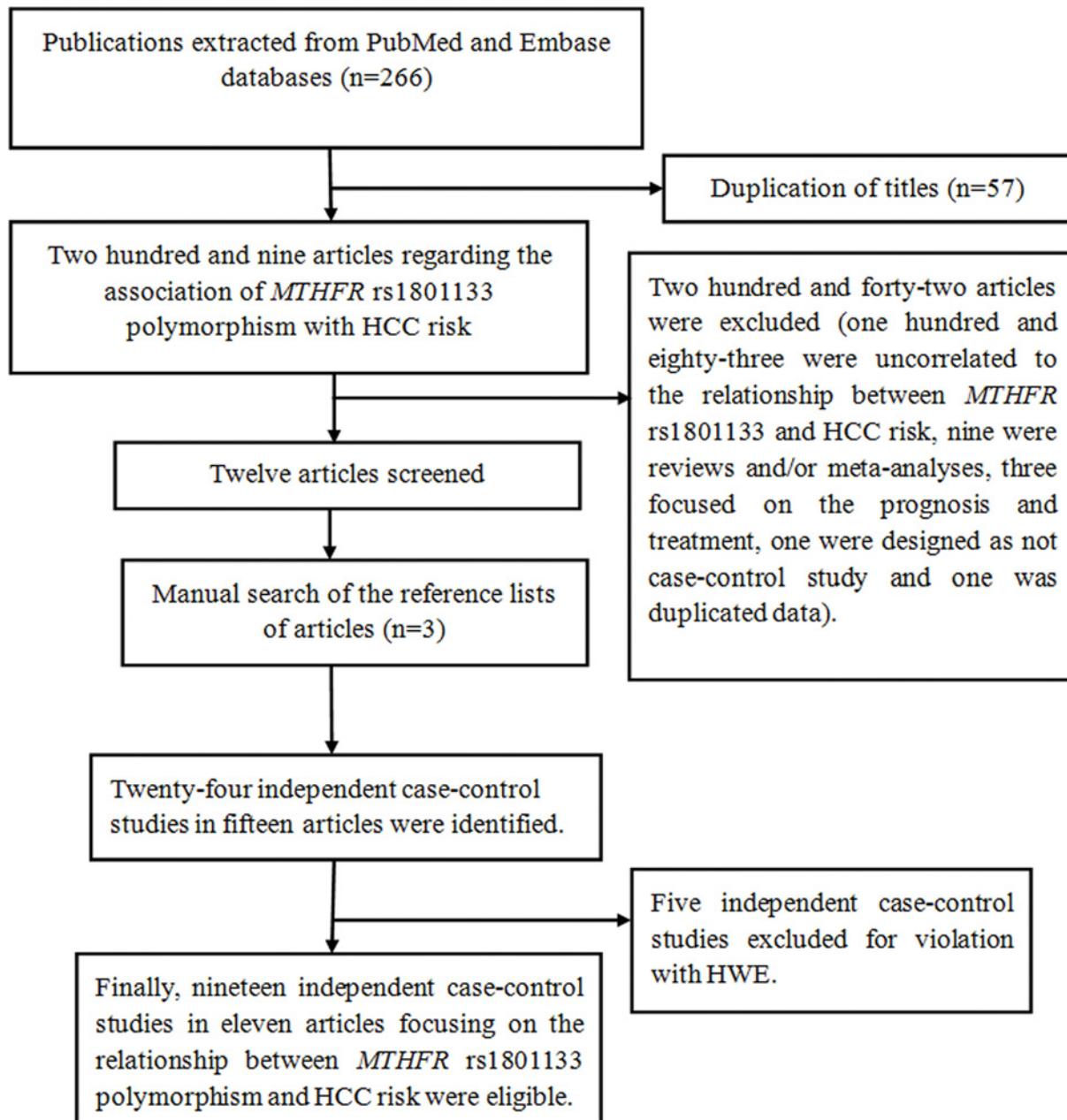


Figure 1. Flow diagram of this meta-analysis

Quality assessment of meta-analysis

Two authors (S.Z. and J.J.) independently extracted the data and calculated the quality score of the included case-control studies. The detailed scores were determined by a quality assessment criteria, which were presented in previous studies [36,37]. If the scores were more than 6.0, the investigation had an acceptable quality [38].

Results

Eligible studies

A total of 11 publications were eligible (Figure 1). Four articles involved several different subgroups, so we considered them as independent investigations. After a screening, 19 independent case-control studies were included. In addition, five publications were excluded for incompatible with HWE [29,39,40–42]. Finally, 6,102 HCC cases and 6,526 controls were recruited to examine the relationship of *MTHFR* rs1801133 polymorphism with HCC risk (Table

