Review Article



Check for updates

Therapeutic targeting of TRAIL death receptors

Francesca Di Cristofano^{1,2,3,6}, Andrew George^{1,2,3,6}, Vida Tajiknia^{1,2,3,6}, Maryam Ghandali^{1,2,3,6}, Laura Wu^{1,2,3,6}, Yiqun Zhang^{1,2,3,6}, Praveen Srinivasan^{1,2,3,6}, Jillian Strandberg^{1,2,3,6}, Marina Hahn^{1,2,3,6}, Ashley Sanchez Sevilla Uruchurtu^{1,2,3,6}, Attila A. Seyhan^{1,2,3,6}, Benedito A. Carneiro^{1,2,3,5,6}, Lanlan Zhou^{1,2,3,6}, Kelsev E. Huntington^{1,2,3,4,6} and [©] Wafik S. El-Deirv^{1,2,3,4,5,6}

¹Laboratory of Translational Oncology and Experimental Cancer Therapeutics, The Warren Alpert Medical School, Brown University, Providence, RI 02903, U.S.A.; ²The Joint Program in Cancer Biology, Brown University and the Lifespan Health System, Providence, RI 02903, U.S.A.; ³Department of Pathology and Laboratory Medicine, The Warren Alpert Medical School, Brown University, Providence, RI 02903, U.S.A.; ⁴Pathobiology Graduate Program, The Warren Alpert Medical School, Brown University, Providence, RI 02903, U.S.A.; ⁵Hematology-Oncology Division, Department of Medicine, Rhode Island Hospital and Brown University, Providence, RI 02903, U.S.A.; ⁶Legorreta Cancer Center at Brown University, The Warren Alpert Medical School, Brown University, Providence, RI 02903, U.S.A.

Correspondence: Wafik S. El-Deiry (wafik@brown.edu)



y and Experimental Cancer Therapeutics, The Warren Alpert Medical School, Brown University, Providence, RI 02903, U.S.A.; ²The Joint Inversity and the Lifespan Health System, Providence, RI 02903, U.S.A.; ¹Department of Pathology and Laboratory Medicine, The Warren sity, Providence, RI 02903, S.A.; ⁴Pathobiology Graduate Program, The Warren Alpert Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Medical School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at School, Brown University, Providence, RI 02903, U.S.A.; ¹Legorreta Cancer Center at Center at Center at Center at

progression and was included in 2000 among the original key hallmarks of cancer. Since then, much ट्रु work has been done to pursue the targeted activation of apoptotic pathways in cancer cells as a viable $\overline{\aleph}$ therapeutic strategy [1,2].

Apoptosis itself was first identified as a controlled cell death pathway in 1972, and years since have seen the emergence of two broad 'pathways' driving the process: cell 'intrinsic' and cell 'extrinsic' [3,4]. Though interplay between the two pathways has been observed, the intrinsic pathway largely relies on the mitochondria and the BCL-2 family of proteins to trigger apoptosis in response to internal stress borne by the cell, while for the extrinsic pathway, ligand binding to cell surface death receptors translates various death signals into a caspase cascade which ultimately leads to cell death [5] (Figure 1).

One such death receptor ligand is tumor necrosis factor-related apoptosis-inducing ligand (TRAIL), which binds death receptors DR4 and DR5 and several decoy receptors. While DR4 and DR5 are tumor cell-expressed TRAIL-binding receptors that contain death domains to trigger the induction of apoptosis, decoy receptors lack the death domain and are primarily expressed on normal cells. This ability of TRAIL to preferentially induce apoptosis in tumor cells, but not normal cells, provides a

Received: 25 August 2022 Revised: 26 November 2022 Accepted: 7 December 2022

Version of Record published: 11 January 2023



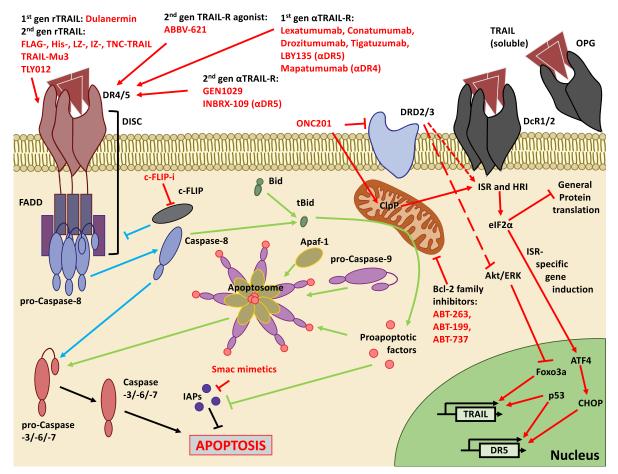


Figure 1. Cell death signaling activated by TRAIL and TRAIL pathway therapeutics.

TRAIL pathway signaling is initiated upon TRAIL binding to DR4 or DR5, driving receptor trimerization and recruitment of the FADD to form the DISC. DR4 and DR5 compete with decoy receptors DcR1, DcR2, and the soluble OPG for TRAIL. Activation of DR4 or DR5 triggers apoptotic signaling initially through initiator caspases, primarily caspase-8. Extrinsic apoptotic signaling (blue arrows) continues through activation of executioner caspases-3, -6, and -7, resulting in apoptotic death. Caspase-8 additionally activates intrinsic apoptotic signaling (green arrows) through facilitating conversion of Bid to tBid, driving proapoptotic factor release from the mitochondrion, apoptosome assembly, and ultimately also converging on executioner caspase activation. Several therapies, as described in the text, have been developed to engage at several points along this path (red labels and red arrows). Some, including first and second generation rTRAIL, TRAs, and antibodies, directly agonize DR4 and DR5 (unless otherwise indicated, these agonists have described activity against both DR4 and DR5). Downstream, inhibitors of c-FLIP prevent its competition with pro-caspase-8 for the FADD to enhance caspase-8 activation. Bcl-2 family inhibitors act to mitigate the function of antiapoptotic members of the Bcl-2 family, especially Bcl-2 and Bcl-xL. Smac mimetics also help to sequester anti-apoptotic IAPs. On a transcriptional level, ONC201 (initially referred to as TIC10 for <u>TRAIL-inducing compound 10</u>) antagonizes DRD2 and DRD3 while agonizing CIpP, ultimately bringing about growth arrest through the integrated stress response (ISR). Moreover, downstream effects of ONC201 (alongside p53) drive increased expression of TRAIL and DR5. Pointed arrows indicate activation; blunted arrows indicate inhibition.

therapeutic window. Thus, following its discovery in 1996, the tumoricidal potential of TRAIL was pursued along with a search for TRAIL-based treatments [6,7] (Figure 1). First explored were TRAIL receptor-based therapies, namely recombinant human TRAIL (rhTRAIL) and death receptor agonistic antibodies. We cloned Death Receptor 5 (DR5; KILLER/DR5) as a p53 target gene which provided a rationale for the combination of TRAIL with DNA damaging therapeutics [8]. These first-generation TRAIL treatments were found to both directly lead to tumor cell death as well as result in the sensitization of tumor cells to other chemotherapies and



radiotherapies [9]. However, such strategies have limitations, ranging from weak binding to death receptors, to poor pharmacokinetics, to acquired resistance. As such, novel second-generation treatment strategies have been sought to address these shortcomings, working to either increase or supplement the core efficacy of TRAIL receptor-based therapies, or to prevent the acquisition of resistance and re-sensitize tumors to first-generation agents. In addition, the recent discovery and emerging clinical success of TIC10/ONC201 have led to its recognition as a 'next-generation' approach.

In this review, we provide a brief overview of key therapies exploiting the TRAIL pathway, along with their limitations and subsequent advances, concluding with a perspective on novel strategies of therapeutic TRAIL pathway-mediated induction of tumor cell death.

TRAIL signaling pathway

TRAIL itself can exist in one of two forms: either as a transmembrane protein form expressed by various cell types, or as a soluble form composed of the shed extracellular domain of membrane-bound TRAIL which self-trimerizes [9]. As previously mentioned, both forms are capable of binding target receptors DR4 and DR5 to trigger an apoptotic signal on the death-receptor expressing cell, but may also bind membrane-bound decoy receptors DcR1 and DcR2 or soluble decoy receptor OPG [10]. Consequently, an initial point of apoptotic signal modulation exists through the expression of such decoy receptors, which ultimately can reduce TRAIL-induced death signaling through competition with DR4 and DR5. Indeed, decoy receptor expression has been implicated, among other mechanisms, in the resistance of normal cells to TRAIL [11,12] as well as in cancer cell resistance to TRAIL [13].

Cell death via the extrinsic apoptotic pathway is initiated by ligands such as TRAIL binding to their target death receptors. Upon ligand binding, the death receptors trimerize, leading to their activation. This is followed by the recruitment of the adaptor protein Fas-associated protein with death domain (FADD) as well as initiator caspases-8 and -10. Together, these molecules assemble into the death-inducing signaling complex (DISC) [14]. While lower-order death receptor clustering (i.e. trimerization) is essential to efficient DISC formation, the higher-order clustering of multiple complexes has emerged as being another important factor in apoptotic signal transduction [15]. Downstream of DISC formation, resultant activation of initiator caspase-8 leads to the activation of executioner caspases-3, -6, and -7, which leads to cell death.

