Check for updates

Review Article

Our evolving understanding of the role of the $\gamma\delta$ T cell receptor in $\gamma\delta$ T cell mediated immunity

Benjamin S. Gully¹,
Jamie Rossjohn^{1,2} and
Martin S. Davey¹

¹Infection and Immunity Program and Department of Biochemistry and Molecular Biology, Biomedicine Discovery Institute, Monash University, Clayton, Victoria 3800, Australia; ²Institute of Infection and Immunity, Cardiff University School of Medicine, Heath Park, Cardiff CF14 4XN, U.K.

Correspondence: Jamie Rossjohn (Jamie.rossjohn@monash.edu)



The $\gamma\delta$ T cell immune cell lineage has remained relatively enigmatic and under-characterised since their identification. Conversely, the insights we have, highlight their central importance in diverse immunological roles and homeostasis. Thus, $\gamma\delta$ T cells are considered as potentially a new translational tool in the design of new therapeutics for cancer and infectious disease. Here we review our current understanding of $\gamma\delta$ T cell biology viewed through a structural lens centred on the how the $\gamma\delta$ T cell receptor mediates ligand recognition. We discuss the limited knowledge of antigens, the structural basis of such reactivities and discuss the emerging trends of $\gamma\delta$ T cell reactivity and implications for $\gamma\delta$ T cell biology.

Introduction

The human adaptive immune system has conserved a tripartite lymphocyte compartment comprising B cells, $\alpha\beta$ and $\gamma\delta$ T cells for over 450 million years of vertebrate evolution [1]. Whilst great strides have been made in B cell and conventional $\alpha\beta$ T cell mediated immunity, the functional role of $\gamma\delta$ T cell mediated immunity, the functional role of $\gamma\delta$ T cells

have been made in B cell and conventional αβ T cell mediated immunity, the functional role of γδ T cells have remained less well defined. Notwithstanding this, $\gamma\delta$ T cells are known to play a central, and somewhat unique, role in both anti-microbial [2] and anti-tumour immunity [3], in addition to roles in tissue homeostasis and mucosal immunity, reviewed in [4].

Current understanding of $\gamma\delta$ T cell lineages

Framed by a lack of clarity on complementary ligands $\gamma\delta$ T cells are often delineated into subsets and mucosal immunity.

based on the expressed V δ gene segment, namely V δ 1⁺, V δ 2⁺ and V δ 3⁺ subsets. The major peripheral blood subset are V δ 2⁺ $\gamma\delta$ T cells, comprising \sim 1–10% of circulating T cells [5]. These prototypic unconventional T cells almost exclusively express a Vγ9 chain, resulting in a focused peripheral repercells offer both an innate-like prenatal protection mediated by $V\delta 2^+V\gamma 9^+$ T cells that persist into adulthood alongside an adaptive-like immunobiology of $V\delta 2^+V\gamma 9^-$ T cells that clonally expand upon cytomegalovirus infection and exhibit effector function [9]. $V\delta 2^+V\gamma 9^+$ T cells that clonally expand upon cytomegalovirus infection and exhibit effector function [9]. $V\delta 2^+V\gamma 9^+$ T cells that clonally expand upon rapid anti-microbial immune response to bacteria, [10] parasites, [11] and tumour cells (Figure 1) [12,13].

 $V\delta 2^- \gamma \delta$ T cells, encompassing $V\delta 1^+$ and $V\delta 3^+$ populations, have been implicated in anti-viral immunity to cytomegalovirus [14,15] and Epstein Barr virus [16,17] as well as potential roles in tumour surveillance [18]. $V\delta 1^+$ T cells are the most abundant neonatal $\gamma\delta$ lineage and continue as the predominating subset in adult peripheral tissues including the gut, skin and liver [19-22]. Although Vδ1⁺ T cells can utilise germline encoded innate receptors, namely NKG2D, NKp30 and NKp46, [23–25] our understanding of this lineage is far from complete, with recent identification of foetal population of invariant $V\delta 1^+$ T cells that precede the development of a postnatal naïve-like and effector $V\delta 1^+$ T cell sub-compartments, dependent on T cell receptor (TCR) selection and eluding towards distinct adaptive-like properties (Figure 1) [26–29].

Received: 2 July 2021 Revised: 17 August 2021 Accepted: 19 August 2021

Version of Record published: 13 September 2021



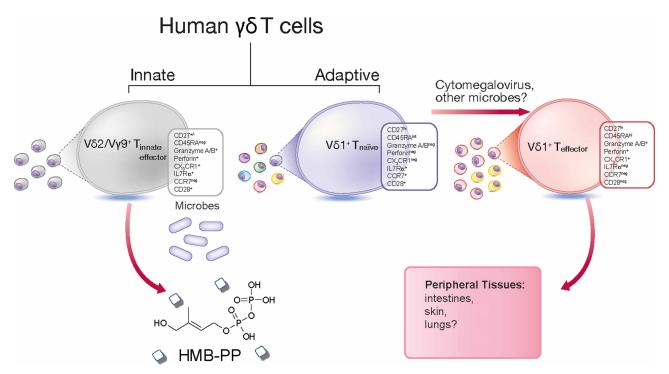


Figure 1. Innate-like and adaptive human $\gamma\delta$ T cell sub-compartments.

 $V\gamma9/V\delta2^+$ T cells are formed of an innate-like T cell population that possess a semi-invariant T cell receptor (TCR) and reactivity to the microbial-derived metabolite HMB-PP. Whereas post-natal $V\delta1^+$ T cells can be separated into naïve-like and effector populations. Naïve-like $V\delta1^+$ T cells display a diverse TCR repertoire and express CD27, CD28, CD62L, CCR7 and IL7R α on their cell surface. In contrast, effector $V\delta1^+$ T cells often possess clonally focussed TCRs, express low levels of CD27, up-regulate expression of the endothelial homing receptor CX3C chemokine receptor 1 (CX3CR1) and cytotoxic granzymes and perforin.

The role of $V\delta 3^+$ $\gamma\delta$ T cells has largely eluded understanding with early insights into immunobiology finding $V\delta 3^+$ T cells comprise a population of gut intraepithelial lymphocytes (IEL) [30]. $V\delta 3^+$ T cells also form a discrete population (~0.2%) of CD4⁻CD8⁻ peripheral T cells that are postulated to be innate-like due to the expression of NKG2D, CD56 and CD161 [31]. Peripheral $V\delta 3^+$ frequencies are known to increase in lupus patients, [32,33] cytomegalovirus [14] and HIV [34] although our knowledge of their immunobiology remains incomplete.

Current understanding of $\gamma\delta$ T cell activation

While bifurcation of $\gamma\delta$ T cells by V δ gene usage has revealed much about their biology, questions remain about antigen reactivity. Indeed, a central tenet of $\gamma\delta$ T cell biology has been their suggested breadth of antigen reactivity. The majority of bona fide ligands identified for human $\gamma\delta$ T cells to date have been described for the V δ 1 lineage, which can recognise proteins that adopt the MHC-like fold CD1b, [35] CD1c, [36] CD1d [37,38] and MR1 [39]. Broader V δ 1 reactivity towards non-MHC-like proteins has been established for Phycoerythrin, [40] and more recently Ephrin type-A receptor 2 [41] alongside V δ 3 and V δ 5 reactivity towards Annexin A2 and endothelial coupled protein-C receptor, respectively (Table 1) [42,43].