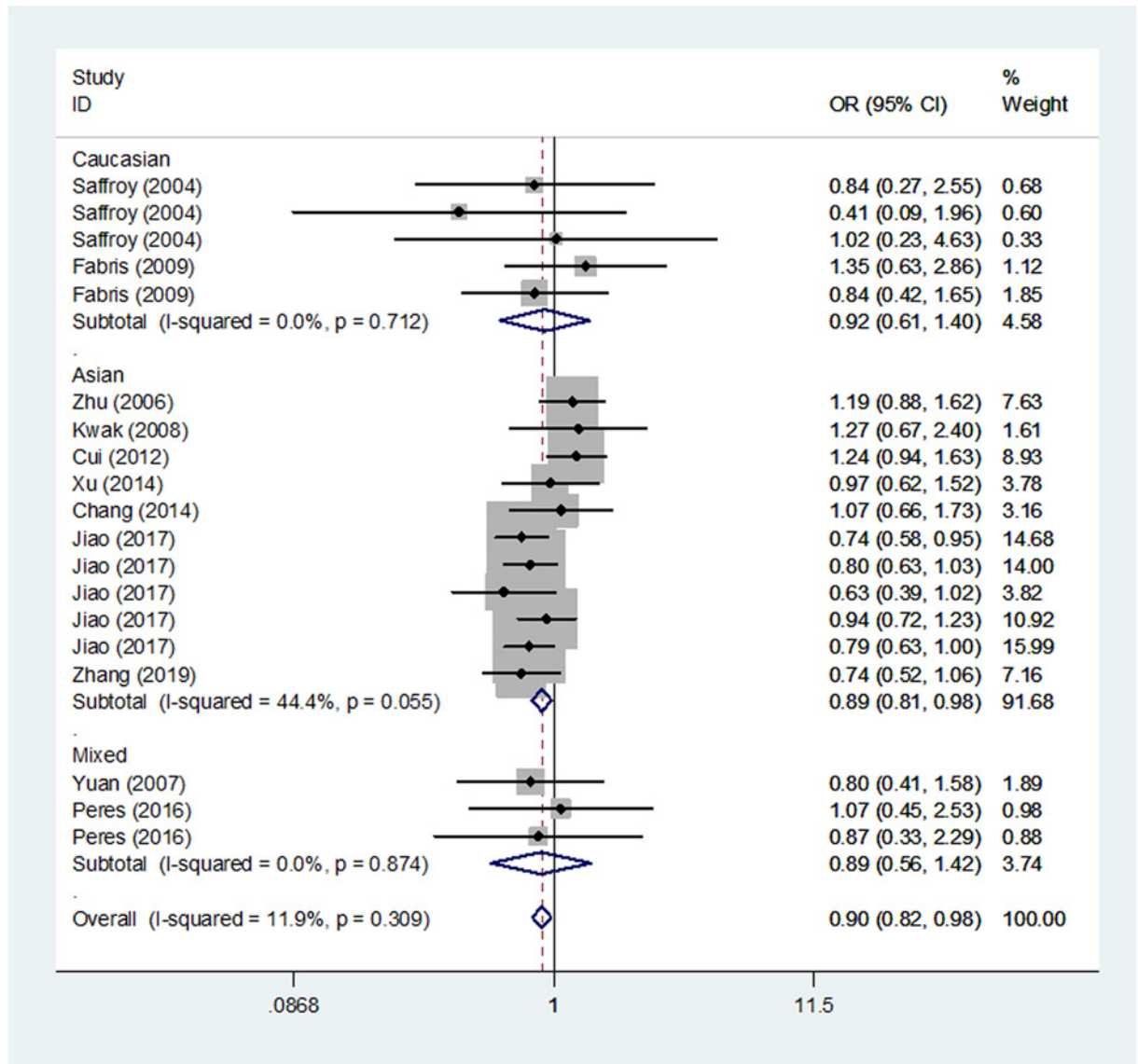


Figure 2. Meta-analysis of the association between *MTHFR* rs1801133 polymorphism and HCC risk (recessive model, fixed-effects model)

1). The publication year covered from 2004 to 2019. These investigations were performed in different populations: three were conducted in mixed populations [28,29], four were carried out in Caucasians [26,27], and twelve involved Asians [19–25]. The *MTHFR* rs1801133 genotypes are summarized in Table 2.

Meta-analysis results

In the eligible investigations, the MAF of *MTHFR* rs1801133 C/T polymorphism was 0.475 in HCC patients (5,670/11,944) and was 0.466 in controls (5,721/12,288). In different race, the MAF of controls was not similar in controls. The MAFs were 0.352 (480/1,362) in mixed populations, 0.328 (214/652) in Caucasians, and that was 0.485 (5,075/10,462) in Asians.

The pooled-analysis findings were reported in four genetic models including 19 independent case-control studies. In recessive model (TT vs. CC/CT), the findings reached statistical significance (OR, 0.90; 95%CI, 0.82–0.98; $P = 0.016$, Table 3 and Figure 2). In other genetic models, we failed to obtain the significance (dominant model: OR, 0.92; 95%CI, 0.81–1.05; $P = 0.209$, homozygote model: OR, 0.88; 95%CI, 0.77–1.01; $P = 0.078$, and allele model: OR, 0.93; 95%CI, 0.85–1.01; $P = 0.077$, Table 3).

Table 1 Characteristics of the studies in meta-analysis

| Study | Year | Sample size | Country | Ethnicity | Sex, male (%) Case/Control | Age (years); Case/Control | Drinking (%) Case/Control | HBsAg, positive (%) Case/Control | Genotype method | Source of control | Type of control |
|---------|------|-------------|---------|-----------|-------------------------------|------------------------------|------------------------------|-------------------------------------|-----------------|-------------------|------------------------------------|
| Cui | 2012 | 356/641 | China | Asian | 83.1/43.5 | 56.6/58.7 | 44.1/30.3 | 77.5/8.6 | Real-time PCR | PB | Normal or healthy control |
| Fabris | 2009 | 65/147 | Italy | Caucasian | NA/NA | NA/NA | NA | 26.2/10.9 | PCR-RFLP | HB | Hepatitis or virus related control |
| Fabris | 2009 | 65/236 | Italy | Caucasian | NA/69.5 | NA/48 | NA | NA/NA | PCR-RFLP | HB | NA |
| Jiao | 2017 | 726/549 | China | Asian | 72.7/54.6 | 56.5/41.5 | 24.5/NA | 89.1/0.0 | TaqMan | HB | Normal or healthy control |
| Jiao | 2017 | 726/558 | China | Asian | 72.7/53.6 | 56.5/33.7 | 24.5/NA | 89.1/0.0 | TaqMan | HB | Normal or healthy control |
| Jiao | 2017 | 726/81 | China | Asian | 72.7/61.7 | 56.5/34.4 | 24.5/NA | 89.1/100 | TaqMan | HB | Hepatitis or virus related control |
| Jiao | 2017 | 726/442 | China | Asian | 72.7/66.5 | 56.5/39.6 | 24.5/13.6 | 89.1/100 | TaqMan | HB | Hepatitis or virus related control |
| Jiao | 2017 | 726/704 | China | Asian | 72.7/64.9 | 56.5/53.7 | 24.5/23.6 | 89.1/88.7 | TaqMan | HB | Hepatitis or virus related control |
| Kwak | 2008 | 96/201 | Korea | Asian | NA | 57.6/53.6 | NA | NA | PCR-RFLP | HB | Normal or healthy control |
| Peres | 2016 | 71/356 | Brazil | Mixed | 73.2/73.3 | NA | 62.0/46.0 | NA | PCR-RFLP | HB | Normal or healthy control |
| Peres | 2016 | 71/116 | Brazil | Mixed | 73.2/74.1 | NA | 62.0/53.4 | NA | PCR-RFLP | HB | Hepatitis or virus related control |
| Saffroy | 2004 | 72/122 | France | Caucasian | 84.7/85.2 | 55/50 | NA | NA | PCR-RFLP | HB | Hepatitis or virus related control |
| Saffroy | 2004 | 27/80 | France | Caucasian | 74.1/86.3 | 54/54 | NA | NA | PCR-RFLP | HB | Normal or healthy control |
| Saffroy | 2004 | 49/30 | France | Caucasian | 85.7/66.7 | 56/52 | NA | NA | PCR-RFLP | HB | Hepatitis or virus related control |
| Xu | 2014 | 205/200 | China | Asian | NA | 52.0/61.0 | NA | NA | PCR | NA | NA |
| Yuan | 2007 | 118/209 | USA | Mixed | 68.6/61.2 | NA | 71.2/68.4 | 28.0/11.5 | TaqMan | PB | Normal or healthy control |
| Zhu | 2006 | 508/543 | China | Asian | 85.8/48.8 | 50/45 | 39.8/17.9 | 72.8/17.9 | PCR-RFLP | HB | Normal or healthy control |
| Chang | 2014 | 204/415 | China | Asian | 77.9/69.2 | 53.9/57.7 | 41.7/35.7 | 64.7/24.6 | PCR-RFLP | PB | Normal or healthy control |
| Zhang | 2019 | 584/923 | China | Asian | 89.9/90.5 | 53.2/53.7 | 29.1/16.0 | 70.6/9.2 | SNPscan | HB | Normal or healthy control |