In so-called Type I cells, this pathway alone is sufficient to activate the executioner caspases and induce cell death. Conversely, in Type II cells, additional amplification via the intrinsic mitochondrial apoptotic pathway is required. In these cells, Bid cleavage by caspase-8 is followed by its myristoylation and translocation to the mitochondria, where it, along with Bax and Bak, allows for the release of proapoptotic factor cytochrome C [16,17]. Cytochrome C complexes with apoptotic peptidase activating factor 1 (Apaf-1) and initiator caspase-9 to form an executioner caspase cascade-activating apoptosome. TRAIL signaling at DR5 receptors has additionally been shown to translocate proapoptotic Bim and Bax to lysosomes, resulting in a release of cathepsin B that further enhances mitochondrial outer membrane permeabilization and executioner caspase activation through the recruitment of phosphofurin acidic cluster sorting protein-2 (PACS-2) to DR5-positive endosomes [18]. C-Myc was identified as a major determinant of TRAIL sensitivity [19].

TRAIL receptor-based therapies: first generation

An early study in the exploration of TRAIL pathway-based therapeutics assessed the expression of TRAIL receptors by immunohistochemistry in primary glioblastoma (GBM) specimens. The study showed that higher receptor expression is correlated with increased patient survival, supporting the important role played by TRAIL in commanding patient prognosis [20,21]. Thus, receptor-targeted therapies garnered much attention based on their potential as effective anticancer agents. Many first-generation TRAIL receptor-targeted agents focused on recombinant forms of TRAIL as well as agonistic antibodies against TRAIL death receptors.

Recombinant forms of TRAIL

One strategy to target the TRAIL pathway is to develop bioactive recombinant soluble forms of TRAIL. One such example is dulanermin (AMG-951), which is a soluble recombinant human Apo2Ligand/TRAIL (rhApo2L/TRAIL) protein that induces apoptosis in target cells via its binding and activation of DR4 (i.e. TRAIL-R1) and DR5 (i.e. TRAIL-R2), though it is also known to bind to decoy receptors [22]. Developed by Amgen/Genentech, dulanermin is the only rhTRAIL to have reached clinical trials. In a phase 1 dose-escalation



study in patients with advanced cancer, dulanermin was shown to have a tolerable safety profile; however, its observed anticancer activity was limited [22]. Of note, the mean terminal phase half-life $(t_{1/2})$ of rhTRAIL ranges from 0.56 to 1.02 h, likely restricting its ability to reach tumor cells [22,23]. Moreover, because rhTRAIL binds to decoy receptors, its delivery to DR4/5 is diluted. This weakened agonism is worsened by the fact that although rhTRAIL induces lower-order receptor trimerization, it has a limited capacity to induce higher-order receptor clustering, resulting in a weak apoptotic signal [24,25]. Meanwhile, other first-generation TRAIL pathway-based therapies were developed.

Death receptor agonist antibodies

The development of agonistic antibodies against DR4/5 presented another strategy to target TRAIL receptors. Unlike rhTRAIL, which binds to both death receptors and decoy receptors, diluting its apoptotic effect, agonistic antibodies specifically target DR4 or DR5 to activate apoptosis in target cells. Moreover, receptor-targeted monoclonal antibodies (TRAIL-R mAbs) have a significantly longer half-life than rhTRAIL. Therefore, the development of these therapeutic antibodies was pursued as an anti-cancer strategy.

Preclinical studies showed promising results, with significant antitumor activity without toxicity seen both in vivo and in vitro [26]. Consequently, several first-generation TRAIL-R mAbs have been tested in clinical trials including DR5-targeted agents such as lexatumumab, conatumumab, drozitumab, tigatuzumab, and LBY135. DR5-targeted mAbs showed results in early clinical trials that only modestly improved upon clinical results observed with rhTRAIL. For example, in a phase 1 trial of lexatumumab in patients with advanced cancers, the mAb was well-tolerated and was associated with stable disease in several patients [27]. Safety was also demonstrated in the first-in-human study of conatumumab in adult patients with advanced solid tumors [28]. In this study, a single patient with colorectal cancer (CRC) had stable disease for 24 weeks and showed a 24% reduction in tumor size by RECIST (Response Evaluation Criteria in Solid Tumors) criteria. Drozitumab is another DR5-targeted mAb that was evaluated in a phase 1 study in patients with advanced malignancies [29]. In this study, three minor responses were observed in patients with CRC, granulosa cell ovarian cancer, and chondrosarcoma, although no objective responses were observed. Moreover, in a phase 1 trial of DR5-targeted tigatuzumab, stable disease and a tolerable safety profile were observed [30]. A DR4-targeted agent, mapatumumab, which is a fully human mAb also entered clinical trials. In a phase 1 study in patients with advanced solid malignancies, mapatumumab was observed to be well-tolerated, however, no objective responses were observed [31]. Another approach using DR4 atrimers was explored although this was not pursued in the clinic [32].

Thus, although successful in preclinical studies, TRAIL-R mAbs have performed modestly in clinical trials. Despite improving upon the pharmacokinetic ability and specificity of rhTRAIL, the agonistic ability of TRAIL-R mAbs remains limited. One factor contributing to this is that mAbs have an inherently bivalent structure, only allowing for their cross-linking to two death receptors. As lower-order receptor trimerization is required to ensure efficient DISC-formation, additional cross-linking is needed for sufficient apoptotic signaling [33]. Another limiting factor is that while first-generation TRAIL-R mAbs are DR4- or DR5-specific, there are differences in the ability of each receptor to transmit apoptotic signals that vary between tissue types [34], pointing to the need for antibodies that target both receptors.

Improving upon TRAIL receptor-based therapies

First-generation therapies were thus divided into two categories: recombinant forms of TRAIL and receptorspecific agonistic antibodies. Although these early TRAIL receptor agonists (TRAs) were shown to be promising in preclinical data [35,36] and well-tolerated in patients [22,37], they ultimately proved to have limited clinical anticancer activity. This can be attributed to several factors, including poor pharmacokinetics, limited agonistic ability, and inherent or acquired resistance [2,38]. Second-generation TRAs were developed to target these limitations by improving upon the pharmacokinetic profile and agonistic ability of these agents and circumventing resistance through combinatorial approaches.

TRAIL derivatives

N-terminal tags

An early second-generation approach was to modulate the rhTRAIL protein through the addition of N-terminal tags to improve its pharmacokinetic profile and agonistic ability. The small molecular weight of rhTRAIL is what leads to its rapid clearance via renal filtration. Adding a molecular tag to rhTRAIL increases



its size and thus improves its half-life [34,39]. Several 'tagged' forms of rhTRAIL were developed, including the addition of Flag (FLAG-TRAIL), poly-histidine (His-TRAIL), leucine zipper (LZ-TRAIL), isoleucine zipper (IZ-TRAIL), or tenascin-C (TNC) oligomerization domain (TNC-TRAIL) [40]. Both FLAG-TRAIL and His-TRAIL were created to facilitate the purification process [38]. On the other hand, LZ-TRAIL, IZ-TRAIL, and TNC-TRAIL were all developed to stabilize rhTRAIL homotrimerization through hydrophobic interactions within the LZ, IZ, and TNC sequences [6,41,42]. This increased stability enhances the agonistic activity and apoptotic potential of rhTRAIL. Despite promising preclinical data, FLAG-TRAIL and His-TRAIL were found to induce significant apoptosis in primary human hepatocytes [43]. While LZ-TRAIL, IZ-TRAIL, and TNC-TRAIL were not found to be significantly hepatotoxic [41], concern over on-target or off-target toxicity hindered the translation of tagged rhTRAIL into the clinic.

TLY012

Another approach to improving rhTRAIL is to covalently link it to molecules that enhance its pharmacokinetic profile. One such fusion product is TLY012, an N-terminal PEGylated rhTRAIL. Like the tags, the covalent attachment of polyethylene glycol (PEG) to a therapeutic protein increases its size, thereby reducing its clear-ance by renal filtration and increasing its half-life [44–46]. Indeed, TLY012 was shown to have a greatly increased half-life compared with rhTRAIL in a molecular weight-dependent manner, with 20K- and 30K-PEG-TRAIL having circulating half-lives of 12 and 18 h, respectively, compared with the 1 h half-life of rhTRAIL in ICR mice [47]. This increased half-life was associated with a greater anti-tumor effect both *in vitro* and *in vivo* in CRC models [47]. Later, TLY012 was shown also to have activity against fibrotic cells [48,49]. While TLY012 is entering clinical trials for the treatment of systemic sclerosis, further exploration of its anticancer potential as a single agent and in combination has been ongoing (further discussed below).

TRAIL-Mu3

A similar approach to enhancing the pharmacokinetic profile of rhTRAIL involves making modifications to the amino acid sequence to alter the chemical properties of the complete peptide. Mutagenesis of amino acid sequence VRERGPQR (114–121) into RRRRRRR, for instance, has been shown to enhance the membrane-penetrating abilities of wild-type TRAIL, leading to the development of a novel membrane-penetrating TRAIL-Mu3 [50]. TRAIL-Mu3 has been shown to have enhanced efficacy both in *in vitro* and *in vivo* preclinical models against pancreatic cancers over both rhTRAIL as well as gencitabine [50]. TRAIL-Mu3 has a greater tendency to activate caspase cascade in pancreatic cancer cell lines compared with rhTRAIL [51]. Though TRAIL-Mu3 is currently confined to the preclinical setting, its deployment into the clinic may allow for a novel therapeutic strategy against gemcitabine-resistant pancreatic cancers.