Conversely, $V\delta 2^+V\gamma 9^+$ T cells are broadly reactive to phosphoantigens that include (E)-4-Hydroxy-3-methyl-but-2-enyl pyrophosphate (HMBPP) a metabolite of the MEP pathway in Plasmodium falciparum, and isopentenyl pyrophosphate (IPP) a metabolite in the mevalonate pathway [10]. Such phosphoantigen (pAg) reactivity is dependent on the B7-protein like butyrophilin (BTN) 2A1 and 3A1 as well as the TCR (Table 1) [44–46]. The pAg activation of $V\delta 2^+$ T cells and the nexus of BTN and TCR molecules remains poorly understood despite growing understanding of their central importance in such reactivities [47]. Thus $\gamma \delta$ T cell receptors seem to enable recognition of a broader range of protein antigens, compared with their $\alpha \beta$ counterparts.



Table 1 Identified $\gamma\delta$ TCR ligands and gene restrictions

Human γδ	_			
lineage	Tissue	Ligand	Structural fold	Ref.
Vδ restriction				
V81	Peripheral blood	CD1b	MHC class I-like	[35]
V81	Peripheral blood	CD1c	MHC class I-like	[36]
Vδ1	Peripheral blood	CD1d	MHC class I-like	[37,38]
Vδ1/Vδ3	Peripheral blood	MR1	MHC class I-like	[39]
Vδ1	Peripheral blood	R-Phycoerythrin	Phycobiliprotein	[40]
<i>Vδ</i> 3	-	Annexin A2	Annexin	[43]
$V\gamma$ restriction (or p	airings)			
Vy4	Enteric	BTNL3-BTNL8	B7-like	[71,73,78]
<i>Vγ</i> 9– <i>Vδ</i> 2	Peripheral blood	BTN3A-BTN2A	B7-like	[47,63]
<i>Vγ3–Vδ2</i>	Intramuscular	Aminoacyl tRNA synthetase	Class II synthetase	[79]
<i>Vγ4–Vδ</i> 5	Peripheral blood	Endothelial protein-C receptor	MHC α 1- α 2-like	[42]
<i>Vγ9–Vδ1</i>	Peripheral blood	Ephrin type A receptor 2	Ephrin receptor tyrosine kinase	[41]

Structural determinants of $\gamma\delta$ T cell reactivity

Clarity around the paradigms for T cell reactivities, and indeed principles of selection, have been greatly advanced through a structural lens. Here, a multitude of $\alpha\beta$ TCR structures have solidified our understanding of MHC-restriction, T cell immunity and autoreactivities for conventional and unconventional $\alpha\beta$ T cells. In contrast, a dichotomy around δ T cell reactivities has provided confusion, here parallels are often drawn to conventional T cell subsets, yet a defining feature of $\gamma\delta$ T cells is the diversity of the antigens they recognise and the suggestion of how such reactivities are achieved.

Although this review is focused on human $\gamma\delta$ T cells, the first structural insight was provided via comparison to a murine $\gamma\delta$ TCR structure in complex with the stress induced $\beta2$ -microglobulin associated MHC class I molecule, T22. Here, polyclonal T cell reactivity to T22 was found to be driven by the CDR loops of the TCR as exemplified by a V $\gamma4$ -V $\alpha11.3$ /V $\delta10$ clone, termed G8 TCR (Figure 2a) [48]. The resultant structure was unlike any other, here the G8 TCR bound the collapsed MHC-like antigen presenting groove of T22 with an interface dominated by the δ -chain which corresponded to \sim 90% of the buried surface area (BSA) and included key germline encoded contacts (Figure 2f,k) [49]. This structure revealed that $\gamma\delta$ T cells could adopt highly unconventional docking strategies in mice although the translation to humans and other antigens was unknown.

The clearest insight we have to date on human $\gamma\delta$ T cell reactivities centres on adaptive V δ 1 T cells responding to MHC-like molecules. A population of peripheral CD1d restricted V δ 1⁺ population of T cells found to exhibit differing antigens specificities and representing 0.05–3.5% of CD1d–tetramer⁺ cells were isolated. Here an element of CD1d autoreactivity was identified with ligand independent tetramer reactivities although some clones showed differing lipid specificities α -galactosylceramide (α -GalCer), β -galactosylceramide (β -GalCer) and β -glucosylceramide (β -GlcCer). Notably the phenotype and functional potential of the CD1d restricted $\gamma\delta$ T cells differed from a type I NKT (CD8⁻/CD4⁻, levels of IFN- γ production). One specific clone, termed 9C2 (V γ 5–V δ 1⁺) was shown to be strongly responsive to CD1d presenting α -GalCer but some autoreactivity to CD1d [37]. Indeed, a resultant structure provided molecular insight into this autoreactivity, with the 9C2 TCR binding the polar extreme of the CD1d A' pocket, distinct from type I and type II NKT TCR–CD1d recognition (Figure 2b) [50,51]. The interface comprised of a majority (75%) of δ -chain contacts which included a mix



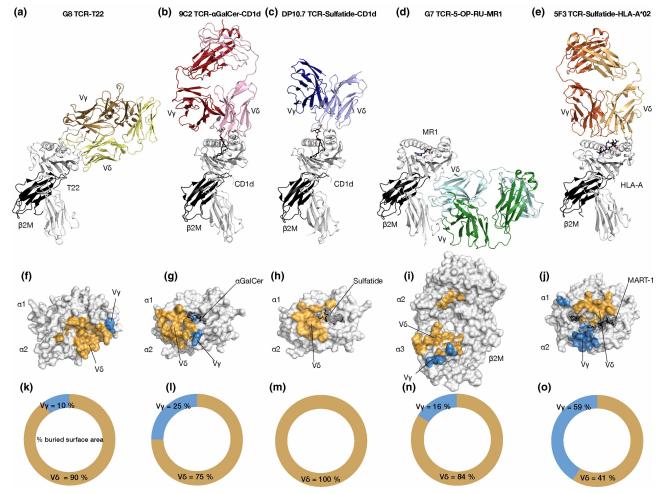


Figure 2. Analysis of the $V\delta$ dominated interface of MHC-like molecule reactivity.

Overall docking mode and interface analysis of yd TCR-antigen ternary structures. Cartoon representation of the Murine G8 ($V_74V_611.3$) TCR mediated recognition of T22 (**a**), 9C2 (V_75V_61) TCR mediated recognition of CD1d-aGalCer (**b**), DP10.7 (V_74V_61) TCR mediated recognition of CD1d-sulfatide (**c**), G7 (V_79V_61) TCR mediated recognition of MR1-5-OP-RU (**d**) and 5F3 (V_78V_61) TCR mediated recognition of HLA*A2-MART-1 (**e**). Interface analysis of the corresponding complex TCR structures with the V_6 and V_7 mediated contacts coloured orange and blue, respectively, with the percentage contributions to the BSA shown, G8-T22 (**f** and **k**), 9C2-CD1d-aGalCer (**g** and **l**), DP10.7-CD1d-sulfatide (**h** and **m**), G7-MR1-5-OP-RU (**i** and **n**) and 5F3-HLA*A2-MART-1 (**j** and **o**).

of germline and non-germline contacts to CD1d whereas the ligand mediating contacts involved the CDR3 γ (25% BSA γ -chain) (Figure 2g,l).