PCR-RFLP: polymerase chain reaction-restriction fragment length polymorphism
 PCR: polymerase chain reaction
 SNP: single-nucleotide polymorphism
 HP: hospital-based
 PB: population-based
 NA: not available

Table 2 Distribution of *MTHFR* rs1801133 C>T polymorphism genotype and allele

| Study | Year | Case TT | Case CT | Case CC | Control TT | Control TC | Control CC | Case T | Case C | Control T | Control C | HWE | Quality assessment |
|---------|------|---------|---------|------------|------------|------------|-------------|--------|--------|-----------|-----------|-----|--------------------|
| Cui | 2012 | 125 | 179 | 52 | 195 | 325 | 121 | 429 | 283 | 715 | 567 | Yes | 7.0 |
| Fabris | 2009 | 13 | – | CC/CT = 52 | 23 | – | CC/CT = 124 | – | – | – | – | Yes | 6.5 |
| Fabris | 2009 | 13 | – | CC/CT = 52 | 54 | 113 | 69 | – | – | – | – | Yes | 6.5 |
| Jiao | 2017 | 188 | 370 | 168 | 176 | 263 | 110 | 746 | 706 | 615 | 483 | Yes | 7.5 |
| Jiao | 2017 | 188 | 370 | 168 | 169 | 268 | 121 | 746 | 706 | 606 | 510 | Yes | 7.5 |
| Jiao | 2017 | 188 | 370 | 168 | 29 | 35 | 17 | 746 | 706 | 93 | 69 | Yes | 6.5 |
| Jiao | 2017 | 188 | 370 | 168 | 120 | 222 | 100 | 746 | 706 | 462 | 422 | Yes | 7.5 |
| Jiao | 2017 | 188 | 370 | 168 | 215 | 338 | 151 | 746 | 706 | 768 | 640 | Yes | 7.5 |
| Kwak | 2008 | 18 | 46 | 32 | 31 | 106 | 64 | 82 | 110 | 168 | 234 | Yes | 6.5 |
| Peres | 2016 | 7 | 36 | 28 | 33 | 174 | 149 | 50 | 92 | 240 | 472 | Yes | 7.0 |
| Peres | 2016 | 7 | 36 | 28 | 13 | 55 | 48 | 50 | 92 | 81 | 151 | Yes | 6.0 |
| Saffroy | 2004 | 5 | 24 | 43 | 10 | 60 | 52 | 34 | 110 | 80 | 164 | Yes | 6.5 |
| Saffroy | 2004 | 2 | 16 | 9 | 13 | 37 | 30 | 20 | 34 | 63 | 97 | Yes | 6.5 |
| Saffroy | 2004 | 5 | 29 | 15 | 3 | 17 | 10 | 39 | 59 | 23 | 37 | Yes | 6.5 |
| Xu | 2014 | 50 | 112 | 43 | 50 | 111 | 39 | 212 | 198 | 211 | 189 | Yes | 6.5 |
| Yuan | 2007 | 14 | 51 | 53 | 30 | 99 | 80 | 79 | 157 | 159 | 259 | Yes | 7.0 |
| Zhu | 2006 | 110 | 226 | 172 | 102 | 268 | 173 | 446 | 570 | 472 | 614 | Yes | 8.0 |
| Chang | 2014 | 30 | 114 | 50 | 57 | 199 | 135 | 174 | 214 | 313 | 469 | Yes | 7.5 |
| Zhang | 2019 | 49 | 227 | 299 | 103 | 446 | 372 | 325 | 825 | 652 | 1190 | Yes | 8.0 |

HWE: Hardy–Weinberg equilibrium.