Eftozanermin alfa

As previously noted, a major limitation of both first- and second-generation rhTRAIL, receptor-specific mAbs, and TRAIL derivatives is their inability to induce efficient lower- and higher-order receptor clustering, leading to reduced apoptotic signaling. In the case of mAbs, it has been shown that due to the bivalent structure of antibodies, additional cross-linking of the Fc region of antibodies to Fc γ receptors (Fc γ R) is necessary for lower-order clustering and trimerization to occur [33,52]. However, IgG is known to compete with these antibodies for this interaction. Mouse models have very low levels of IgG as compared with cancer patients, providing a likely explanation for the discrepancy between preclinical studies and clinical trials of receptor-specific mAbs [33]. With this in mind, a new family of TRAIL-R agonists was produced.

APG350 is a first-in-class prototype of Eftozanermin alfa (ABBV-621). It was designed to increase receptor clustering while remaining independent of Fc γ R cross-linking. The molecule is composed of two single chain TRAIL trimers dimerized via fusion to the Fc region of IgG1 and is therefore able to bind to six TRAIL receptors (both DR4 and DR5) with limited Fc γ R binding capability [33]. APG350 was shown to have an enhanced lower-order clustering efficiency as compared with TRAIL-R mAbs, and because it can simultaneously bind two death receptor trimers, it has a greater ability to induce higher-order receptor clustering as compared with rhTRAIL and its derivatives [53,54]. Moreover, the later developed Eftozanermin alfa has an estimated human half-life of ~2 days [54]. Thus, Eftozanermin alfa clearly improves upon the agonistic ability and pharmacokinetic profile of first-generation TRAIL therapeutics and has consequently progressed into clinical trials (Table 1). A first-in-human study showed Eftozanermin alfa was well-tolerated in patients with advanced solid



Table 1 Ongoing clinical trials involving TRAIL-based therapeutics

Cancer type	Combined with	Phase	Status	Clinicaltrials.gov ID
Eftozanermin alfa				
Multiple myeloma	-	1	Recruiting	NCT04570631
Advanced solid tumors, hematological malignancies	-	1	Completed	NCT03082209
SCB-313				
Peritoneal malignancies	-	1	Completed	NCT03443674
Malignant pleural effusions	-	1	Completed	NCT03869697
Peritoneal carcinomatosis	-	1	Recruiting	NCT04047771
Malignant ascites	-	1	Completed	NCT04051112
Malignant pleural effusions	-	1	Completed	NCT04123886
GEN1029				
Solid tumors	-	1/2	Terminated	NCT03576131
INBRX-109				
Metastatic solid tumors, including sarcomas	-	1	Recruiting	NCT03715933
Conventional chondrosarcoma	-	2	Recruiting	NCT04950075
IGM-8444				
Solid tumors	Numerous agents	1	Recruiting	NCT04553692
ONC201				
Neuroendocrine	-	2	Active, not recruiting	NCT03034200
Acute myeloid leukemia	-	1	Recruiting	NCT03932643
Leukemia	-	1/2	Recruiting	NCT02392572
Multiple myeloma	-	1/2	Active, not recruiting	NCT02863991
Metastatic colorectal cancer	-	1/2	Terminated	NCT03791398
Ovarian cancer	Paclitaxel	2	Recruiting	NCT04055649
Glioma	-	2	Active, not recruiting	NCT03295396
H3 K27M Glioma	-	1	Recruiting	NCT03416530
Recurrent GBM, H3 K27M Glioma, midline glioma	-	2	Active, not recruiting	NCT02525692
Midline glioma	Radiation, Paxalisib, or Panobinostat	2	Recruiting	NCT05009992

tumors and hematological malignancies alone and in combination with venetoclax or chemotherapeutics [55,56] (NCT03082209). The results of dose-escalation and dose-optimization cohorts of patients with advanced solid tumors receiving Eftozanermin alfa monotherapy showed partial responses according to RECIST in three patients (two with CRC and one with pancreatic cancer) among the 105 patients participating in the clinical trial (PMID 35467243). Pharmacodynamics studies demonstrated saturation of Eftozanermin alfa binding sites on the TRIAL receptors and increased levels of M30 and M65 markers of apoptosis in the peripheral blood. Analysis of paired tumor specimens collected during the clinical trial also showed increase tumor infiltration of immune cells including CD4+ T cells in post-treatment biopsies compared with baseline tumor specimens as well as increased PARP cleavage and down-regulation of MEK/Erk/AKT pathway. Another phase 1B study is currently recruiting patients with relapsed or refractory multiple myeloma (NCT04570631).



Modified TRAIL-R antibodies

Besides TRAIL derivatives, modified TRAIL-R mAbs are another class of second-generation therapeutics. Unlike their bivalent first-generation counterparts which are unable to induce efficient receptor trimerization, these antibodies are multivalent, which allows them to bind more than just two receptors, thereby enhancing receptor clustering.

GEN1029 (HexaBody-DR5/DR5) is novel a mixture of two DR5-specific IgG1 mAbs, each targeting two distinct DR5 epitopes and containing E430G mutations that enhance antibody hexamerization via Fc-Fc interactions [57–59]. This unique structure results in the ability to induce lower- and higher-order receptor clustering in an Fc γ R-independent manner. GEN1029 has shown potent antitumor activity in preclinical models in a variety of solid and hematological malignancies [59,60]. A two-part safety trial consisting of a first-in-human dose-escalating phase 1 and an expansion phase 2A for a variety of solid tumors was recently terminated by its sponsor, Genmab (Copenhagen, Denmark), for unspecified reasons (NCT03576131) (Table 1).

INBRX-109 (Inhibrx, La Jolla, CA, U.S.A.) is another multivalent DR5 agonistic antibody. Previous attempts to produce multivalent antibodies led to the realization that excessive receptor clustering can cause hepatotoxicity (as shown by the terminated phase 1 trial for TAS266, a DR5-targeting tetravalent nanobody [61]). INBRX-109 was engineered to avoid hepatotoxicity by limiting its valency to four. Preclinical studies indeed showed that INBRX-109 maintained superior induction of apoptosis in cancer cells while sparing human hepatocytes. INBRX-109 entered an ongoing phase 1 clinical study to evaluate its single agent efficacy in subjects with locally advanced or metastatic tumors (NCT04950075). The drug was shown to be well-tolerated, which led to the FDA granting it orphan drug designation (ODD) in 2021 as well as a phase 2 study of INBRX-109 in conventional chondrosarcoma (NCT03715933) (Table 1).

Finally, IGM-8444 is an IgG DR5 antibody. With 10 binding units, it is capable of binding multiple receptors at once, facilitating receptor clustering [62]. IGM-8444 was found to induce apoptosis in a broad panel of solid and hematological malignancies [62]. A phase I study is currently recruiting patients to test IGM-8444 alone and in combination in subjects with cancer (NCT04553692).

Combination therapies

Second-generation approaches have clearly improved upon first-generation therapeutics, with larger, multivalent molecules enhancing the pharmacokinetic profiles and agonistic abilities of rhTRAIL and TRAIL-R mAbs. However, it is established that resistance to monotherapy with these TRAs is inherent or acquired [9,38,63,64]. Combinations of TRAs with chemotherapeutics and TRAIL-sensitizers are being explored to help overcome this resistance.

TRAs have been tested in combination with chemotherapeutics including gemcitabine, cisplatin, carboplatin, irinotecan, 5-FU, oxaliplatin, paclitaxel, capecitabine and doxorubicin. In the case of rhTRAIL, in a phase 1B study of patients with CRC, dulanermin was evaluated in combination with FOLFOX6 regimen plus bevacizumab. Study results showed that the combination was well-tolerated. A partial response (PR) was observed in 57% of patients, stable disease was observed in 30% of patients, and progressive disease (PD) was observed in the remaining 13% [65]. Dulanermin was also evaluated in combination with paclitaxel, carboplatin and bevacizumab in a randomized phase 2 study in patients with advanced non-small-cell lung cancer (NSCLC). The addition of dulanermin to these therapeutic regimes was tolerated but did not significantly improve patient outcomes [66]. In a phase 3 study in patients with advanced NSCLC, the combination of dulanermin plus vinorelbine and cisplatin was evaluated. Here, researchers noted that the addition of dulanermin to the vinorelbine and cisplatin regime significantly improved overall response rate (ORR) and progression-free survival (PFS) [67]. Finally, in a phase 1b open-label study, a patient with BRAF-mutant metastatic colon cancer received dulanermin and FOLFIRI. The patient maintained stable disease through 25 doses administered every two weeks, a period which extended beyond the median overall survival (OS) for patients with this disease [68]. An ongoing phase 1 trial with Eftozanermin alfa is investigating the combination with FOLFIRI and bevacizumab for treatment of patients with KRAS-mutant CRC (NCT03082209).

In the case of TRAIL-R antibodies, in a phase 2 study in chemotherapy-naïve patients with unresectable or metastatic pancreatic cancer, the combination of tigatuzumab with gemcitabine was well-tolerated. Here, an ORR of 13.1% was reported while 45.9% of patients had stable disease and 23% of patients had progressive disease [71]. LBY135 has been evaluated in combination with capecitabine in a phase 1 clinical trial of patients with advanced solid tumors. Here, there were some signs of clinical activity and the mAb was shown to be well-



tolerated, once again [72]. Numerous mechanisms have been proposed to explain chemotherapy-induced TRA-sensitization, including receptor up-regulation and suppression of anti-apoptotic proteins [38]. Nonetheless, this sensitization remains limited due to the previously mentioned agonistic and pharmacokinetic issues associated with rhTRAIL and TRAIL-R mAbs.

We discovered a potent synergy between rhTRAIL and sorafenib through effects on NF-kB and Mcl-1 [69]. This strategy was pursued in the clinic through a randomized phase II study using mapatumumab in combination with sorafenib in hepatocellular cancer, but with limited efficacy [70].