Further insight into this T cell niche was afforded upon the isolation of a $\gamma\delta$ T cell population responsive to CD1d presenting sulfatide, a sulfated β -galactosylceramide that is abundant in myelin and associated in demyelination diseases [52]. Here a rarefied population, 1% of peripheral V δ 1⁺ T cells, of CD1d-sulfatide restricted $\gamma\delta$ T-cells was insufficient to enable phenotyping although one clone termed DP10.7 (V γ 4V δ 1), was shown to confer sulfatide specificity [38]. Here the DP10.7 TCR docked similarly to the 9C2 clone, but closer to the type II NKT $\alpha\beta$ counterpart (Figure 2c) [51]. As with the similar docking manner, the interface was biased towards δ -chain contacts here accounting for the entire interface (Figure 2h,m) [38]. As observed with 9C2, germline encoded regions mediated contact to the CD1d surface however for DP10.7 the CDR3 δ provided the ligand specific contacts to the presented lipid antigen. Thus, the V γ played no role in TCR recognition of CD1d-sulfatide and only ligand deciphering contacts to CD1d presenting α -GalCer.

Following the isolation of a V δ 1⁺ CD1d $\gamma\delta$ subset, an MR1 reactive peripheral blood and tissue $\gamma\delta$ T cell population was identified. These cells ranged from <0.1–5% of $\gamma\delta$ T cells and were mostly CD8⁻/CD4⁻ with



variable CD161 expression, in contrast with typical MR1-restricted $\alpha\beta$ T cells. Many of the responding $\gamma\delta$ T cell clones displayed varied activation thresholds and antigen specificity with suggested auto reactivity. A clone termed G7 (V γ 9V δ 1) bound MR1 with moderate affinity whether presenting 5-(2-oxopropylideneamino)-6-Dribitylaminouracil (5-OP-RU), 6-formylpterin (6-FP) or acetyl-6-formylpterin (Ac-6-FP). Structural analysis of this clone revealed a completely novel docking topology binding the underneath of the MR1 antigen presenting groove, unlike any $\gamma\delta$ TCR (Figure 2e). Indeed, the interaction was beyond anything observed for MR1 reactive $\alpha\beta$ T cells which converge on the antigen presenting groove following conventional paradigms.

Despite such a novel interaction with MR1, the interface was directed by the δ -chain which comprised 84% of the BSA (16% γ -chain) (Figure 2i,n). The δ -chain contacts to the α 3 domain of MR1 was principally germline encoded, including the CDR1 δ , δ -chain framework residues alongside some non-germline CDR3 δ contacts. Thus, the building picture of $\gamma\delta$ TCR mediated reactivity is one where the δ -chain plays a principle role, indeed mutagenesis studies of MR1 to disrupt the paratope of the δ -chain showed both its central role in the docking mechanism employed by the G7 TCR, and showed other clones converged on neighbouring regions of MR1.

The exception to this evolving picture stemmed from $ex\ vivo$ culturing of haematopoietic stem progenitor cells to yield MART-1-HLA*A2 reactive CD8⁺ $\alpha\beta$ and $\gamma\delta$ T cells, with analogous populations derived from CD4⁻ T cells from umbilical cord blood. Functional testing was limited due to the scarcity of such cells however MART-1-specific $\gamma\delta$ TCR transduced cell lines produced interferon- γ and displayed effector function after stimulation, expressing granzyme B) and CD107a. Structural insight into one clone, termed 5F3 was structurally resolved to reveal an unorthodox docking mode again. Here using a δ - α comparisons the 5F3 TCR docked over the antigen binding groove although in reverse orientation, when compared with an $\alpha\beta$ restricted MART-1 HLA*A2, [53] akin to a few $\alpha\beta$ exceptions (Figure 2d) [54,55]. In separation from other $\gamma\delta$ structures the 5F3 had a relatively balanced interface, whereby the γ - and δ -chains contributed equally (59 & 41% BSA, respectively) to the interaction (Figure 2j,0). The interface included contacts from each CDR loop, except for the CDR3 γ , although a key reactivity conferring interaction stemmed from W98 of the CDR3 δ . Here W98 made prominent contacts to the MART-1 peptide replicating analogous peptide contacts from a key hydrophobic observed in $\alpha\beta$ restriction. Whilst questions remain around the generality of such melanoma specific T cells and their existence in the periphery, this work illustrates that $\gamma\delta$ T cells can expand and fill immunological voids as seen in diseases that deplete MAIT cells [56].

Thus, in conclusion, the paradigms that have held for $\alpha\beta$ restriction to MHC over decades are yet to hold for any determined $\gamma\delta$ TCR mediated antigen recognition pairing. Instead the emergent theme is one whereby each TCR adopts a recognition mechanism unique to the clone, or a small sub-population of the responding cells. The antigen recognition interfaces include a dominant role of the δ -chain, sometimes exclusively, leaving the γ -chain either to play an ancillary role in ligand deciphering or completely free from the interface. Early studies have noted that the CDR δ chain is the most polymorphic and diverse, correlating with its ability to endow such varied docking modes, however the role of the γ -chain remains ill-defined, at least in an adaptive capacity.

The functional role of $V\gamma$ gene segments in BTN/L reactivity

Intriguingly, the γ -chain is of central importance in some innate-like $\gamma\delta$ T cell subsets, whereby strong restrictions of the V γ gene usage correlate with TCR mediated reactivities to infectious disease or tumour associated metabolites. Namely, V γ 9V δ 2 cells were shown to respond to non-peptide pAg (IPP & HMPP) [57–61]. Since these seminal discoveries, it was found that such $\gamma\delta$ T cells are activated in a V γ 9V δ 2 TCR dependant manner that is dependent on members of the butyrophilin 3A family (BTN3A1, BTN3A2, BTN3A3) [62,6344]. Structural insight into the BTN3A molecules revealed the architecture of the extracellular IgV and IgC domains which provided insight into homodimerization [64].

Contention arose over how the pAg induced BTN3A mediated $V\gamma9V\delta2$ activation was achieved. Initial suggestion of a possible antigen presentation role for BTN3A molecules is now viewed as unlikely due to mounting evidence of the central role of the B30.2 domain in pAg binding [65]. Structures of the B30.2 domain of BTN3A1 in complex with pAgs showed the ligand binds within a positively charged pocket, [46,66] in close agreement with mutational studies of the binding site, [67] with binding shown to induce conformational changes within the B30.2 domain [68]. Furthermore, this study suggested that BTN3A1 formed a V-shaped homodimer on a resting cell-surface, with di-sulphide locking of the homodimer shown to impede pAg detection [68]. Recently, BTN2A1 has been implicated in pAg reactivity by $V\gamma9V\delta2T$ cells. Here, BTN2A1



expression was closely associated with BTN3A1 at the cell-surface with co-expression required for pAg reactivity, suggesting a potential role for heterodimerisation although direct evidence of this is outstanding despite the observation of BTN3A1 heterodimers [47,69]. Furthermore, BTN2A1 was shown to directly bind a charged patch of the TCR in a germline encoded region of the V γ 9, distal to the CDR loops that was found to be central to the reactivity [47]. Thus the evolving picture of V γ 9V δ 2 reactivity to pAgs involves BTN3A1 & BTN2A1 which may heterodimerise and then transmits an extracellular signal or conformational change upon intracellular binding of pAg binding via the B30.2 domain (Figure 3). However, recent identification of BTN3A1 interacting with CD45-phosphatases, provides an alternative hypothesis, that pAg binding BTN3A1 then sequesters CD45 from the immune synapse allowing an as yet unidentified CDR3-dependent antigen to be recognised by the V γ 9V δ 2 TCR [70].