Table 3 Results of the meta-analysis from different comparative genetic models

| | No. of studies | T vs. C | | | | TT vs. CC | | | | TT/CT vs. CC | | | | TT vs. CT/CC | | | |
|----------------------------|----------------|------------------------|------------------|----------------|-----------|------------------------|--------------|----------------|-----------|------------------------|------------------|----------------|-----------|------------------------|--------------|----------------|-----------|
| | | OR (95% CI) | P | I ² | P(Q-test) | OR(95% CI) | P | I ² | P(Q-test) | OR(95% CI) | P | I ² | P(Q-test) | OR(95% CI) | P | I ² | P(Q-test) |
| Total | 19 | 0.93(0.85–1.01) | 0.077 | 48.7% | 0.013 | 0.88(0.77–1.01) | 0.078 | 25.8% | 0.158 | 0.92(0.81–1.05) | 0.209 | 47.3% | 0.016 | 0.90(0.82–0.98) | 0.016 | 11.9% | 0.309 |
| Ethnicity | | | | | | | | | | | | | | | | | |
| Asians | 11 | 0.94(0.85–1.03) | 0.182 | 63.4% | 0.002 | 0.90(0.75–1.06) | 0.210 | 50.1% | 0.029 | 0.94(0.80–1.09) | 0.380 | 59.3% | 0.006 | 0.90(0.79–1.03) | 0.133 | 44.4% | 0.055 |
| Caucasians | 5 | 0.79(0.57–1.09) | 0.153 | 0.0% | 0.405 | 0.67(0.30–1.50) | 0.331 | 0.0% | 0.781 | 0.73(0.47–1.13) | 0.155 | 42.2% | 0.177 | 0.92(0.61–1.40) | 0.693 | 0.0% | 0.712 |
| Mixed | 3 | 0.94(0.76–1.17) | 0.592 | 0.0% | 0.549 | 0.86(0.52–1.41) | 0.550 | 0.0% | 0.721 | 0.94(0.70–1.27) | 0.679 | 0.0% | 0.494 | 0.89(0.56–1.42) | 0.619 | 0.0% | 0.874 |
| Sample sizes | | | | | | | | | | | | | | | | | |
| <1000 | 13 | 1.02(0.93–1.13) | 0.631 | 25.8% | 0.198 | 1.06(0.86–1.31) | 0.559 | 0.0% | 0.488 | 1.05(0.90–1.22) | 0.537 | 31.3% | 0.149 | 1.02(0.87–1.19) | 0.857 | 0.0% | 0.686 |
| ≥1000 | 6 | 0.88(0.80–0.96) | 0.006 | 52.6% | 0.061 | 0.80(0.70–0.92) | 0.001 | 29.1% | 0.217 | 0.84(0.73–0.97) | 0.020 | 48.8% | 0.082 | 0.85(0.76–0.94) | 0.002 | 35.3% | 0.172 |
| Source of control | | | | | | | | | | | | | | | | | |
| P-B | 3 | 1.10(0.89–1.36) | 0.374 | 53.7% | 0.116 | 1.30(0.97–1.73) | 0.080 | 39.9% | 0.190 | 1.19(0.81–1.75) | 0.385 | 65.0% | 0.057 | 1.14(0.91–1.43) | 0.248 | 0.0% | 0.490 |
| H-B | 15 | 0.88(0.83–0.93) | <0.001 | 22.9% | 0.212 | 0.81(0.71–0.92) | 0.001 | 0.0% | 0.642 | 0.84(0.77–0.93) | <0.001 | 23.2% | 0.209 | 0.85(0.77–0.94) | 0.002 | 0.0% | 0.498 |
| NA | 1 | 0.96(0.73–1.26) | 0.766 | – | – | 0.91(0.51–1.63) | 0.743 | – | – | 0.91(0.56–1.48) | 0.712 | – | – | 0.97(0.62–1.52) | 0.887 | – | – |
| Control type | | | | | | | | | | | | | | | | | |
| Normal or healthy | 10 | 0.95(0.84–1.08) | 0.428 | 66.3% | 0.002 | 0.92(0.74–1.16) | 0.487 | 53.9% | 0.021 | 0.95(0.79–1.15) | 0.590 | 65.0% | 0.002 | 0.94(0.79–1.11) | 0.439 | 42.3% | 0.076 |
| Hepatitis or virus related | 7 | 0.90(0.81–0.99) | 0.028 | 0.0% | 0.533 | 0.82(0.67–1.00) | 0.054 | 0.0% | 0.905 | 0.90(0.56–1.06) | 0.208 | 0.0% | 0.463 | 0.85(0.72–0.99) | 0.035 | 0.0% | 0.695 |
| NA | 2 | 0.96(0.73–01.26) | 0.766 | – | – | 0.91(0.51–1.63) | 0.743 | – | – | 0.91(0.56–1.48) | 0.712 | – | – | 0.93(0.64–1.35) | 0.684 | 0.0% | 0.729 |

P-B: population-based;
 H-B: hospital-based
 NA: not available

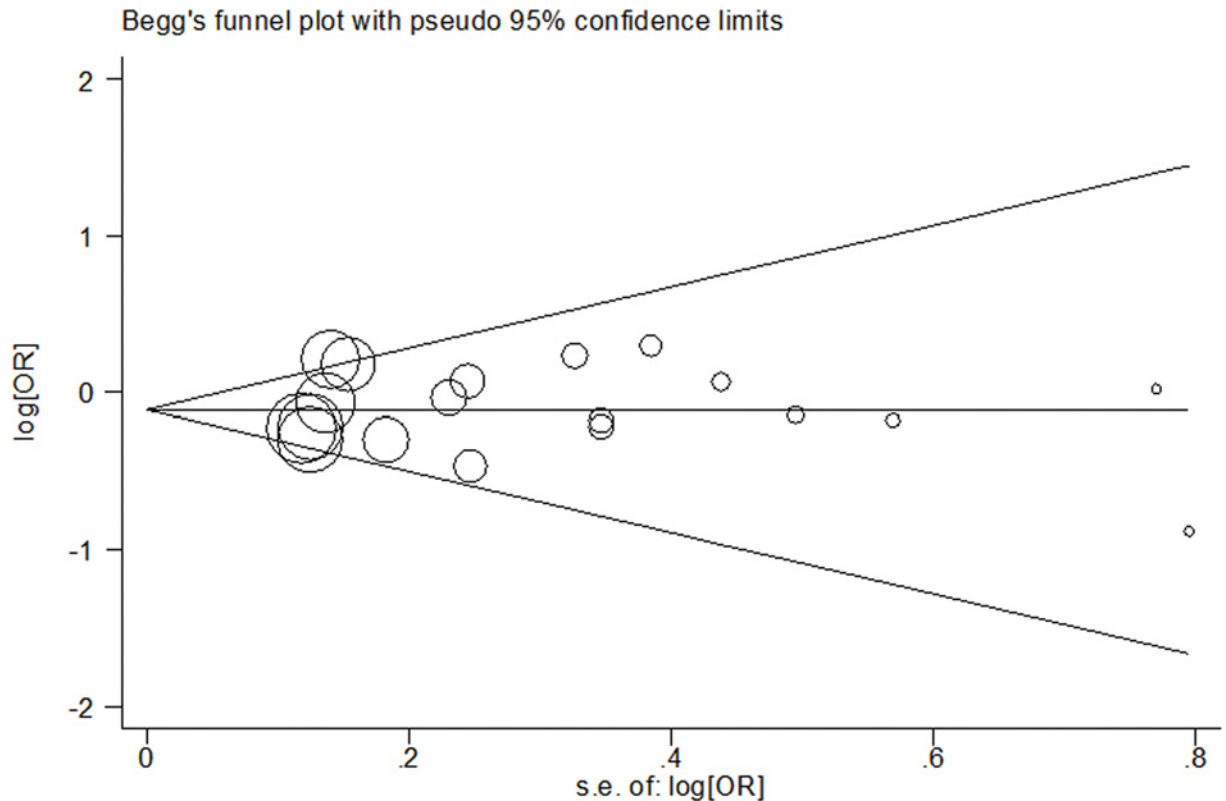


Figure 3. Begg's funnel plot of meta-analysis of the association between *MTHFR* rs1801133 polymorphism and HCC risk (recessive model, fixed-effects model)