TRAs have also been tested in combination with apoptotic pathway agents that act as TRAIL-sensitizers. The rationale is that cancer cells often become resistant to TRAIL therapy through cross-talk between the extrinsic and intrinsic pathways and by up-regulating anti-apoptotic proteins [73]. For instance, c-FLIP is known to inhibit DISC formation by competing with caspase-8/-10 for binding to FADD. FLIP is often overexpressed in cancer [19], leading to the development of FLIP inhibitors and siRNAs that silence the protein [74]. Such approaches were shown to sensitize cancer cells to TRAIL therapy [75]. In addition, XIAP interacts with and inhibits caspases, and its high expression is associated with TRAIL resistance. Smac mimetics have been developed to mimic the binding site of the XIAP antagonist Smac and have shown promising preclinical activity [76]. siRNA silencing of XIAP has been shown to effectively sensitize cells to TRAIL [77]. Finally, overexpression of BCL-2 family anti-apoptotic proteins, including BCL-2, Mcl-1, and Bcl-xl, are associated with TRAIL resistance [78]. BH3-mimetics such as ABT-263, ABT-199 and ABT-737 bind to and inhibit one or more of these proteins and have also been shown to sensitize cells to TRAIL therapy [54,79,80].

Moving away from TRAIL-R agonists: an alternative approach

Regardless of the improvements made by combination therapies, resistance continues to be a limiting factor for some receptor agonist-based therapies, as cancer cells often up-regulate anti-apoptotic proteins and down-regulate TRAIL and its receptors [9]. Thus, agents that increase the expression of TRAIL and its receptors have some therapeutic potential. One such agent is TIC10/ONC201 [81].

Originally discovered as TRAIL-inducing compound 10 (TIC10), ONC201 is a first-in-class molecule and a founding member of the imipridone small molecule family. ONC201 acts directly to agonize ClpP, which leads to the activation of the integrated stress response (ISR) and downstream ATF4 and CHOP-mediated up-regulation of DR5 [82]. Earlier work has shown that ONC201 also inhibits Akt and ERK, which allows for the nuclear translocation of Foxo3a and subsequent up-regulation of TRAIL [83]. Thus, ONC201 up-regulates TRAIL in all cells and DR5 selectively in tumor cells. This up-regulation has been shown to translate to increased expression of TRAIL and DR5 at the cell surface which leads to downstream induction of apoptosis [84,85]. In addition, ONC201 inhibits cancer stem cells and has documented immunostimulatory effects [86,87]. Thus, ONC201 induces tumor-selective apoptosis and promotes immune function through several mechanisms, many of which are still being studied.

ONC201 has shown preclinical activity as a single agent against a wide variety of solid and hematological [88] tumors, including lung [89], breast [90], pancreatic [91], ovarian [92], colorectal [81], prostate [81], hepatocellular [93], leukemia [94] and lymphoma [95]. Of note, ONC201 has shown efficacy against neuroendocrine tumors and brain tumors [96,97]. Such activity can be attributed to ONC201's ability to cross the blood-brain barrier as well as its ability to antagonize DRD2/3. Indeed, ONC201's activity against neuroendocrine tumors including pheochromocytomas and paragangliomas is correlated with their high expression of dopamine receptor type II [97]. Treatment with ONC201 reduced tumor growth in multiple glioma xenograft models, and a single dose doubled the overall survival of mice with an intracranial xenograft of human GBM [81,98]. ONC201 has also shown preclinical anti-metastatic activity in various tissue types, including breast, CRC, and endometrial cancers [99]. This is explained by the fact that ONC201 inhibits cell adhesion, migration, and invasion in a TRAIL-dependent manner [99,100].

The preclinical outcomes of ONC201 led to its approval by the FDA for testing in phase 1 trials for patients with various cancers. ONC201 has been evaluated in 16 clinical trials as a single agent (Table 1). The first-in-human study found ONC201 was well-tolerated in patients with advanced solid tumors. Although objective responses were not achieved by RECIST criteria, a subset of the patients experienced tumor regression, including one patient with chemotherapy-resistant endometrial cancer [101]. More recently, a phase 2 study of ONC201 monotherapy in patients with recurrent GBM once again found that ONC201 was well-



tolerated. Of note, this study was the second report of a patient with a H3K27M mutation achieving a complete regression of enhancing multifocal lesions that was maintained for >1.5 years. It is noteworthy that the more potent ONC201 analog ONC212 is active against H3K27M-mutated DIPG cells in preclinical studies, but this analog has not been reported to antagonize dopamine receptors [102].

Although ONC201 has shown some success as a single agent, its activity is limited by resistance-inducing factors such as PI3K/AKT/mTOR upregulation [103] and insufficient TRAIL induction [104,105]. This, along with its multifaceted mechanism of action makes it a good candidate for combination therapy. There is extensive preclinical work showing synergism between ONC201 and radiation [106], chemotherapeutics, targeted therapies [107], and immune-checkpoint agents [96,99]. Most relevant to this review is the combination between ONC201 and TRAs. ONC201 was shown to be ineffective as a single agent in triple-negative breast cancer (TNBC) due to insufficient TRAIL induction [104,105]. TRAIL itself has activity only in a subset of TNBC patients. Combination therapy between ONC201 and rhTRAIL produced a potent apoptotic effect in breast cancer cells which was replicated in other cancer types [104]. A similar effect was seen in non-TNBC cells with the combination between lexatumumab and ONC201 [108]. Finally, the combination of ONC201 with TLY012 was shown to produce a synergistic apoptotic effect both *in vitro* and *in vivo* in pancreatic cancer models [109]. Such combinations have potential but remain to be tested in clinical trials. Meanwhile, clinical trials have been initiated using combinations between ONC201 and radiation (NCT05009992) or chemotherapeutics (NCT04055649) (Table 1). Preclinical data demonstrate synergy between ONC201 or other imipridones and EZH2 or HDAC inhibitors and this has been correlated with H3K27 acetylation [102,110].

Conclusion

Unlike other members of the TNF-family, TRAIL possesses the ability to selectively target cancer cells. This exciting realization led to several decades of intense research focused on the development of TRAIL receptorbased therapies. However, clinical studies have demonstrated that such therapies have failed to meet their expectations, with many possessing an inability to induce lower- or higher-order receptor clustering, poor pharmacokinetic profiles including short half-lives, and rapidly built resistance. While novel TRAs are being developed to specifically target these shortcomings and have shown promise in the clinic, it has become clear that a more imaginative approach must be taken to advance the field. The discovery of ONC201/TIC10 provides such an approach. Unlike TRAs, ONC201 acts independently of receptor binding and instead induces TRAIL and DR5 at the transcriptional level. This unique mechanism of action represents a new generation of TRAIL-pathway based approaches. ONC201 has shown promising preclinical and clinical results. Nevertheless, it must be recognized that the nature of cancer is complex, with tumor heterogeneity, evolution, and resistance to anti-cancer therapy acting as major limiting factors to single-drug therapies. Thus, further exploration of ONC201 and other TRAIL-based approaches is needed to address such obstacles.

Perspectives

- The TRAIL pathway is a powerful innate immune system whose therapeutic potential is yet to be realized.
- It is likely that harnessing the TRAIL pathway in cancer therapy alone or in combination with other cancer therapeutic modalities holds promise.
- Mechanistic studies with first- and second-generation TRAIL receptor agonists will help inform translational directions and combinatorial cancer therapeutics.

Competing Interests

W.S.E.-D. is a co-founder of Oncoceutics, Inc., a subsidiary of Chimerix. Dr. El-Deiry has disclosed his relationship with Oncoceutics/Chimerix and potential conflict of interest to his academic institution/employer and is fully compliant with NIH and institutional policy that is managing this potential conflict of interest.



Acknowledgements

W.S.E-D. is an American Cancer Society Research Professor and is supported by the Mencoff Family University Professorship at Brown University. This work was supported by an NIH grant (CA173453) and by a sponsored research agreement from D&D Pharmatech to W.S.E-D.

Abbreviations

CRC, colorectal cancer; DISC, death-inducing signaling complex; FADD, Fas-associated protein with death domain; GBM, glioblastoma; ISR, integrated stress response; IZ, isoleucine zipper; LZ, leucine zipper; NSCLC, non-small-cell lung cancer; ODD, Orphan Drug Designation; ORR, overall response rate; PEG, polyethylene glycol; RECIST, Response Evaluation Criteria in Solid Tumors; TIC10, TRAIL-inducing compound 10; TNBC, triple-negative breast cancer; TNC, tenascin-C; TRAIL, tumor necrosis factor-related apoptosis-inducing ligand; TRAs, TRAIL receptor agonists; rhTRAIL, recombinant human TRAIL.

