Corrections

Regulation of gene expression by ambient pH in Aspergillus: genes expressed at acidic pH

By S. SARKAR, M.X. CADDICK, E. BIGNELL, J. TILBURN and H.N. ARST, JR

Volume 24 Part 2 (1996) 360–363

On page 362, the sequence shown in Figure 2 is not that of *pacA* but instead corresponds to a gene on chromosome VI (http://www-genome.wi.mit.edu/annotation/fungi/aspergillus/) which can rescue *pacA*$^{-}$ mutations by transformation. This gene has been renamed *suApacA* and the database entry (Z79750) has been corrected.

Application of monoclonal antibody libraries for the measurement of glycation adducts

By R. NAGAI and S. HORIUCHI

Volume 31 Part 6 (2003) 1438–1440

The authors should like to point out that Dr Yuka Unno, featured in the acknowledgements section, should have been accorded the status of a co-author on this paper.

Exceptionally diverse morphotypes and genomes of crenarchaeal hyperthermophilic viruses

By D. PRANGISHVILI and R.A. GARRETT

Volume 32 Part 2 (2003) 204–208

The legend to Figure 2 on page 207 carried an incorrect permissions acknowledgement credit line. The final sentence of the legend should have read as follows: Reprinted from M. Bettstetter, X. Peng, R.A. Garrett and D. Prangishvili, "AFV1, a novel virus infecting hyperthermophilic archaea of the genus *Acidianus*", Virology, vol. 315, pp. 68–79. © 2003, with permission from Elsevier.

The outer membrane of the hyperthermophilic archaeon Ignicoccus: dynamics, ultrastructure and composition