Subgroup analyses were carried out according to the following terms: ethnicity (Caucasians or Asians or mixed), sample sizes (<1000 or ≥ 1000 subjects), control type [normal/healthy subjects or hepatitis/virus related patients or not available (NA)] and source of control [hospital-based (HB) or population-based (PB) or NA]. We pooled seven case-control studies (including 2,435 HCC cases and 1,642 hepatitis/virus related patients) and found an association between *MTHFR* rs1801133 polymorphism and decreased risk of HCC in hepatitis/virus related patients (recessive model: OR, 0.85; 95%CI, 0.72–0.99; $P = 0.035$, and allele model: OR, 0.90; 95%CI, 0.81–0.99; $P = 0.028$, Table 3). When we conducted a subgroup analysis by ethnicity, null association between *MTHFR* rs1801133 C>T polymorphism and the risk of HCC was found.

Heterogeneity assessment

In some genetic models, heterogeneity was significant (Table 3). Subgroup analyses indicated that extreme heterogeneity existed in Asian populations, larger sample size investigation, HB study and normal/healthy control subgroups. If we excluded these subgroups in our meta-analysis, the heterogeneity significantly decreased.

Bias evaluation

We used Begg's and Egger's tests to identify the bias of publication among the included investigations. The shape of Begg's test seemed symmetrical (Figure 3). Egger's linear regression test also confirmed these evaluations.

Sensitivity analyses

By sequentially omitting an individual investigation, sensitivity analysis was carried out. This method is considered as a criterion for meta-analysis. The results indicated that the significance of the present study could not be altered by removing any case-control study (Figure 4), suggesting that our findings were stable.

Quality assessment

Table 2 presents the results of the quality evaluation. Each eligible study had an acceptable quality (scores ≥ 6).

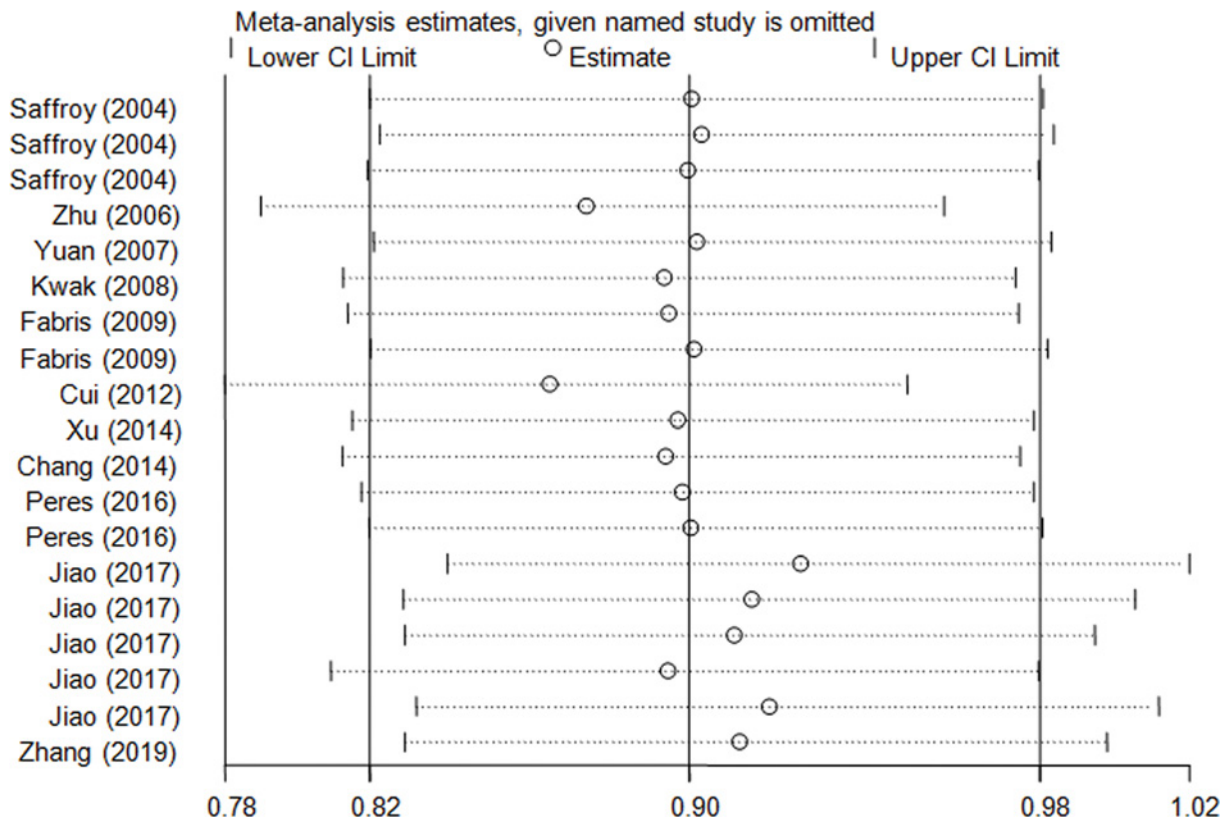


Figure 4. Sensitivity analysis of the influence of *MTHFR* rs1801133 polymorphism to HCC risk (recessive model, fixed-effects model)

Discussion

Accumulating investigations highlight that *MTHFR* rs1801133 polymorphism may be associated with the development of HCC. However, the findings of the previous case-control studies were conflicting, with several investigations suggesting a potential relationship, whereas others did not support the correlation. In this investigation, to explore whether the *MTHFR* rs1801133 polymorphism was implicated in the etiology of HCC, we carried out a pooled-analysis of 19 eligible studies, which recruited 6,102 HCC cases and 6,526 controls. This meta-analysis indicated that the *MTHFR* rs1801133 polymorphism was a protective factor for the development of HCC in the overall comparison. Compared with the previous study, this pooled-analysis first confirmed the association of *MTHFR* rs1801133 polymorphism with a decreased risk of HCC.