A tissue restricted population of $V\gamma 4^+ \gamma \delta$ T cells have been found to display reactivity to another butyrophilin family member, the butyrophilin-like (BTNL) molecule BTNL3, which heterodimerises with BTNL8 and resulted in a TCR-dependent response [69,71,72]. Indeed BTNL3 recognition by $V\gamma 4^+ \gamma \delta$ T cells led to CD69 up-regulation and TCR internalisation [71]. Sequence comparisons of BTNL3 reactive and non-reactive clones in conjunction with mutational studies of the TCR revealed key roles for germline encoded CDR2 γ and hypervariable region 4, termed HV4. Furthermore, characterisation of a soluble $V\gamma 4^+$ TCR termed LES, was shown

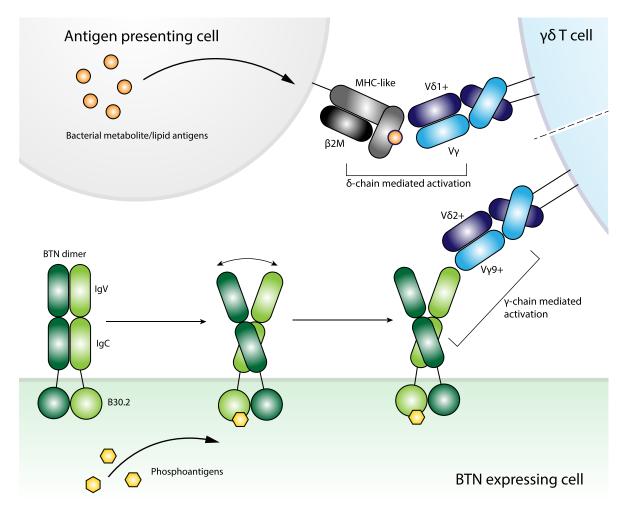


Figure 3. The divergent activation pathways of adaptive-like and innate-like $\gamma\delta$ T cells.

The activation mechanisms of adaptive-like $\gamma\delta$ T cells, so far, centre of V δ 1 mediated recognition of MHC-like molecules employing a heavy V δ -dominance for cognate antigen ligation. Innate-like $\gamma\delta$ T cells recognise bacterial lipid antigens (HMBPP & IPP) in a butyrophilin dependent manner and display a restricted V γ gene pairing. Although the molecular basis remains unknown V γ 9⁺ and V γ 4⁺ TCR confer reactivity to BTN3A1 and BTNL3, respectively, thus implicating the γ -chain at the interface.

to directly bind the IgV domain of BTNL3 with moderate affinity (\sim 15–25 μ M) with mutations of the CDR2 γ and HV4 region mitigating BTNL3 reactivity or conferring responsiveness upon inclusion of such motifs within typically non-responsive $\gamma\delta$ TCRs [71]. Mutational analysis of the BTNL3 molecule suggested that the CFG face of the IgV severely impacted binding by the LES TCR [71]. The mutational work led to a proposed model for the potential interaction although atomic structural data on this front is currently lacking.

Further work on IEL gut resident $V\gamma 4^+$ TCRs suggested a contesting hypothesis of BTNL3 restriction in celiac patients and are implicated in the inflammatory response therein [73]. Here single cell sequence analysis of the V $\delta 1^+$ IEL TCR repertoires identified a CDR3 γ histidine motif neighbouring the TRGJ1-encoded J γ segment (termed H-J1) that was overrepresented in patients with active celiac relative to healthy controls and celiac patients with gluten-free diets [73]. Further investigation showed remodelling of the IEL $\gamma\delta$ T cell compartment within celiac patients with altered ability to respond to BTNL3/8 correlated with disease progression. The underlying mechanism and the importance of the H-J1 motif are still to be determined with a large scale single-cell study of $\gamma\delta$ TCR sequences from celiac patients unable to identify the H-J1 motif [74].

Thus, $V\gamma 4^+ \gamma \delta$ T cells directly respond to BTNL3 in isolation from BTNL8, although they are thought to heterodimerise prior to cell surface expression [69]. Contesting hypotheses currently exist over the molecular basis for such restriction with elegant cellular work implicating either the germline HV4 regions or the non-germline H-J1 TCR regions of the γ -chain.

Discussion

The relative abundance of bona fide $\gamma\delta$ T cell ligands although threadbare has yielded pivotal insight into the molecular determinants of these enigmatic cells. What has become clear is that the paradigms of conventional T cell restrictions do not translate. Instead each subset must be treated individually despite the desire for universal truths. One general trend is the V δ 1-chain to dictate much of the interface in adaptive $\gamma\delta$ T cell responses, at least thus far. Such δ -chain dominance, in combination with the unique and extreme polymorphic capacity of this chain, [75] has enabled extremely unorthodox T cell recognition mechanisms that resemble those observed for antibody mediated B cell immunity (Figure 3).

In contrast with the δ -chain dominance of the adaptive antigen driven responses, a subset of innate-like $\gamma\delta$ T cells show a strong V γ restriction. Here peripheral V $\gamma9^+$ $\gamma\delta$ T cells confer reactivity to foreign lipid antigens in a BTN3A1 and BTN2A1 dependent manner. Whereas, V $\gamma4^+$ IEL $\gamma\delta$ T cells respond to the BTNL3 and BTNL8 heterodimer although the biological context for this interaction and any disease associated triggers remains unclear. Comprehensive mutational work has identified regions within the V γ domain that confers BTN/L reactivity and map to a site distinct from the antigen binding paratope of their adaptive counterparts (Figure 3). Notably due to the δ -chain dominance described above, the BTN/L binding regions of the V γ domain are typically accessible in adaptive antigen-driven complexes. Thus, one of the central unanswered questions of such innate-like $\gamma\delta$ T cell biology is the basis for the V γ restrictions in both BTN and BTNL mediated axes of $\gamma\delta$ T cell activation. Indeed, structural clarity on this front will be particularly revealing.

The importance and function of defined $\gamma\delta$ T cell interactions with cognate ligands within disease settings is not well understood. An expanded understanding of the potential ligand repertoire of $\gamma\delta$ T cells will likely serve as useful tools in deciphering such immune roles within infections and cancer. A tantalising premise is the potential convergence of the adaptive and innate capacities of such $\gamma\delta$ T cells, namely are antigen recognition and BTN and BTNL ligation events mutual exclusive or compounding. Previous investigations with MHC or B2M deficient cell-lines suggested that MHC and MHC-like molecules are not required although another protein is likely involved in $\gamma\delta$ T cell activation [76,77]. Thus, antigen identification experiments and structural analyses of $\gamma\delta$ T cells have provided profound insight into their biological functional and unique ligand bindings properties. However, central questions remain about their activation mechanisms, the true diversity of potential antigens and the adaptive or innate differences in $\gamma\delta$ TCR ligation. Answering of these questions will aid in our understanding of $\gamma\delta$ T cells and unlock their therapeutic potential moving forward.