References

- 1 Hanahan, D. and Weinberg, R.A. (2000) The hallmarks of cancer. Cell 100, 57–70 https://doi.org/10.1016/s0092-8674(00)81683-9
- 2 Carneiro, B.A. and El-Deiry, W.S. (2020) Targeting apoptosis in cancer therapy. Nat. Rev. Clin. Oncol. 17, 395–417 https://doi.org/10.1038/ s41571-020-0341-y
- Kerr, J.F., Wyllie, A.H. and Currie, A.R. (1972) Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. Br. J. Cancer 26, 239–257 https://doi.org/10.1038/bjc.1972.33
- 4 Igney, F.H. and Krammer, P.H. (2002) Death and anti-death: tumour resistance to apoptosis. Nat. Rev. Cancer 2, 277–288 https://doi.org/10.1038/ nrc776
- 5 Elmore, S. (2007) Apoptosis: a review of programmed cell death. Toxicol. Pathol. 35, 495–516 https://doi.org/10.1080/01926230701320337
- 6 Walczak, H., Miller, R.E., Ariail, K., Gliniak, B., Griffith, T.S., Kubin, M. et al. (1999) Tumoricidal activity of tumor necrosis factor-related apoptosis-inducing ligand in vivo. *Nat. Med.* **5**, 157–163 https://doi.org/10.1038/5517
- 7 Pitti, R.M., Marsters, S.A., Ruppert, S., Donahue, C.J., Moore, A., and Ashkenazi, A. (1996) Induction of apoptosis by Apo-2 ligand, a new member of the tumor necrosis factor cytokine family. *J. Biol. Chem.* **271**, 12687–12690 https://doi.org/10.1074/jbc.271.22.12687
- 8 Wu, G.S., Burns, T.F., McDonald, E.R., 3rd, Jiang, W., Meng, R., Krantz, I.D. et al. (1997) KILLER/DR5 is a DNA damage-inducible p53-regulated death receptor gene. *Nat. Genet.* **17**, 141–143 https://doi.org/10.1038/ng1097-141
- 9 Yuan, X., Gajan, A., Chu, Q., Xiong, H., Wu, K. and Wu, G.S. (2018) Developing TRAIL/TRAIL death receptor-based cancer therapies. *Cancer Metastasis Rev.* 37, 733–748 https://doi.org/10.1007/s10555-018-9728-y
- 10 Wang, S. and El-Deiry, W.S. (2003) TRAIL and apoptosis induction by TNF-family death receptors. Oncogene 22, 8628-8633 https://doi.org/10.1038/ sj.onc.1207232
- 11 Meng, R.D., McDonald, III, E.R., Sheikh, M.S., Fornace, Jr, A.J. and El-Deiry, W.S. (2000) The TRAIL decoy receptor TRUNDD (DcR2, TRAIL-R4) is induced by adenovirus-p53 overexpression and can delay TRAIL-, p53-, and KILLER/DR5-dependent colon cancer apoptosis. *Mol. Ther.* **1**, 130–144 https://doi.org/10.1006/mthe.2000.0025
- 12 Pan, G., Ni, J., Wei, Y.F., Yu, G., Gentz, R. and Dixit, V.M. (1997) An antagonist decoy receptor and a death domain-containing receptor for TRAIL. *Science* **277**, 815–818 https://doi.org/10.1126/science.277.5327.815
- 13 Sanlioglu, A.D., Dirice, E., Aydin, C., Erin, N., Koksoy, S. and Sanlioglu, S. (2005) Surface TRAIL decoy receptor-4 expression is correlated with TRAIL resistance in MCF7 breast cancer cells. *BMC Cancer* 5, 54 https://doi.org/10.1186/1471-2407-5-54
- 14 Wu, G.S. (2009) TRAIL as a target in anti-cancer therapy. Cancer Lett. 285, 1–5 https://doi.org/10.1016/j.canlet.2009.02.029
- 15 Naval, J., de Miguel, D., Gallego-Lleyda, A., Anel, A. and Martinez-Lostao, L. (2019) Importance of TRAIL molecular anatomy in receptor oligomerization and signaling. implications for cancer therapy. *Cancers (Basel)* **11**, 444 https://doi.org/10.3390/cancers11040444
- Huang, K., Zhang, J., O'Neill, K.L., Gurumurthy, C.B., Quadros, R.M., Tu, Y. et al. (2016) Cleavage by caspase 8 and mitochondrial membrane association activate the BH3-only protein bid during TRAIL-induced apoptosis. J. Biol. Chem. 291, 11843–11851 https://doi.org/10.1074/jbc.M115. 711051
- 17 Ozoren, N. and El-Deiry, W.S. (2002) Defining characteristics of types I and II apoptotic cells in response to TRAIL. *Neoplasia* **4**, 551–557 https://doi. org/10.1038/sj.neo.7900270
- 18 Werneburg, N.W., Bronk, S.F., Guicciardi, M.E., Thomas, L., Dikeakos, J.D., Thomas, G. et al. (2012) Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) protein-induced lysosomal translocation of proapoptotic effectors is mediated by phosphofurin acidic cluster sorting protein-2 (PACS-2). J. Biol. Chem. 287, 24427–24437 https://doi.org/10.1074/jbc.M112.342238
- 19 Ricci, M.S., Jin, Z., Dews, M., Yu, D., Thomas-Tikhonenko, A., Dicker, D.T. et al. (2004) Direct repression of FLIP expression by c-myc is a major determinant of TRAIL sensitivity. *Mol. Cell. Biol.* 24, 8541–8555 https://doi.org/10.1128/MCB.24.19.8541-8555.2004
- 20 Kuijlen, J.M., Mooij, J.J., Platteel, I., Hoving, E.W., van der Graaf, W.T., Span, M.M. et al. (2006) TRAIL-receptor expression is an independent prognostic factor for survival in patients with a primary glioblastoma multiforme. J. Neurooncol. 78, 161–171 https://doi.org/10.1007/ s11060-005-9081-1
- 21 Finnberg, N., Klein-Szanto, A.J. and El-Deiry, W.S. (2008) TRAIL-R deficiency in mice promotes susceptibility to chronic inflammation and tumorigenesis. J. Clin. Invest. 118, 111–123 https://doi.org/10.1172/JCl29900
- 22 Herbst, R.S., Eckhardt, S.G., Kurzrock, R., Ebbinghaus, S., O'Dwyer, P.J., Gordon, M.S. et al. (2010) Phase I dose-escalation study of recombinant human Apo2L/TRAIL, a dual proapoptotic receptor agonist, in patients with advanced cancer. J. Clin. Oncol. 28, 2839–2846 https://doi.org/10.1200/ JC0.2009.25.1991