MTHFR rs1801133 polymorphism locates on 11796321 (NCBI Build 38) of Chromosome 1. Zhu and her/his colleagues first reported that *MTHFR* rs1801133 polymorphism might confer a risk to HCC [24]. In addition, Cui et al. also suggested that this polymorphism could increase the risk of HCC [20]. However, some case-control studies indicated that the rs1801133 polymorphism in *MTHFR* gene might decrease the susceptibility of HCC [21,25]. And most studies reported that this SNP in *MTHFR* gene could not alter the risk of HCC. Thus, the association of *MTHFR* rs1801133 polymorphism with the susceptibility of HCC was more conflicting. Here, we performed a pooled-analysis of nineteen eligible studies involving 6,102 HCC cases and 6,526 controls to explore the correlation of rs1801133 with the etiology of HCC. The results indicated that this SNP in *MTHFR* gene could be a protective factor for the occurrence of HCC. Two meta-analyses suggested that rs1801133 was not associated with HCC development [43,44]. Others pooled-analyses reported that *MTHFR* rs1801133 polymorphism was associated with an increased risk of HCC [45–48]. Compared with these early meta-analyses, our analysis included more large sample size studies [21,25]. It is worth mentioning that these more recent case-control studies have recruited more participants and reported that rs1801133 polymorphism was a protective factor for the development of HCC. Compared with the most recent meta-analysis [23], the merit of our study was the larger sample size and the detailed subgroup analysis. Combined the eligible studies, we observed that rs1801133 decreased the susceptibility of HCC in the overall comparison.

The quality score was evaluated in our study. Each eligible study had an acceptable quality (scores ≥ 6). This indicated that our findings were reliable. We also found an association between *MTHFR* rs1801133 polymorphism and decreased risk of HCC in hepatitis/virus related patients. Of late, in Asian population, some meta-analyses identified that *MTHFR* rs1801133 polymorphism decreased the risk of colorectal cancer [49,50]. Some publications [51,52] suggested that *MTHFR* rs1801133 C>T polymorphism (Ala→Val) could promote the level of 5,10-methylene-THF for DNA synthesis, which might be protective to carcinogenesis. In the future, a functional study should be carried out to address how this Ala→Val substitution could decrease the risk of HCC.

Heterogeneity was identified in the overall comparison. In the present study, we conducted subgroup analyses to explore the major source among the eligible studies. Subgroup analysis suggested that major heterogeneity might be due to different populations, sample size, and characteristics of controls.

Some potential limitations should be addressed in this pooled-analysis. First, only published investigations were eligible in our study. Thus, the number of included case-control studies might be inadequate. Second, for lacking of sufficient data, only crude ORs and CIs were calculated. Third, the controls in some of the case-control studies were hepatitis or virus related patients. Fourth, a recent investigation contained some subgroups, we treated them as independent case-control studies. However, in this literature, the same HCC group was used in different stratified analysis. Finally, our study did not focus on the gene-gene and gene-environment interactions.

In summary, the present pooled-analysis highlights that *MTHFR* rs1801133 polymorphism is a protective factor for the occurrence of HCC, especially in hepatitis/virus related patients. The relationship of *MTHFR* rs1801133 polymorphism with HCC risk warrants a further determination.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Funding

The project was supported by the Application and Basic Research Project of Changzhou City [grant number CJ20180068].

Author Contribution

Conceived and designed the experiments: S.Z. and L.L. Performed the experiments: S.Z. and J.J. Analyzed the data: W.T. and S.Z. Contributed reagents/materials/analysis tools: L.L. Wrote the manuscript: S.Z. and J.J.

Acknowledgements

We wish to thank Dr Yan Liu (Genesky Biotechnologies Inc., Shanghai, China) for technical support.

Abbreviations

CI, confidence interval; HCC, hepatocellular carcinoma; *MTHFR*, methylenetetrahydrofolate reductase; OR, odds ratio.

References

- Bray, F., Ferlay, J., Soerjomataram, I. et al. (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **68**, 394–424
- Pinero, F., Pages, J., Marciano, S. et al. (2018) Fatty liver disease, an emerging etiology of hepatocellular carcinoma in Argentina. *World J. Hepatol.* **10**, 41–50, PubMed Central PMCID: PMC5787683
- Liew, Z.H., Goh, G.B., Hao, Y. et al. (2018) Comparison of Hepatocellular Carcinoma in Patients with Cryptogenic Versus Hepatitis B Etiology: A Study of 1079 Cases Over 3 Decades. *Dig. Dis. Sci.* **64**, 585–590
- Ioannou, G.N., Green, P., Lowy, E. et al. (2018) Differences in hepatocellular carcinoma risk, predictors and trends over time according to etiology of cirrhosis. *PLoS One* **13**, e0204412, PubMed Central PMCID: PMC6160079, <https://doi.org/10.1371/journal.pone.0204412>
- Jaquet, A., Tchounga, B., Tanon, A. et al. (2018) Etiology of hepatocellular carcinoma in West Africa, a case-control study. *Int. J. Cancer* **143**, 869–877, PubMed Central PMCID: PMC6041181, <https://doi.org/10.1002/ijc.31393>
- Ghouri, Y.A., Mian, I. and Rowe, J.H. (2017) Review of hepatocellular carcinoma: Epidemiology, etiology, and carcinogenesis. *J. Carcinogenesis* **16**, 1, PubMed Central PMCID: PMC5490340
- Kopp, M., Morisset, R. and Rychlik, M. (2017) Characterization and Interrelations of One-Carbon Metabolites in Tissues, Erythrocytes, and Plasma in Mice with Dietary Induced Folate Deficiency. *Nutrients* **9**, pii: E462, <https://doi.org/10.3390/nu9050462>
- Jain, M., Pandey, P., Tiwary, N.K. et al. (2012) MTHFR C677T polymorphism is associated with hyperlipidemia in women with polycystic ovary syndrome. *J. Human Reprod. Sci.* **5**, 52–56, PubMed Central PMCID: PMC3409921, <https://doi.org/10.4103/0974-1208.97802>