Perspectives

 γδ T cells comprise a central yet often overlooked component of the tripartite adaptive immune system with particularly key roles in epithelial immunity and protection.



- Current knowledge on the activating ligands and recognition principles of $\gamma\delta$ T cells are incomplete. Despite this $\gamma\delta$ T cells are already being developed within the context of cellular immunotherapies.
- To fully realise the importance of $\gamma\delta$ T cells in protective immunity, and unleash their true immunotherapeutic potential, we must decipher the breadth of antigens that $\gamma\delta$ T cells can recognise and elucidate the activation paradigms across $\gamma\delta$ T cell subtypes.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Open Access Statement

Open access for this article was enabled by the participation of Monash University in an all-inclusive Read & Publish pilot with Portland Press and the Biochemical Society under a transformative agreement with CAUL.

Author Contributions

B.S.G, M.S.D and J.R wrote the manuscript.

Acknowledgements

We acknowledge funding from the National Health and Medical Research Council (NHMRC) (Australia), Australian Research Council (ARC) and National Institutes of Health (NIH). B.S.G is supported by an ARC Discovery Project (DP200103462 to B.S.G.). J.R. is supported by an ARC Australian Laureate Fellowship and the National Institute of Allergy and Infectious Diseases of the National Institutes of Health (NIH; R01Al148407 to J.R). M.S.D. is supported by an ARC Discovery Early Career Researcher Award and ARC Discovery Project (DP210103327 to M.S.D).

Abbreviations

5-OP-RU, 5-(2-oxopropylideneamino)-6-D-ribitylaminouracil; 6-FP, 6-formylpterin; Ac-6-FP, acetyl-6-formylpterin; BTN, Butyrophilin; BTNL, Butyrophilin-like; HMBPP, (E)-4-Hydroxy-3-methyl-but-2-enyl pyrophosphate; IPP, isopentenyl pyrophosphate; TCR, T cell receptor; termed H-J1, histidine motif neighbouring the TRGJ1-encoded J γ segment; α -GalCer, α -galactosylceramide; β -GalCer, β -galactosylceramide.

References

- 1 Willcox, B.E. and Willcox, C.R. (2019) Γδ TCR ligands: the quest to solve a 500-million-year-old mystery. *Nat. Immunol.* **20**, 121–128 https://doi.org/10.1038/s41590-018-0304-y
- Zheng, J., Liu, Y., Lau, Y.-L. and Tu, W. (2013) γδ-T cells: an unpolished sword in human anti-infection immunity. Cell. Mol. Immunol. 10, 50–57 https://doi.org/10.1038/cmi.2012.43
- 3 Silva-Santos, B., Serre, K. and Norell, H. (2015) Γδ T cells in cancer. Nat. Rev. Immunol. 15, 683–691 https://doi.org/10.1038/nri3904
- 4 Vantourout, P. and Hayday, A. (2013) Six-of-the-best: unique contributions of γ8 T cells to immunology. *Nat. Rev. Immunol.* **13**, 88–100 https://doi.org/10.1038/nri3384
- De Libero, G., Casorati, G., Giachino, C., Carbonara, C., Migone, N., Matzinger, P. et al. (1991) Selection by two powerful antigens may account for the presence of the major population of human peripheral gamma/delta T cells. *J. Exp. Med.* **173**, 1311–1322 https://doi.org/10.1084/jem.173.6.1311
- 6 Dimova, T., Brouwer, M., Gosselin, F., Tassignon, J., Leo, O., Donner, C. et al. (2015) Effector Vγ9Vδ2T cells dominate the human fetal γδ T-cell repertoire. *Proc. Natl Acad. Sci. U.S.A.* **112**, E556 https://doi.org/10.1073/pnas.1412058112
- 7 Davodeau, F., Peyrat, M.A., Hallet, M.M., Houde, I., And, H.V. and Bonneville, M. (1993) Peripheral selection of antigen receptor junctional features in a major human γδ subset. Eur. J. Immunol. 23, 804–808 https://doi.org/10.1002/ejj.1830230405
- 8 Parker, C.M., Groh, V., Band, H., Porcelli, S.A., Morita, C., Fabbi, M. et al. (1990) Evidence for extrathymic changes in the T cell receptor gamma/delta repertoire. *J. Exp. Med.* **171**, 1597–1612 https://doi.org/10.1084/jem.171.5.1597
- 9 Davey, M.S., Willcox, C.R., Hunter, S., Kasatskaya, S.A., Remmerswaal, E.B.M., Salim, M. et al. (2018) The human Vδ2+ T-cell compartment comprises distinct innate-like Vγ9+ and adaptive Vγ9- subsets. *Nat. Commun.* **9**, 1760 https://doi.org/10.1038/s41467-018-04076-0
- Morita, C.T., Jin, C., Sarikonda, G. and Wang, H. (2007) Nonpeptide antigens, presentation mechanisms, and immunological memory of human Vgamma2Vdelta2T cells: discriminating friend from foe through the recognition of prenyl pyrophosphate antigens. *Immunol. Rev.* 215, 59–76 https://doi. org/10.1111/j.1600-065X.2006.00479.x