- 23 Soria, J.C., Smit, E., Khayat, D., Besse, B., Yang, X., Hsu, C.P. et al. (2010) Phase 1b study of dulanermin (recombinant human Apo2L/TRAIL) in combination with paclitaxel, carboplatin, and bevacizumab in patients with advanced non-squamous non-small-cell lung cancer. J. Clin. Oncol. 28, 1527–1533 https://doi.org/10.1200/JC0.2009.25.4847
- 24 Graves, J.D., Kordich, J.J., Huang, T.H., Piasecki, J., Bush, T.L., Sullivan, T. et al. (2014) Apo2I/TRAIL and the death receptor 5 agonist antibody AMG 655 cooperate to promote receptor clustering and antitumor activity. *Cancer Cell* **26**, 177–189 https://doi.org/10.1016/j.ccr.2014.04.028
- 25 von Karstedt, S., Montinaro, A. and Walczak, H. (2017) Exploring the TRAILs less travelled: TRAIL in cancer biology and therapy. Nat. Rev. Cancer 17, 352–366 https://doi.org/10.1038/nrc.2017.28
- 26 Adams, C., Totpal, K., Lawrence, D., Marsters, S., Pitti, R., Yee, S. et al. (2008) Structural and functional analysis of the interaction between the agonistic monoclonal antibody Apomab and the proapoptotic receptor DR5. *Cell Death Differ*. **15**, 751–761 https://doi.org/10.1038/sj.cdd.4402306
- 27 Plummer, R., Attard, G., Pacey, S., Li, L., Razak, A., Perrett, R. et al. (2007) Phase 1 and pharmacokinetic study of lexatumumab in patients with advanced cancers. *Clin. Cancer Res.* **13**, 6187–6194 https://doi.org/10.1158/1078-0432.CCR-07-0950
- Herbst, R.S., Kurzrock, R., Hong, D.S., Valdivieso, M., Hsu, C.P., Goyal, L. et al. (2010) A first-in-human study of conatumumab in adult patients with advanced solid tumors. *Clin. Cancer Res.* 16, 5883–5891 https://doi.org/10.1158/1078-0432.CCR-10-0631
- 29 Camidge, D.R., Herbst, R.S., Gordon, M.S., Eckhardt, S.G., Kurzrock, R., Durbin, B. et al. (2010) A phase I safety and pharmacokinetic study of the death receptor 5 agonistic antibody PR095780 in patients with advanced malignancies. *Clin. Cancer Res.* 16, 1256–1263 https://doi.org/10.1158/ 1078-0432.CCR-09-1267
- 30 Forero-Torres, A., Shah, J., Wood, T., Posey, J., Carlisle, R., Copigneaux, C. et al. (2010) Phase I trial of weekly tigatuzumab, an agonistic humanized monoclonal antibody targeting death receptor 5 (DR5). *Cancer Biother. Radiopharm.* 25, 13–19 https://doi.org/10.1089/cbr.2009.0673
- 31 Hotte, S.J., Hirte, H.W., Chen, E.X., Siu, L.L., Le, L.H., Corey, A. et al. (2008) A phase 1 study of mapatumumab (fully human monoclonal antibody to TRAIL-R1) in patients with advanced solid malignancies. *Clin. Cancer Res.* **14**, 3450–3455 https://doi.org/10.1158/1078-0432.CCR-07-1416
- 32 Allen, J.E., Ferrini, R., Dicker, D.T., Batzer, G., Chen, E., Oltean, D.I. et al. (2012) Targeting TRAIL death receptor 4 with trivalent DR4 Atrimer complexes. *Mol. Cancer Ther.* **11**, 2087–2095 https://doi.org/10.1158/1535-7163.Mct-12-0366
- 33 Gieffers, C., Kluge, M., Merz, C., Sykora, J., Thiemann, M., Schaal, R. et al. (2013) APG350 induces superior clustering of TRALL receptors and shows therapeutic antitumor efficacy independent of cross-linking via fcgamma receptors. *Mol. Cancer Ther.* **12**, 2735–2747 https://doi.org/10.1158/ 1535-7163.MCT-13-0323
- 34 Thapa, B., Kc, R. and Uludag, H. (2020) TRAIL therapy and prospective developments for cancer treatment. J. Control. Release **326**, 335–349 https://doi.org/10.1016/j.jconrel.2020.07.013
- 35 Ashkenazi, A., Pai, R.C., Fong, S., Leung, S., Lawrence, D.A., Marsters, S.A. et al. (1999) Safety and antitumor activity of recombinant soluble Apo2 ligand. J. Clin. Invest. 104, 155–162 https://doi.org/10.1172/JCI6926
- 36 Pollack, I.F., Erff, M. and Ashkenazi, A. (2001) Direct stimulation of apoptotic signaling by soluble Apo2l/tumor necrosis factor-related apoptosis-inducing ligand leads to selective killing of glioma cells. *Clin. Cancer Res.* **7**, 1362–1369
- 37 Tolcher, A.W., Mita, M., Meropol, N.J., von Mehren, M., Patnaik, A., Padavic, K. et al. (2007) Phase I pharmacokinetic and biologic correlative study of mapatumumab, a fully human monoclonal antibody with agonist activity to tumor necrosis factor-related apoptosis-inducing ligand receptor-1. *J. Clin.* Oncol. 25, 1390–1395 https://doi.org/10.1200/JC0.2006.08.8898
- 38 Lemke, J., von Karstedt, S., Zinngrebe, J. and Walczak, H. (2014) Getting TRAIL back on track for cancer therapy. Cell Death Differ. 21, 1350–1364 https://doi.org/10.1038/cdd.2014.81
- 39 Liu, H., Su, D., Zhang, J., Ge, S., Li, Y., Wang, F. et al. (2017) Improvement of pharmacokinetic profile of TRAIL via trimer-tag enhances its antitumor activity in vivo. Sci. Rep. 7, 8953 https://doi.org/10.1038/s41598-017-09518-1
- 40 de Miguel, D., Lemke, J., Anel, A., Walczak, H. and Martinez-Lostao, L. (2016) Onto better TRAILs for cancer treatment. *Cell Death Differ.* 23, 733–747 https://doi.org/10.1038/cdd.2015.174
- 41 Ganten, T.M., Koschny, R., Sykora, J., Schulze-Bergkamen, H., Büchler, P., Haas, T.L. et al. (2006) Preclinical differentiation between apparently safe and potentially hepatotoxic applications of TRAIL either alone or in combination with chemotherapeutic drugs. *Clin. Cancer Res.* 12, 2640–2646 https://doi.org/10.1158/1078-0432.CCR-05-2635
- 42 Berg, D., Lehne, M., Müller, N., Siegmund, D., Münkel, S., Sebald, W. et al. (2007) Enforced covalent trimerization increases the activity of the TNF ligand family members TRAIL and CD95L. *Cell Death Differ.* **14**, 2021–2034 https://doi.org/10.1038/sj.cdd.4402213
- 43 Jo, M., Kim, T.H., Seol, D.W., Esplen, J.E., Dorko, K., Billiar, T.R. et al. (2000) Apoptosis induced in normal human hepatocytes by tumor necrosis factor-related apoptosis-inducing ligand. *Nat. Med.* **6**, 564–567 https://doi.org/10.1038/75045
- 44 Harris, J.M. and Chess, R.B. (2003) Effect of pegylation on pharmaceuticals. Nat. Rev. Drug Discov. 2, 214–221 https://doi.org/10.1038/nrd1033
- 45 Crawford, J. (2002) Clinical benefits of pegylated proteins in oncology. Cancer Treat. Rev. 28, 1–2 https://doi.org/10.1016/s0305-7372(02)80001-9
- 46 Chae, S.Y., Kim, T.H., Park, K., Jin, C.H., Son, S., Lee, S. et al. (2010) Improved antitumor activity and tumor targeting of NH(2)-terminal-specific PEGylated tumor necrosis factor-related apoptosis-inducing ligand. *Mol. Cancer Ther.* 9, 1719–1729 https://doi.org/10.1158/1535-7163.MCT-09-1076
- Kim, T.H., Youn, Y.S., Jiang, H.H., Lee, S., Chen, X. and Lee, K.C. (2011) PEGylated TNF-related apoptosis-inducing ligand (TRAIL) analogues: pharmacokinetics and antitumor effects. *Bioconjug. Chem.* 22, 1631–1637 https://doi.org/10.1021/bc200187k
- 48 Park, J.S., Oh, Y., Park, Y.J., Park, O., Yang, H., Slania, S. et al. (2019) Targeting of dermal myofibroblasts through death receptor 5 arrests fibrosis in mouse models of scleroderma. *Nat. Commun.* **10**, 1128 https://doi.org/10.1038/s41467-019-09101-4
- 49 Oh, Y., Park, O., Swierczewska, M., Hamilton, J.P., Park, J.S., Kim, T.H. et al. (2016) Systemic PEGylated TRAIL treatment ameliorates liver cirrhosis in rats by eliminating activated hepatic stellate cells. *Hepatology* **64**, 209–223 https://doi.org/10.1002/hep.28432
- 50 Huang, M., Zhu, H., Yi, C., Yan, J., Wei, L., Yang, X. et al. (2018) A novel TRAIL mutant-TRAIL-Mu3 enhances the antitumor effects by the increased affinity and the up-expression of DR5 in pancreatic cancer. *Cancer Chemother. Pharmacol.* **82**, 829–838 https://doi.org/10.1007/s00280-018-3658-9
- 51 Huang, M., Yi, C., Huang, X.Z., Yan, J., Wei, L.J., Tang, W.J. et al. (2021) Recombinant protein TRAIL-Mu3 enhances the antitumor effects in pancreatic cancer cells by strengthening the apoptotic signaling pathway. *Oncol. Lett.* **21**, 438 https://doi.org/10.3892/ol.2021.12699
- 52 Wilson, N.S., Yang, B., Yang, A., Loeser, S., Marsters, S., Lawrence, D. et al. (2011) An fcgamma receptor-dependent mechanism drives antibody-mediated target-receptor signaling in cancer cells. *Cancer Cell* **19**, 101–113 https://doi.org/10.1016/j.ccr.2010.11.012