- 9 Zara-Lopes, T., Galbiatti-Dias, A.L.S., Castanhole-Nunes, M.M.U. et al. (2019) Polymorphisms in MTHFR, MTR, RFC1 and CtsS genes involved in folate metabolism and thyroid cancer: a case-control study. *Archiv. Med. Sci.* **15**, 522–530, PubMed Central PMCID: PMC6425207, <https://doi.org/10.5114/aoms.2018.73091>
- 10 Lin, K.M., Yang, M.D., Tsai, C.W. et al. (2018) The Role of MTHFR Genotype in Colorectal Cancer Susceptibility in Taiwan. *Anticancer Res.* **38**, 2001–2006
- 11 Zhang, S., Chen, S., Chen, Y. et al. (2017) Investigation of methylenetetrahydrofolate reductase tagging polymorphisms with colorectal cancer in Chinese Han population. *Oncotarget* **8**, 63518–63527, PubMed Central PMCID: PMC5609940
- 12 Hesari, A., Maleksabet, A., Tirkani, A.N. et al. (2018) Evaluation of the two polymorphisms rs1801133 in MTHFR and rs10811661 in CDKN2A/B in breast cancer. *J. Cell. Biochem.*, <https://doi.org/10.1002/jcb.27517>
- 13 Ding, G., Wang, Y., Chen, Y. et al. (2017) Methylenetetrahydrofolate reductase tagging polymorphisms are associated with risk of esophagogastric junction adenocarcinoma: a case-control study involving 2,740 Chinese Han subjects. *Oncotarget* **8**, 111482–111494, PubMed Central PMCID: PMC5762337
- 14 Ding, H., Wang, Y., Chen, Y. et al. (2017) Methylenetetrahydrofolate reductase tagging polymorphisms are associated with risk of non-small cell lung cancer in eastern Chinese Han population. *Oncotarget* **8**, 110326–110336, PubMed Central PMCID: PMC5746385
- 15 Xia, X., Duan, Y., Cui, J. et al. (2017) Association of methylenetetrahydrofolate reductase gene-gene interaction and haplotype with susceptibility to acute lymphoblastic leukemia in Chinese children. *Leuk. Lymphoma* **58**, 1887–1892
- 16 Ramirez-Pacheco, A., Moreno-Guerrero, S., Alamillo, I. et al. (2016) Mexican Childhood Acute Lymphoblastic Leukemia: A Pilot Study of the MDR1 and MTHFR Gene Polymorphisms and Their Associations with Clinical Outcomes. *Genetic Test. Mole. Biomark.* **20**, 597–602
- 17 Wei, L., Niu, F., Wu, J. et al. (2019) Association study between genetic polymorphisms in folate metabolism and gastric cancer susceptibility in Chinese Han population: A case-control study. *Mole. Genet. Genomic Med.* **7**, e633, PubMed Central PMCID: PMC6503009
- 18 Lv, C., Bai, Z., Liu, Z. et al. (2015) Renal cell carcinoma risk is associated with the interactions of APOE, VHL and MTHFR gene polymorphisms. *Int. J. Clin. Exp. Pathol.* **8**, 5781–5786, PubMed Central PMCID: PMC4503168
- 19 Chang, S.C., Chang, P.Y., Butler, B. et al. (2014) Single nucleotide polymorphisms of one-carbon metabolism and cancers of the esophagus, stomach, and liver in a Chinese population. *PLoS One* **9**, e109235, PubMed Central PMCID: PMC4206280, <https://doi.org/10.1371/journal.pone.0109235>
- 20 Cui, L.H., Song, Y., Si, H. et al. (2012) Folate metabolism-related gene polymorphisms and susceptibility to primary liver cancer in North China. *Med. Oncol.* **29**, 1837–1842, <https://doi.org/10.1007/s12032-011-0066-y>
- 21 Jiao, X., Luo, Y., Yang, B. et al. (2017) The MTHFR C677T mutation is not a risk factor recognized for HBV-related HCC in a population with a high prevalence of this genetic marker. *Infect. Genet. Evol.: J. Mole. Epidemiol. Evol. Genet. Infect. Dis.* **49**, 66–72, <https://doi.org/10.1016/j.meegid.2017.01.008>
- 22 Kwak, S.Y., Kim, U.K., Cho, H.J. et al. (2008) Methylenetetrahydrofolate reductase (MTHFR) and methionine synthase reductase (MTRR) gene polymorphisms as risk factors for hepatocellular carcinoma in a Korean population. *Anticancer Res.* **28**, 2807–2811
- 23 Su, H. (2019) Correlation Between MTHFR Polymorphisms and Hepatocellular Carcinoma: A Meta-analysis. *Nutr. Cancer* **71**, 1055–1060, <https://doi.org/10.1080/01635581.2019.1577985>
- 24 Zhu, Z.Z., Cong, W.M., Liu, S.F. et al. (2006) A study on the association of MTHFR C677T polymorphism with genetic susceptibility to hepatocellular carcinoma. *Zhonghua Gan Zang Bing Za Zhi* **14**, 196–198
- 25 Zhang, S., Lin, J., Jiang, J. et al. (2019) Association between methylenetetrahydrofolate reductase tagging polymorphisms and susceptibility of hepatocellular carcinoma: a case-control study. *Biosci. Rep.* **39**, pii: BSR20192517, PubMed Central PMCID: PMC6852349, <https://doi.org/10.1042/BSR20192517>
- 26 Fabris, C., Toniutto, P., Falletti, E. et al. (2009) MTHFR C677T polymorphism and risk of HCC in patients with liver cirrhosis: role of male gender and alcohol consumption. *Alcohol Clin. Exp. Res.* **33**, 102–107, <https://doi.org/10.1111/j.1530-0277.2008.00816.x>
- 27 Saffroy, R., Pham, P., Chiappini, F. et al. (2004) The MTHFR 677C > T polymorphism is associated with an increased risk of hepatocellular carcinoma in patients with alcoholic cirrhosis. *Carcinogenesis* **25**, 1443–1448
- 28 Peres, N.P., Galbiatti-Dias, A.L., Castanhole-Nunes, M.M. et al. (2016) Polymorphisms of folate metabolism genes in patients with cirrhosis and hepatocellular carcinoma. *World J. Hepatol.* **8**, 1234–1243, PubMed Central PMCID: PMC5067443, <https://doi.org/10.4254/wjh.v8.i29.1234>
- 29 Yuan, J.M., Lu, S.C., Van Den Berg, D. et al. (2007) Genetic polymorphisms in the methylenetetrahydrofolate reductase and thymidylate synthase genes and risk of hepatocellular carcinoma. *Hepatology* **46**, 749–758, PubMed Central PMCID: PMC2391240, <https://doi.org/10.1002/hep.21735>
- 30 Higgins, J.P., Thompson, S.G., Deeks, J.J. et al. (2003) Measuring inconsistency in meta-analyses. *BMJ* **327**, 557–560, PubMed Central PMCID: PMC192859, <https://doi.org/10.1136/bmj.327.7414.557>
- 31 DerSimonian, R. and Laird, N. (1986) Meta-analysis in clinical trials. *Control. Clin. Trials* **7**, 177–188, [https://doi.org/10.1016/0197-2456\(86\)90046-2](https://doi.org/10.1016/0197-2456(86)90046-2)
- 32 Mantel, N.H.W. (1959) Statistical aspects of the analysis of data from retrospective studies of disease. *J. Natl. Cancer Inst.* **22**, 719–748
- 33 Tang, W., Wang, Y., Pan, H. et al. (2019) Association of miRNA-499 rs3746444 A>G variants with adenocarcinoma of esophagogastric junction (AEG) risk and lymph node status. *OncoTargets Ther.* **12**, 6245–6252, PubMed Central PMCID: PMC6690596
- 34 Tang, W., Liu, J., Zhong, Z. et al. (2019) Association of metabolism-related genes polymorphisms with adenocarcinoma of the oesophagogastric junction: Evidence from 2261 subjects. *J. Cell. Biochem.* **120**, 18689–18701, PubMed Central PMCID: PMC6771939, <https://doi.org/10.1002/jcb.29167>
- 35 Rong, G., Tang, W., Wang, Y. et al. (2019) Investigation of leptin receptor rs1137101 G>A polymorphism with cancer risk: evidence from 35936 subjects. *Biosci. Rep.* **39**, pii: BSR20182240, PubMed Central PMCID: PMC6597850
- 36 Thakkinstian, A., McEvoy, M., Minelli, C. et al. (2005) Systematic review and meta-analysis of the association between {beta}2-adrenoceptor polymorphisms and asthma: a HuGE review. *Am. J. Epidemiol.* **162**, 201–211, <https://doi.org/10.1093/aje/kwi184>