- 11 Costa, G., Loizon, S., Guenot, M., Mocan, I., Halary, F., de Saint-Basile, G. et al. (2011) Control of *Plasmodium falciparum* erythrocytic cycle: γδ T cells target the red blood cell-invasive merozoites. *Blood* **118**, 6952–6962 https://doi.org/10.1182/blood-2011-08-376111
- 12 Sebestyen, Z., Prinz, I., Déchanet-Merville, J., Silva-Santos, B. and Kuball, J. (2020) Translating gammadelta (γδ) T cells and their receptors into cancer cell therapies. *Nat. Rev. Drug Discov.* **19**, 169–184 https://doi.org/10.1038/s41573-019-0038-z
- 13 Fisch, P., Malkovsky, M., Kovats, S., Sturm, E., Braakman, E., Klein, B.S. et al. (1990) Recognition by human V gamma 9/V delta 2T cells of a GroEL homolog on Daudi Burkitt's lymphoma cells. *Science* **250**, 1269 https://doi.org/10.1126/science.1978758
- 14 Déchanet, J., Merville, P., Lim, A., Retière, C., Pitard, V., Lafarge, X. et al. (1999) Implication of γδ T cells in the human immune response to cytomegalovirus. *J. Clin. Invest.* 103, 1437–1449 https://doi.org/10.1172/JCl5409
- 15 Couzi, L., Pitard, V., Netzer, S., Garrigue, I., Lafon, M.-E., Moreau, J.-F. et al. (2009) Common features of γδ T cells and CD8+ αβ T cells responding to human cytomegalovirus infection in kidney transplant recipients. *J. Infect. Dis.* **200**, 1415–1424 https://doi.org/10.1086/644509
- Farnault, L., Gertner-Dardenne, J., Gondois-Rey, F., Michel, G., Chambost, H., Hirsch, I. et al. (2013) Clinical evidence implicating gamma-delta T cells in EBV control following cord blood transplantation. *Bone Marrow Transplant.* **48**, 1478–1479 https://doi.org/10.1038/bmt.2013.75
- Fujishima, N., Hirokawa, M., Fujishima, M., Yamashita, J., Saitoh, H., Ichikawa, Y. et al. (2007) Skewed T cell receptor repertoire of Vdelta1(+) gammadelta T lymphocytes after human allogeneic haematopoietic stem cell transplantation and the potential role for Epstein-Barr virus-infected B cells in clonal restriction. *Clin. Exp. Immunol.* **149**, 70–79 https://doi.org/10.1111/j.1365-2249.2007.03388.x
- Halary, F., Pitard, V., Dlubek, D., Krzysiek, R., de la Salle, H., Merville, P. et al. (2005) Shared reactivity of Vδ2neg γδ T cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells. J. Exp. Med. 201, 1567–1578 https://doi.org/10.1084/jem.20041851
- 19 Morita, C.T., Parker, C.M., Brenner, M.B. and Band, H. (1994) TCR usage and functional capabilities of human gamma delta T cells at birth. *J. Immunol.* **153**, 3979 PMID:7930606
- 20 Deusch, K., Lüling, F., Reich, K., Classen, M., Wagner, H. and Pfeffer, K. (1991) A major fraction of human intraepithelial lymphocytes simultaneously expresses the γ/δ T cell receptor, the CD8 accessory molecule and preferentially uses the Vδ1 gene segment. *Eur. J. Immunol.* **21**, 1053–1059 https://doi.org/10.1002/eji.1830210429
- 21 Brandtzaeg, P., Bosnes, V., Halstensen, T.S., Scott, H., Sollid, L.M. and Valnes, K.N. (1989) T lymphocytes in human gut epithelium preferentially express the α/β antigen receptor and are often CD45/UCHL1-positive. *Scand. J. Immunol.* **30**, 123–128 https://doi.org/10.1111/j.1365-3083.1989. tb01196.x
- 22 Bos, J.D., Teunissen, M.B., Cairo, I., Krieg, S.R., Kapsenberg, M.L., Das, P.K. et al. (1990) T-cell receptor gamma delta bearing cells in normal human skin. *J. Invest. Dermatol.* **94**, 37–42 https://doi.org/10.1111/1523-1747.ep12873333
- 23 Correia, D.V., Fogli, M., Hudspeth, K., da Silva, M.G., Mavilio, D. and Silva-Santos, B. (2011) Differentiation of human peripheral blood Vδ1+ T cells expressing the natural cytotoxicity receptor NKp30 for recognition of lymphoid leukemia cells. *Blood* 118, 992–1001 https://doi.org/10.1182/blood-2011-02-339135
- 24 Hudspeth, K., Fogli, M., Correia, D.V., Mikulak, J., Roberto, A., Della Bella, S. et al. (2012) Engagement of NKp30 on Vδ1T cells induces the production of CCL3, CCL4, and CCL5 and suppresses HIV-1 replication. Blood 119, 4013–4016 https://doi.org/10.1182/blood-2011-11-390153
- 25 Dunne, M.R., Elliott, L., Hussey, S., Mahmud, N., Kelly, J., Doherty, D.G. et al. (2013) Persistent changes in circulating and intestinal γδ T cell subsets, invariant natural killer T cells and mucosal-associated invariant T cells in children and adults with coeliac disease. PLoS ONE 8, e76008 https://doi.org/10.1371/journal.pone.0076008
- 26 Davey, M.S., Willcox, C.R., Joyce, S.P., Ladell, K., Kasatskaya, S.A., McLaren, J.E. et al. (2017) Clonal selection in the human Vδ1T cell repertoire indicates γδ TCR-dependent adaptive immune surveillance. *Nat. Commun.* 8, 14760 https://doi.org/10.1038/ncomms14760
- 27 Davey, M.S., Willcox, C.R., Baker, A.T., Hunter, S. and Willcox, B.E. (2018) Recasting human Vδ1 lymphocytes in an adaptive role. *Trends Immunol.* 39, 446–459 https://doi.org/10.1016/j.it.2018.03.003
- Tieppo, P., Papadopoulou, M., Gatti, D., McGovern, N., Chan, J.K.Y., Gosselin, F. et al. (2020) The human fetal thymus generates invariant effector gammadelta T cells. J. Exp. Med. 217, jem.20190580 https://doi.org/10.1084/jem.20190580
- 29 Vermijlen, D., Brouwer, M., Donner, C., Liesnard, C., Tackoen, M., Van Rysselberge, M. et al. (2010) Human cytomegalovirus elicits fetal gammadelta T cell responses in utero. J. Exp. Med. 207, 807–821 https://doi.org/10.1084/jem.20090348
- 30 Falk, M.C., Ng, G., Zhang, G.Y., Fanning, G.C., Kamath, K.R. and Knight, J.F. (1994) Predominance of T cell receptor V delta 3 in small bowel biopsies from coeliac disease patients. *Clin. Exp. Immunol.* **98**, 78–82 https://doi.org/10.1111/j.1365-2249.1994.tb06610.x
- 31 Mangan, B.A., Dunne, M.R., O'Reilly, V.P., Dunne, P.J., Exley, M.A., O'Shea, D. et al. (2013) Cutting edge: CD1d restriction and Th1/Th2/Th17 cytokine secretion by human V63T cells. *J. Immunol.* **191**, 30–34 https://doi.org/10.4049/jimmunol.1300121
- 32 Robak, E., Błoński, J.Z., Bartkowiak, J., Niewiadomska, H., Sysa-Jedrzejowska, A. and Robak, T. (1999) Circulating TCR gammadelta cells in the patients with systemic lupus erythematosus. *Mediators Inflamm.* **8**, 305–312 https://doi.org/10.1080/09629359990315
- 33 Robak, E., Niewiadomska, H., Robak, T., Bartkowiak, J., Błoński, J.Z., Wozniacka, A. et al. (2001) Lymphocyctes tgammadelta in clinically normal skin and peripheral blood of patients with systemic lupus erythematosus and their correlation with disease activity. *Mediators Inflamm.* **10**, 179–189 https://doi.org/10.1080/09629350124724
- 34 Taupin, J.L., Halary, F., Déchanet, J., Peyrat, M.A., Ragnaud, J.M., Bonneville, M. et al. (1999) An enlarged subpopulation of T lymphocytes bearing two distinct gammadelta TCR in an HIV-positive patient. *Int. Immunol.* **11**, 545–552 https://doi.org/10.1093/intimm/11.4.545
- 35 Reijneveld, J.F., Ocampo, T.A., Shahine, A., Gully, B.S., Vantourout, P., Hayday, A.C. et al. (2020) Human γδ T cells recognize CD1b by two distinct mechanisms. *Proc. Natl Acad. Sci. U.S.A.* 117, 22944–22952 https://doi.org/10.1073/pnas.2010545117
- 36 Roy, S., Ly, D., Castro, C.D., Li, N.-S., Hawk, A.J., Altman, J.D. et al. (2016) Molecular analysis of lipid-reactive Vδ1 γδ T cells identified by CD1c tetramers. *J. Immunol.* **196**, 1933–1942 https://doi.org/10.4049/jimmunol.1502202
- 37 Uldrich, A.P., Le Nours, J., Pellicci, D.G., Gherardin, N.A., McPherson, K.G., Lim, R.T. et al. (2013) CD1d-lipid antigen recognition by the γδ TCR. *Nat. Immunol.* **14**, 1137–1145 https://doi.org/10.1038/ni.2713
- 38 Luoma, A.M., Castro, C.D., Mayassi, T., Bembinster, L.A., Bai, L., Picard, D. et al. (2013) Crystal structure of Vδ1T cell receptor in complex with CD1d-sulfatide shows MHC-like recognition of a self-lipid by human γδ T cells. *Immunity.* **39**, 1032–1042 https://doi.org/10.1016/j.immuni.2013.11.