- 53 Legler, K., Hauser, C., Egberts, J.H., Willms, A., Heneweer, C., Boretius, S. et al. (2018) The novel TRAIL-receptor agonist APG350 exerts superior therapeutic activity in pancreatic cancer cells. *Cell Death Dis.* **9**, 445 https://doi.org/10.1038/s41419-018-0478-0
- 54 Phillips, D.C., Buchanan, F.G., Cheng, D., Solomon, L.R., Xiao, Y., Xue, J. et al. (2021) Hexavalent TRAIL fusion protein eftozanermin alfa optimally clusters apoptosis-inducing TRAIL receptors to induce on-target antitumor activity in solid tumors. *Cancer Res.* **81**, 3402–3414 https://doi.org/10.1158/0008-5472.CAN-20-2178
- 55 LoRusso, P., Ratain, M.J., Doi, T., Rasco, D.W., de Jonge, M.J.A., Moreno, V. et al. (2022) Eftozanermin alfa (ABBV-621) monotherapy in patients with previously treated solid tumors: findings of a phase 1, first-in-human study. *Invest. New Drugs* 40, 762–772 https://doi.org/10.1007/ s10637-022-01247-1
- 56 de Jonge, M.J.A., Carneiro, B.A., Devriese, L., Doi, T., Penugonda, S., Petrich, A.M. et al. (2019) First-in-Human study of abbv-621, a TRAIL receptor agonist fusion protein, in patients (Pts) with relapsed/refractory (RR) acute myeloid leukemia (AML) and diffuse large B-cell lymphoma (DLBCL). *Blood* **134**, 3924–3924 https://doi.org/10.1182/blood-2019-129783
- 57 Cook, E.M., Lindorfer, M.A., van der Horst, H., Oostindie, S., Beurskens, F.J., Schuurman, J. et al. (2016) Antibodies that efficiently form hexamers upon antigen binding Can induce complement-dependent cytotoxicity under complement-Limiting conditions. *J. Immunol.* **197**, 1762–1775 https://doi. org/10.4049/jimmunol.1600648
- 58 de Jong, R.N., Beurskens, F.J., Verploegen, S., Strumane, K., van Kampen, M.D., Voorhorst, M. et al. (2016) A novel platform for the potentiation of therapeutic antibodies based on antigen-dependent formation of IgG hexamers at the cell surface. *PLoS Biol.* 14, e1002344 https://doi.org/10.1371/ journal.pbio.1002344
- 59 Overdijk, M.B., Strumane, K., Beurskens, F.J., Ortiz Buijsse, A., Vermot-Desroches, C., Vuillermoz, B.S. et al. (2020) Dual epitope targeting and enhanced hexamerization by DR5 antibodies as a novel approach to induce potent antitumor activity through DR5 agonism. *Mol. Cancer Ther.* **19**, 2126–2138 https://doi.org/10.1158/1535-7163.MCT-20-0044
- 60 van der Horst, H.J., Gelderloos, A.T., Chamuleau, M.E.D., Breij, E.C.W., Zweegman, S., Nijhof, I.S. et al. (2021) Potent preclinical activity of HexaBody-DR5/DR5 in relapsed and/or refractory multiple myeloma. *Blood Adv.* 5, 2165–2172 https://doi.org/10.1182/bloodadvances.2020003731
- 61 Papadopoulos, K.P., Isaacs, R., Bilic, S., Kentsch, K., Huet, H.A., Hofmann, M. et al. (2015) Unexpected hepatotoxicity in a phase I study of TAS266, a novel tetravalent agonistic Nanobody(R) targeting the DR5 receptor. *Cancer Chemother. Pharmacol.* **75**, 887–895 https://doi.org/10.1007/s00280-015-2712-0
- 62 Wang, B.T., Kothambawala, T., Wang, L., Matthew, T.J., Calhoun, S.E., Saini, A.K. et al. (2021) Multimeric anti-DR5 IgM agonist antibody IGM-8444 is a potent inducer of cancer cell apoptosis and synergizes with chemotherapy and BCL-2 inhibitor ABT-199. *Mol. Cancer Ther.* 20, 2483–2494 https://doi.org/10.1158/1535-7163.MCT-20-1132
- 63 Dianat-Moghadam, H., Heidarifard, M., Mahari, A., Shahgolzari, M., Keshavarz, M., Nouri, M. et al. (2020) TRAIL in oncology: from recombinant TRAIL to nano- and self-targeted TRAIL-based therapies. *Pharmacol. Res.* **155**, 104716 https://doi.org/10.1016/j.phrs.2020.104716
- 64 Wong, S.H.M., Kong, W.Y., Fang, C.M., Loh, H.S., Chuah, L.H., Abdullah, S. et al. (2019) The TRAIL to cancer therapy: hindrances and potential solutions. *Crit. Rev. Oncol. Hematol.* **143**, 81–94 https://doi.org/10.1016/j.critrevonc.2019.08.008
- 65 Wainberg, Z.A., Messersmith, W.A., Peddi, P.F., Kapp, A.V., Ashkenazi, A., Royer-Joo, S. et al. (2013) A phase 1B study of dulanermin in combination with modified FOLFOX6 plus bevacizumab in patients with metastatic colorectal cancer. *Clin. Colorectal Cancer* **12**, 248–254 https://doi.org/10.1016/j. clcc.2013.06.002
- 66 Soria, J.C., Márk, Z., Zatloukal, P., Szima, B., Albert, I., Juhász, E. et al. (2011) Randomized phase II study of dulanermin in combination with paclitaxel, carboplatin, and bevacizumab in advanced non-small-cell lung cancer. J. Clin. Oncol. 29, 4442–4451 https://doi.org/10.1200/JC0.2011.37.2623
- 67 Ouyang, X., Shi, M., Jie, F., Bai, Y., Shen, P., Yu, Z. et al. (2018) Phase III study of dulanermin (recombinant human tumor necrosis factor-related apoptosis-inducing ligand/Apo2 ligand) combined with vinorelbine and cisplatin in patients with advanced non-small-cell lung cancer. *Invest. New Drugs* 36, 315–322 https://doi.org/10.1007/s10637-017-0536-y
- 68 Lim, B., Scicchitano, A., Beachler, C., Gusani, N., Sarwani, N., Yang, Z. et al. (2013) FOLFIRI plus dulanermin (rhApo2L/TRAIL) in a patient with BRAF-mutant metastatic colon cancer. *Cancer Biol. Ther.* 14, 711–719 https://doi.org/10.4161/cbt.25310
- 69 Ricci, M.S., Kim, S.H., Ogi, K., Plastaras, J.P., Ling, J., Wang, W. et al. (2007) Reduction of TRAIL-induced Mcl-1 and cIAP2 by c-Myc or sorafenib sensitizes resistant human cancer cells to TRAIL-induced death. *Cancer Cell* **12**, 66–80 https://doi.org/10.1016/j.ccr.2007.05.006
- 70 Ciuleanu, T., Bazin, I., Lungulescu, D., Miron, L., Bondarenko, I., Deptala, A. et al. (2016) A randomized, double-blind, placebo-controlled phase II study to assess the efficacy and safety of mapatumumab with sorafenib in patients with advanced hepatocellular carcinoma. *Ann. Oncol.* 27, 680–687 https://doi.org/10.1093/annonc/mdw004
- 71 Forero-Torres, A., Infante, J.R., Waterhouse, D., Wong, L., Vickers, S., Arrowsmith, E. et al. (2013) Phase 2, multicenter, open-label study of tigatuzumab (CS-1008), a humanized monoclonal antibody targeting death receptor 5, in combination with gemcitabine in chemotherapy-naive patients with unresectable or metastatic pancreatic cancer. *Cancer Med.* **2**, 925–932 https://doi.org/10.1002/cam4.137
- 72 Sharma, S., de Vries, E.G., Infante, J.R., Oldenhuis, C., Chiang, L., Goldbrunner, S., Bilic, M. et al. (2008) Phase I trial of LBY135, a monoclonal antibody agonist to DR5, alone and in combination with capecitabine in advanced solid tumors. *J. Clin. Oncol.* **26**, 3538–3538 https://doi.org/10.1200/jco.2008.26.15_suppl.3538
- 73 Allen, J.E., Prabhu, V.V., Talekar, M., van den Heuvel, A.P., Lim, B., Dicker, D.T. et al. (2015) Genetic and pharmacological screens converge in identifying FLIP, BCL2, and IAP proteins as key regulators of sensitivity to the TRAIL-inducing anticancer agent ONC201/TIC10. *Cancer Res.* **75**, 1668–1674 https://doi.org/10.1158/0008-5472.CAN-14-2356
- 74 Zhang, S. and El-Deiry, W.S. (2022) Abstract 5854: Therapeutic targeting of p73-activated c-FLIP switches cell fate from growth arrest and survival to apoptosis in p53-deficient cancer cells. *Cancer Res.* **82**, 5854–5854 https://doi.org/10.1158/1538-7445.Am2022-5854
- 75 Chawla-Sarkar, M., Bae, S.I., Reu, F.J., Jacobs, B.S., Lindner, D.J. and Borden, E.C. (2004) Down-regulation of Bcl-2, FLIP or IAPs (XIAP and survivin) by siRNAs sensitizes resistant melanoma cells to Apo2L/TRAIL-induced apoptosis. *Cell Death Differ.* **11**, 915–923 https://doi.org/10.1038/sj.cdd. 4401416
- 76 Fulda, S., Wick, W., Weller, M. and Debatin, K.M. (2002) Smac agonists sensitize for Apo2L/TRAIL- or anticancer drug-induced apoptosis and induce regression of malignant glioma *in vivo. Nat. Med.* 8, 808–815 https://doi.org/10.1038/nm735