- 37 Camargo, M.C., Mera, R., Correa, P. et al. (2006) Interleukin-1beta and interleukin-1 receptor antagonist gene polymorphisms and gastric cancer: a meta-analysis. *Cancer Epidemiol. Biomark. Prevent.* **15**, 1674–1687
- 38 Guo, J., Jin, M., Zhang, M. et al. (2012) A genetic variant in miR-196a2 increased digestive system cancer risks: a meta-analysis of 15 case-control studies. *PLoS One* **7**, e30585, PubMed Central PMCID: PMC3265498, <https://doi.org/10.1371/journal.pone.0030585>
- 39 D'Amico, M., Pasta, L. and Sammarco, P. (2009) MTHFR C677T, PAI1 4G-4G, V Leiden Q506, and prothrombin G20210A in hepatocellular carcinoma with and without portal vein thrombosis. *J. Thromb. Thrombol.* **28**, 70–73
- 40 Qiao, K., Zhang, S., Trieu, C. et al. (2017) Genetic Polymorphism of MTHFR C677T Influences Susceptibility to HBV-Related Hepatocellular Carcinoma in a Chinese Population: a Case-Control Study. *Clin. Lab.* **63**, 787–795
- 41 Wang, C., Xie, H., Lu, D. et al. (2018) The MTHFR polymorphism affect the susceptibility of HCC and the prognosis of HCC liver transplantation. *Clin. Transl. Oncol.* **20**, 448–456
- 42 Zhang, H., Liu, C., Han, Y.C. et al. (2015) Genetic variations in the one-carbon metabolism pathway genes and susceptibility to hepatocellular carcinoma risk: a case-control study. *Tumour Biol.* **36**, 997–1002, <https://doi.org/10.1007/s13277-014-2725-z>
- 43 Su, H. and Zhang, G. (2019) Correlation between Methylenetetrahydrofolate Reductase Polymorphisms and Hepatocellular Carcinoma: A Meta-Analysis. *Ann. Nutr. Metab.* **74**, 251–256
- 44 Qin, X., Peng, Q., Chen, Z. et al. (2013) The association between MTHFR gene polymorphisms and hepatocellular carcinoma risk: a meta-analysis. *PLoS One* **8**, e56070, PubMed Central PMCID: PMC3573065, <https://doi.org/10.1371/journal.pone.0056070>
- 45 Jin, F., Qu, L.S. and Shen, X.Z. (2009) Association between the methylenetetrahydrofolate reductase C677T polymorphism and hepatocellular carcinoma risk: a meta-analysis. *Diagn. Pathol.* **4**, 39, PubMed Central PMCID: PMC2788519, <https://doi.org/10.1186/1746-1596-4-39>
- 46 Qi, X., Sun, X., Xu, J. et al. (2014) Associations between methylenetetrahydrofolate reductase polymorphisms and hepatocellular carcinoma risk in Chinese population. *Tumour Biol.* **35**, 1757–1762, <https://doi.org/10.1007/s13277-013-1529-x>
- 47 Qi, Y.H., Yao, L.P., Cui, G.B. et al. (2014) Meta-analysis of MTHFR C677T and A1298C gene polymorphisms: association with the risk of hepatocellular carcinoma. *Clin. Res. Hepatol. Gastroenterol.* **38**, 172–180, <https://doi.org/10.1016/j.clinre.2013.10.002>
- 48 Sun, H., Han, B., Zhai, H. et al. (2014) Significant association between MTHFR C677T polymorphism and hepatocellular carcinoma risk: a meta-analysis. *Tumour Biol.* **35**, 189–193, <https://doi.org/10.1007/s13277-013-1023-5>
- 49 Teng, Z., Wang, L., Cai, S. et al. (2013) The 677C>T (rs1801133) polymorphism in the MTHFR gene contributes to colorectal cancer risk: a meta-analysis based on 71 research studies. *PLoS One* **8**, e55332, PubMed Central PMCID: PMC3577825
- 50 Guo, X.P., Wang, Y., Zhao, H. et al. (2014) Association of MTHFR C677T polymorphisms and colorectal cancer risk in Asians: evidence of 12,255 subjects. *Clin. Transl. Oncol.* **16**, 623–629
- 51 Taioli, E., Garza, M.A., Ahn, Y.O. et al. (2009) Meta- and pooled analyses of the methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism and colorectal cancer: a HuGE-GSEC review. *Am. J. Epidemiol.* **170**, 1207–1221, PubMed Central PMCID: PMC2781761, <https://doi.org/10.1093/aje/kwp275>
- 52 Frosst, P., Blom, H.J., Milos, R. et al. (1995) A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. *Nat. Genet.* **10**, 111–113, <https://doi.org/10.1038/ng0595-111>