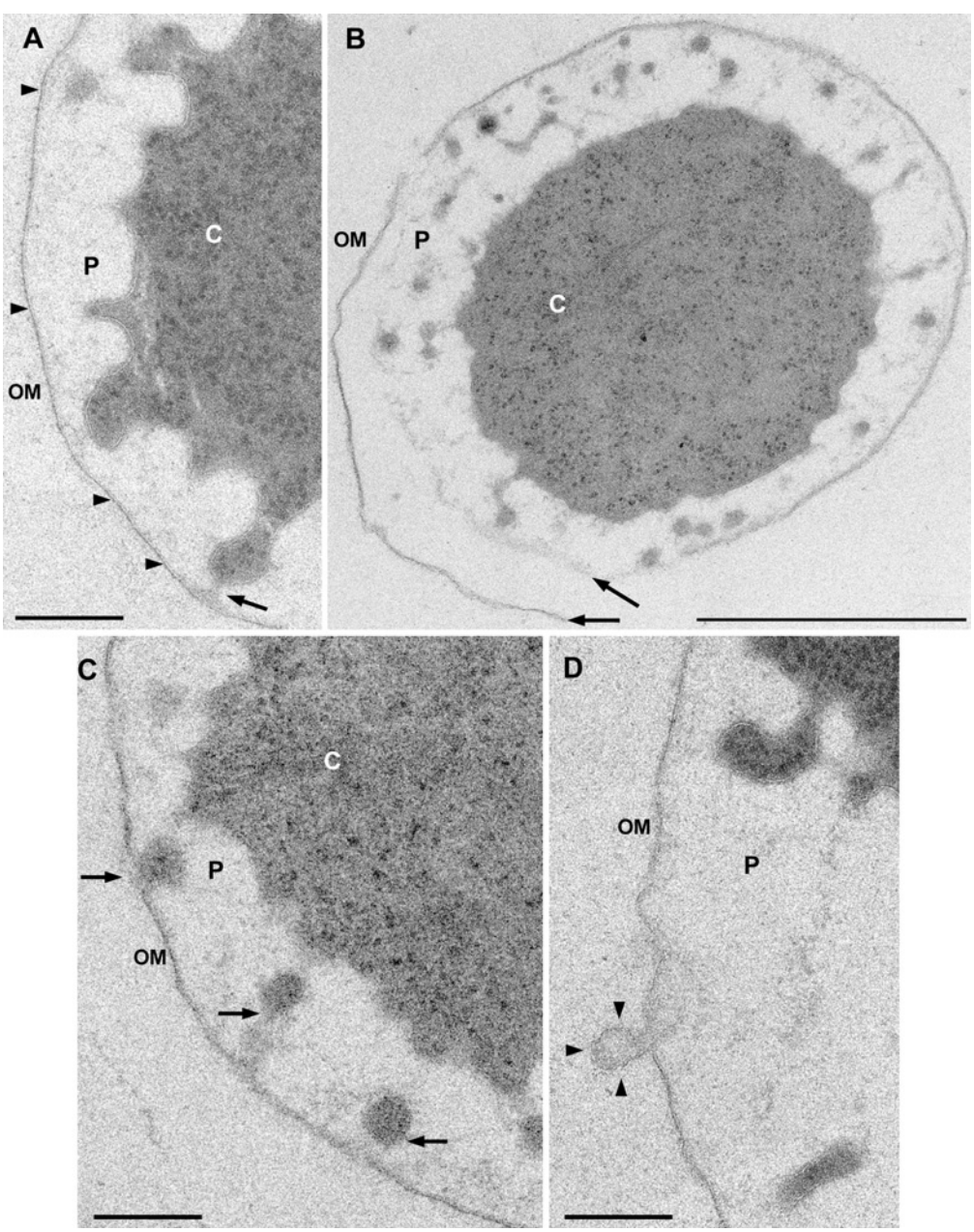
By D.J. NÄTHER and R. RACHEL

Volume 32 Part 2 (2004) 199–203

Owing to a problem at the printing stage, Figure 1 on page 200 did not appear at a sufficiently high resolution to enable the double lines of both the inner and the outer membrane of the *Ignicoccus* cells to be clearly visible. The Figure is reprinted here for this purpose, together with the associated legend.

Figure 1 | Structure, asymmetry and dynamics of the *Ignicoccus* outer membrane

(**A**, **C**, **D**) Parts of thin-sectioned cells; scale bar, 200 nm; (**B**) whole sectioned cell; scale bar, 1 μm. C, cytoplasm; P, periplasm; OM, outer membrane. Arrowheads in (**A**) show areas of the OM where two lines are visible; the arrow marks a vesicle getting into close contact with the OM. Arrows in (**B**) show the inner and outer leaflets of the OM, where they are separated. Arrows in (**C**) show vesicles getting into close contact with the OM, and one vesicle just fusing with the OM. Arrowheads in (**D**) mark a vesicle blebbing outwards from the OM.



Regulation of chloroplast translation: interactions of RNA elements, RNA-binding proteins and the plastid ribosome

By A. MANUELL, M.V. BELIGNI, K. YAMAGUCHI and S.P. MAYFIELD

Volume 32 Part 4 (2004) 601–605

Owing to a technical problem at the printing stage, the colours for Figure 2 as featured on page 604 were misprinted. The printed version of this Figure should have appeared as shown below (together with the associated Figure legend). The online version of the Figure was not affected.

Figure 2 | Ribosomal small-subunit proteins containing chloroplast unique domains

The bacterial 30 S ribosomal subunit (PDB 1J5E; [48]) is shown from the solvent side. rRNA (light green) and most of the proteins (light blue) are shown as backbone trace. Ribosomal proteins S2 (yellow), S3 (green) and S5 (blue) are shown as surfaces. Marked in red on each of these subunits is the residue where the unique chloroplast protein domain would be connected. For perspective, the additional domains from these three proteins would equal an additional S2, two additional S3s and three additional S5s. Arrows mimic the travel of mRNA around the back of the neck of the 30 S subunit, with S3 and S5 proteins flanking the site where the mRNA leaves the ribosome. GRASP [49], Molscript and POV-ray were used for creating this Figure.