- 39 Le Nours, J., Gherardin, N.A., Ramarathinam, S.H., Awad, W., Wiede, F., Gully, B.S. et al. (2019) A class of γδ T cell receptors recognize the underside of the antigen-presenting molecule MR1. Science 366, 1522 https://doi.org/10.1126/science.aav3900
- 40 Zeng, X., Wei, Y.-L., Huang, J., Newell, E.W., Yu, H., Kidd, B.A. et al. (2012) Γδ T cells recognize a microbial encoded B cell antigen to initiate a rapid antigen-specific interleukin-17 response. *Immunity* 37, 524–534 https://doi.org/10.1016/j.immuni.2012.06.011
- 41 Harly, C., Joyce, S.P., Domblides, C., Bachelet, T., Pitard, V., Mannat, C. et al. (2021) Human γδ T cell sensing of AMPK-dependent metabolic tumor reprogramming through TCR recognition of EphA2. *Sci. Immunol.* **6**, eaba9010 https://doi.org/10.1126/sciimmunol.aba9010
- 42 Willcox, C.R., Pitard, V., Netzer, S., Couzi, L., Salim, M., Silberzahn, T. et al. (2012) Cytomegalovirus and tumor stress surveillance by binding of a human γδ T cell antigen receptor to endothelial protein C receptor. *Nat. Immunol.* **13**, 872–879 https://doi.org/10.1038/ni.2394
- 43 Marlin, R., Pappalardo, A., Kaminski, H., Willcox, C.R., Pitard, V., Netzer, S. et al. (2017) Sensing of cell stress by human γδ TCR-dependent recognition of annexin A2. *Proc. Natl Acad. Sci. U.S.A.* 114, 3163 https://doi.org/10.1073/pnas.1621052114
- 44 Harly, C., Guillaume, Y., Nedellec, S., Peigné, C.M., Mönkkönen, H., Mönkkönen, J. et al. (2012) Key implication of CD277/butyrophilin-3 (BTN3A) in cellular stress sensing by a major human γδ T-cell subset. *Blood* **120**, 2269–2279 https://doi.org/10.1182/blood-2012-05-430470
- 45 Rhodes, D.A., Chen, H.-C., Price, A.J., Keeble, A.H., Davey, M.S., James, L.C. et al. (2015) Activation of human γδ T cells by cytosolic interactions of BTN3A1 with soluble phosphoantigens and the cytoskeletal adaptor periplakin. *J. Immunol.* **194**, 2390 https://doi.org/10.4049/jimmunol.1401064
- 46 Sandstrom, A., Peigné, C.-M., Léger, A., Crooks James, E., Konczak, F., Gesnel, M.-C. et al. (2014) The intracellular B30.2 domain of butyrophilin 3A1 binds phosphoantigens to mediate activation of human Vγ9Vδ2T cells. *Immunity* 40, 490–500 https://doi.org/10.1016/j.immuni.2014.03.003
- 47 Rigau, M., Ostrouska, S., Fulford, T.S., Johnson, D.N., Woods, K., Ruan, Z. et al. (2020) Butyrophilin 2A1 is essential for phosphoantigen reactivity by γ8 T cells. *Science* **367**, eaay5516 https://doi.org/10.1126/science.aay5516
- 48 Shin, S., El-Diwany, R., Schaffert, S., Adams, E.J., Garcia, K.C., Pereira, P. et al. (2005) Antigen recognition determinants of γδ T cell receptors. *Science* **308**, 252 https://doi.org/10.1126/science.1106480
- 49 Adams, E.J., Chien, Y.-H. and Garcia, K.C. (2005) Structure of a γδ T cell receptor in complex with the nonclassical MHC T22. Science 308, 227 https://doi.org/10.1126/science.1106885
- 50 Borg, N.A., Wun, K.S., Kjer-Nielsen, L., Wilce, M.C.J., Pellicci, D.G., Koh, R. et al. (2007) CD1d—lipid-antigen recognition by the semi-invariant NKT T-cell receptor. *Nature* **448**, 44–49 https://doi.org/10.1038/nature05907
- 51 Patel, O., Pellicci, D.G., Gras, S., Sandoval-Romero, M.L., Uldrich, A.P., Mallevaey, T. et al. (2012) Recognition of CD1d-sulfatide mediated by a type II natural killer T cell antigen receptor. *Nat. Immunol.* **13**, 857–863 https://doi.org/10.1038/ni.2372
- 52 Bai, L., Picard, D., Anderson, B., Chaudhary, V., Luoma, A., Jabri, B. et al. (2012) The majority of CD1d-sulfatide-specific T cells in human blood use a semiinvariant Vδ1 TCR. Eur. J. Immunol. 42, 2505–2510 https://doi.org/10.1002/eji.201242531
- 53 Cole, D.K., Yuan, F., Rizkallah, P.J., Miles, J.J., Gostick, E., Price, D.A. et al. (2009) Germ line-governed recognition of a cancer epitope by an immunodominant human T-cell receptor. *J. Biol. Chem.* **284**, 27281–9 https://doi.org/10.1074/jbc.M109.022509
- 54 Beringer, D.X., Kleijwegt, F.S., Wiede, F., van der Slik, A.R., Loh, K.L., Petersen, J. et al. (2015) T cell receptor reversed polarity recognition of a self-antigen major histocompatibility complex. *Nat. Immunol.* **16**, 1153–1161 https://doi.org/10.1038/ni.3271
- 55 Gras, S., Chadderton, J., Del Campo Claudia, M., Farenc, C., Wiede, F., Josephs Tracy, M. et al. (2016) Reversed T cell receptor docking on a major histocompatibility class I complex limits involvement in the immune response. *Immunity* **45**, 749–760 https://doi.org/10.1016/j.immuni.2016.09.007
- Howson, L.J., Awad, W., von Borstel, A., Lim, H.J., McWilliam, H.E.G. Sandoval-Romero, M.L. et al. (2020) Absence of mucosal-associated invariant T cells in a person with a homozygous point mutation in MR1. *Sci. Immunol.* **5**, eabc9492 https://doi.org/10.1126/sciimmunol.abc9492
- 57 Constant, P., Davodeau, F., Peyrant, M.-A., Poquet, Y., Puzo, G., Bonneville, M. et al. (1994) Stimulation of human gamma delta T cells by nonpeptidic mycobacterial ligands. *Science* **264**, 267–270 https://doi.org/10.1126/science.8146660
- 58 Tanaka, Y., Morita, C.T., Tanaka, Y., Nieves, E., Brenner, M.B. and Bloom, B.R. (1995) Natural and synthetic non-peptide antigens recognized by human γδ T cells. *Nature* **375**, 155–158 https://doi.org/10.1038/375155a0
- 59 Espinosa, E., Belmant, C., Sicard, H., Poupot, R., Bonneville, M. and Fournié, J.-J. (2001) Y2k+1 state-of-the-art on non-peptide phosphoantigens, a novel category of immunostimulatory molecules. *Microbes Infect.* **3**, 645–654 https://doi.org/10.1016/S1286-4579(01)01420-4
- Morita, C.T., Lee, H.K., Wang, H., Li, H., Mariuzza, R.A. and Tanaka, Y. (2001) Structural features of nonpeptide prenyl pyrophosphates that determine their antiqenicity for human γδ T cells. *J. Immunol.* **167**, 36 https://doi.org/10.4049/jimmunol.167.1.36
- 61 Hintz, M., Reichenberg, A., Altincicek, B., Bahr, U., Gschwind, R.M., Kollas, A.-K. et al. (2001) Identification of (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human γδ T cells in *escherichia coli. FEBS Lett.* **509**, 317–322 https://doi.org/10.1016/S0014-5793(01) 03191-X
- 62 Bukowski, J.F., Morita, C.T., Band, H. and Brenner, M.B. (1998) Crucial role of TCRγ chain junctional region in prenyl pyrophosphate antigen recognition by γδ T cells. *J. Immunol.* **161**, 286 PMID:9647235
- 63 Wang, H., Henry, O., Distefano, M.D., Wang, Y.-C., Räikkönen, J., Mönkkönen, J. et al. (2013) Butyrophilin 3A1 plays an essential role in prenyl pyrophosphate stimulation of human Vγ2Vδ2T cells. *J. Immunol.* 191, 1029 https://doi.org/10.4049/jimmunol.1300658
- 64 Palakodeti, A., Sandstrom, A., Sundaresan, L., Harly, C., Nedellec, S., Olive, D. et al. (2012) The molecular basis for modulation of human Vγ9V82T cell responses by CD277/butyrophilin-3 (BTN3A)-specific antibodies. *J. Biol. Chem.* 287, 32780–32790 https://doi.org/10.1074/jbc.M112.384354
- 65 Vavassori, S., Kumar, A., Wan, G.S., Ramanjaneyulu, G.S., Cavallari, M., El Daker, S. et al. (2013) Butyrophilin 3A1 binds phosphorylated antigens and stimulates human γδ T cells. *Nat. Immunol.* **14**, 908–916 https://doi.org/10.1038/ni.2665
- 66 Yang, Y., Li, L., Yuan, L., Zhou, X., Duan, J., Xiao, H. et al. (2019) A structural change in butyrophilin upon phosphoantigen binding underlies phosphoantigen-mediated Vγ9Vδ2T cell activation. *Immunity* 50, 1043–53.e5 https://doi.org/10.1016/j.immuni.2019.02.016
- 67 Wang, H. and Morita, C.T. (2015) Sensor function for butyrophilin 3A1 in prenyl pyrophosphate stimulation of human Vγ2V82T cells. J. Immunol. 195, 4583 https://doi.org/10.4049/jimmunol.1500314
- 68 Gu, S., Sachleben, J.R., Boughter, C.T., Nawrocka, W.I., Borowska, M.T., Tarrasch, J.T. et al. (2017) Phosphoantigen-induced conformational change of butyrophilin 3A1 (BTN3A1) and its implication on Vγ9V82T cell activation. *Proc. Natl Acad. Sci. U.S.A.* 114, E7311–E7320 https://doi.org/10.1073/ pnas.1707547114
- 69 Vantourout, P., Laing, A., Woodward, M.J., Zlatareva, I., Apolonia, L., Jones, A.W. et al. (2018) Heteromeric interactions regulate butyrophilin (BTN) and BTN-like molecules governing γδ T cell biology. *Proc. Natl Acad. Sci. U.S.A.* 115, 1039 https://doi.org/10.1073/pnas.1701237115