- 77 Spee, B., Jonkers, M.D., Arends, B., Rutteman, G.R., Rothuizen, J. and Penning, L.C. (2006) Specific down-regulation of XIAP with RNA interference enhances the sensitivity of canine tumor cell-lines to TRAIL and doxorubicin. *Mol. Cancer* **5**, 34 https://doi.org/10.1186/1476-4598-5-34
- 78 Kim, K., Nakagawa, H., Fei, P., Rustgi, A.K. and El-Deiry, W.S. (2004) Targeting Bcl-xL in esophageal squamous cancer to sensitize to chemotherapy plus TRAIL-induced apoptosis while normal epithelial cells are protected by blockade of caspase 9. *Cell Death Differ.* **11**, 583–587 https://doi.org/10. 1038/sj.cdd.4401388
- 79 Huang, S. and Sinicrope, F.A. (2008) BH3 mimetic ABT-737 potentiates TRAIL-mediated apoptotic signaling by unsequestering Bim and Bak in human pancreatic cancer cells. *Cancer Res.* 68, 2944–2951 https://doi.org/10.1158/0008-5472.CAN-07-2508
- 80 Wang, G., Zhan, Y., Wang, H. and Li, W. (2012) ABT-263 sensitizes TRALL-resistant hepatocarcinoma cells by downregulating the Bcl-2 family of anti-apoptotic protein. *Cancer Chemother. Pharmacol.* 69, 799–805 https://doi.org/10.1007/s00280-011-1763-0
- 81 Allen, J.E., Krigsfeld, G., Mayes, P.A., Patel, L., Dicker, D.T., Patel, A.S. et al. (2013) Dual inactivation of Akt and ERK by TIC10 signals Foxo3a nuclear translocation, TRAIL gene induction, and potent antitumor effects. *Sci. Transl. Med.* **5**, 171ra117 https://doi.org/10.1126/scitranslmed.3004828
- 82 Kline, C.L., Van den Heuvel, A.P., Allen, J.E., Prabhu, V.V., Dicker, D.T. and El-Deiry, W.S. (2016) ONC201 kills solid tumor cells by triggering an integrated stress response dependent on ATF4 activation by specific elF2alpha kinases. *Sci. Signal.* **9**, ra18 https://doi.org/10.1126/scisignal.aac4374
- 83 Prabhu, V.V., Allen, J.E., Dicker, D.T. and El-Deiry, W.S. (2015) Small-molecule ONC201/TIC10 targets chemotherapy-resistant colorectal cancer stem-like cells in an Akt/Foxo3a/TRAIL-dependent manner. *Cancer Res.* **75**, 1423–1432 https://doi.org/10.1158/0008-5472.CAN-13-3451
- 84 Ray, J.E., Ralff, M.D., Jhaveri, A., Zhou, L., Dicker, D.T., Ross, E.A. et al. (2021) Antitumorigenic effect of combination treatment with ONC201 and TRAIL in endometrial cancer *in vitro* and *in vivo. Cancer Biol. Ther.* **22**, 554–563 https://doi.org/10.1080/15384047.2021.1977067
- 85 Parker, C.S., Zhou, L., Prabhu, V., Allen, J. and El-Deiry, W. (2021) Abstract 1044: ONC201 as a novel anti-cancer therapeutic via modulation of inhibitors of apoptosis and up-regulation of DR5 in gastric adenocarcinoma. *Cancer Res.* 81, 1044–1044 https://doi.org/10.1158/1538-7445. Am2021-1044
- 86 Prabhu, V.V., Lulla, A.R., Madhukar, N.S., Ralff, M.D., Zhao, D., Kline, C.L. B. et al. (2017) Cancer stem cell-related gene expression as a potential biomarker of response for first-in-class imipridone ONC201 in solid tumors. *PLoS ONE* 12, e0180541 https://doi.org/10.1371/journal.pone.0180541
- 87 Stein, M.N., Malhotra, J., Tarapore, R.S., Malhotra, U., Silk, A.W., Chan, N. et al. (2019) Safety and enhanced immunostimulatory activity of the DRD2 antagonist ONC201 in advanced solid tumor patients with weekly oral administration. J. Immunother. Cancer 7, 136 https://doi.org/10.1186/ s40425-019-0599-8
- Prabhu, V.V., Talekar, M.K., Lulla, A.R., Kline, C.L.B., Zhou, L., Hall, J. et al. (2018) Single agent and synergistic combinatorial efficacy of first-in-class small molecule imipridone ONC201 in hematological malignancies. *Cell Cycle* **17**, 468–478 https://doi.org/10.1080/15384101.2017.1403689
- 89 Feng, Y., Zhou, J., Li, Z., Jiang, Y. and Zhou, Y. (2016) Small molecular TRAIL inducer ONC201 induces death in lung cancer cells: a preclinical study. PLoS ONE 11, e0162133 https://doi.org/10.1371/journal.pone.0162133
- 90 Yuan, X., Kho, D., Xu, J., Gajan, A., Wu, K. and Wu, G.S. (2017) ONC201 activates ER stress to inhibit the growth of triple-negative breast cancer cells. Oncotarget 8, 21626–21638 https://doi.org/10.18632/oncotarget.15451
- 91 Zhang, Q., Wang, H., Ran, L., Zhang, Z. and Jiang, R. (2016) The preclinical evaluation of TIC10/ONC201 as an anti-pancreatic cancer agent. Biochem. Biophys. Res. Commun. 476, 260–266 https://doi.org/10.1016/j.bbrc.2016.05.106
- 92 Baumeister, M.D., Küçükkase, O.C., Prabhu, V.V., Dicker, D.T., Allen, J.E. and El-Deiry, W.S. (2017) Abstract 3212: ONC201 shows efficacy in BRCA-deficient cancer cells and synergy with PARP inhibitors in glioblastoma, breast, prostate, and ovarian cancers. *Cancer Res.* 77, 3212–3212 https://doi.org/10.1158/1538-7445.Am2017-3212
- 93 Cheng, L., Liu, Y.Y., Lu, P.H., Peng, Y., Yuan, Q., Gu, X.S. et al. (2017) Identification of DNA-PKcs as a primary resistance factor of TIC10 in hepatocellular carcinoma cells. Oncotarget 8, 28385–28394 https://doi.org/10.18632/oncotarget.16073
- 94 Ishizawa, J., Zarabi, S.F., Davis, R.E., Halgas, O., Nii, T., Jitkova, Y. et al. (2019) Mitochondrial ClpP-mediated proteolysis induces selective cancer cell lethality. *Cancer Cell* **35**, 721–737.e729 https://doi.org/10.1016/j.ccell.2019.03.014
- 95 Talekar, M.K., Allen, J.E., Dicker, D.T. and El-Deiry, W.S. (2015) ONC201 induces cell death in pediatric non-Hodgkin's lymphoma cells. *Cell Cycle* 14, 2422–2428 https://doi.org/10.1080/15384101.2015.1054086
- 96 Prabhu, V.V., Morrow, S., Rahman Kawakibi, A., Zhou, L., Ralff, M., Ray, J. et al. (2020) ONC201 and imipridones: anti-cancer compounds with clinical efficacy. *Neoplasia* 22, 725–744 https://doi.org/10.1016/j.neo.2020.09.005
- 97 Anderson, P.M., Trucco, M.M., Tarapore, R.S., Zahler, S., Thomas, S., Gortz, J. et al. (2022) Phase II study of ONC201 in neuroendocrine tumors including pheochromocytoma-paraganglioma and desmoplastic small round cell tumor. *Clin. Cancer Res.* 28, 1773–1782 https://doi.org/10.1158/ 1078-0432.CCR-21-4030
- 98 Ishida, C.T., Zhang, Y., Bianchetti, E., Shu, C., Nguyen, T.T.T., Kleiner, G. et al. (2018) Metabolic reprogramming by dual AKT/ERK inhibition through imipridones elicits unique vulnerabilities in glioblastoma. *Clin. Cancer Res.* 24, 5392–5406 https://doi.org/10.1158/1078-0432.CCR-18-1040
- 99 Wagner, J., Kline, C.L., Zhou, L., Campbell, K.S., MacFarlane, A.W., Olszanski, A.J. et al. (2018) Dose intensification of TRAIL-inducing ONC201 inhibits metastasis and promotes intratumoral NK cell recruitment. *J. Clin. Invest.* **128**, 2325–2338 https://doi.org/10.1172/JCl96711
- 100 Fang, Z., Wang, J., Clark, L.H., Sun, W., Yin, Y., Kong, W. et al. (2018) ONC201 demonstrates anti-tumorigenic and anti-metastatic activity in uterine serous carcinoma *in vitro. Am. J. Cancer Res.* **8**, 1551–1563
- 101 Stein, M.N., Bertino, J.R., Kaufman, H.L., Mayer, T., Moss, R., Silk, A. et al. (2017) First-in-human clinical trial of oral ONC201 in patients with refractory solid tumors. *Clin. Cancer Res.* 23, 4163–4169 https://doi.org/10.1158/1078-0432.CCR-16-2658
- 102 Borsuk, R., Zhou, L., Chang, W.I., Zhang, Y., Sharma, A., Prabhu, V.V. et al. (2021) Potent preclinical sensitivity to imipridone-based combination therapies in oncohistone H3K27M-mutant diffuse intrinsic pontine glioma is associated with induction of the integrated stress response, TRAIL death receptor DR5, reduced ClpX and apoptosis. *Am. J. Cancer Res.* **11**, 4607–4623
- 103 Jin, Z.Z., Wang, W., Fang, D.L. and Jin, Y.J. (2016) mTOR inhibition sensitizes ONC201-induced anti-colorectal cancer cell activity. *Biochem. Biophys. Res. Commun.* 478, 1515–1520 https://doi.org/10.1016/j.bbrc.2016.08.126
- 104 Ralff, M.D., Kline, C.L.B., Küçükkase, O.C., Wagner, J., Lim, B., Dicker, D.T. et al. (2017) ONC201 demonstrates antitumor effects in both triple-negative and non-triple-negative breast cancers through TRAIL-dependent and TRAIL-independent mechanisms. *Mol. Cancer Ther.* 16, 1290–1298 https://doi.org/10.1158/1535-7163.MCT-17-0121

69



- 105 Ralff, M.D., Jhaveri, A., Ray, J.E., Zhou, L., Lev, A., Campbell, K.S. et al. (2020) TRAIL receptor agonists convert the response of breast cancer cells to ONC201 from anti-proliferative to apoptotic. *Oncotarget* **11**, 3753–3769 https://doi.org/10.18632/oncotarget.27773
- 106 Zhou, L., Wu, J.L., Safran, H.P. and El-Deiry, W.S. (2022) Abstract 5448: Synergistic antitumor effect of ONC201, radiotherapy and temozolomide in glioblastoma mouse orthotopic models. *Cancer Res.* 82, 5448–5448 https://doi.org/10.1158/1538-7445.Am2022-5448
- 107 Cristofano, F.R.D., Fong, M., Zhou, L. and El-Deiry, W.S. (2022) Abstract 3709: Synergistic activity of ABT-263 and ONC201 against solid tumor cell lines is associated with suppression of BAG3, Mcl-1, pAkt, and up-regulation of Noxa along with Bax cleavage during apoptosis. *Cancer Res.* 82, 3709–3709 https://doi.org/10.1158/1538-7445.Am2022-3709
- 108 Ralff, M.D., Ray, J.E., Lev, A., Zhou, L., Dicker, D.T. and El-Deiry, W.S. (2019) Abstract 258: Recombinant human TRAIL or a DR5 agonistic antibody convert the response of non-triple negative breast cancer cells to ONC201 from anti-proliferative to apoptotic. *Cancer Res.* **79**, 258–258 https://doi.org/ 10.1158/1538-7445.Am2019-258
- 109 Jhaveri, A.V., Zhou, L., Ralff, M.D., Lee, Y.S., Navaraj, A., Carneiro, B.A. et al. (2021) Combination of ONC201 and TLY012 induces selective, synergistic apoptosis in vitro and significantly delays PDAC xenograft growth *in vivo. Cancer Biol. Ther.* 22, 607–618 https://doi.org/10.1080/ 15384047.2021.1976567
- 110 Zhang, Y., Zhou, L., Safran, H., Borsuk, R., Lulla, R., Tapinos, N. et al. (2021) EZH2i EPZ-6438 and HDACi vorinostat synergize with ONC201/TIC10 to activate integrated stress response, DR5, reduce H3K27 methylation, ClpX and promote apoptosis of multiple tumor types including DIPG. *Neoplasia* 23, 792–810 https://doi.org/10.1016/j.neo.2021.06.007