- 70 Payne, K.K., Mine, J.A., Biswas, S., Chaurio, R.A., Perales-Puchalt, A., Anadon, C.M. et al. (2020) BTN3A1 governs antitumor responses by coordinating αβ and γδ T cells. *Science* **369**, 942 https://doi.org/10.1126/science.aay2767
- 71 Melandri, D., Zlatareva, I., Chaleil, R.A.G., Dart, R.J., Chancellor, A., Nussbaumer, O. et al. (2018) The γδTCR combines innate immunity with adaptive immunity by utilizing spatially distinct regions for agonist selection and antigen responsiveness. *Nat. Immunol.* 19, 1352–1365 https://doi.org/10.1038/s41590-018-0253-5
- 72 Di Marco Barros, R., Roberts, N.A., Dart, R.J., Vantourout, P., Jandke, A., Nussbaumer, O. et al. (2016) Epithelia use butyrophilin-like molecules to shape organ-specific γδ T cell compartments. *Cell* **167**, 203–218 https://doi.org/10.1016/j.cell.2016.08.030
- 73 Mayassi, T., Ladell, K., Gudjonson, H., McLaren, J.E., Shaw, D.G., Tran, M.T. et al. (2019) Chronic inflammation permanently reshapes tissue-resident immunity in celiac disease. *Cell* **176**, 967–81.e19 https://doi.org/10.1016/j.cell.2018.12.039
- 74 Eggesbø, L.M., Risnes, L.F., Neumann, R.S., Lundin, K.E.A., Christophersen, A. and Sollid, L.M. (2020) Single-cell TCR sequencing of gut intraepithelial γδ T cells reveals a vast and diverse repertoire in celiac disease. *Mucosal Immunol.* **13**, 313–321 https://doi.org/10.1038/s41385-019-0222-9
- 75 Rock, E.P., Sibbald, P.R., Davis, M.M. and Chien, Y.H. (1994) CDR3 length in antigen-specific immune receptors. *J. Exp. Med.* **179**, 323–328 https://doi.org/10.1084/jem.179.1.323
- 76 Correa, I., Bix, M., Liao, N.S., Zijlstra, M., Jaenisch, R. and Raulet, D. (1992) Most gamma delta T cells develop normally in beta 2-microglobulin-deficient mice. *Proc. Natl Acad. Sci. U.S.A.* 89, 653–657 https://doi.org/10.1073/pnas.89.2.653
- 77 Morita, C.T., Beckman, E.M., Bukowski, J.F., Tanaka, Y., Band, H., Bloom, B.R. et al. (1995) Direct presentation of nonpeptide prenyl pyrophosphate antigens to human gamma delta T cells. *Immunity* **3**, 495–507 https://doi.org/10.1016/1074-7613(95)90178-7
- 78 Willcox, C.R., Vantourout, P., Salim, M., Zlatareva, I., Melandri, D., Zanardo, L. et al. (2019) Butyrophilin-like 3 directly binds a human Vγ4⁺ T cell receptor using a modality distinct from clonally-restricted antigen. *Immunity*. 51, 813–25.e4 https://doi.org/10.1016/j.immuni.2019.09.006
- 79 Bruder, J., Siewert, K., Obermeier, B., Malotka, J., Scheinert, P., Kellermann, J. et al. (2012) Target specificity of an autoreactive pathogenic human γδ-T cell receptor in myositis. *J. Biol. Chem.* **287**, 20986–20995 https://doi.org/10.1074/jbc.M112.356709