

Review Article

Molecular mechanism of Fast Endophilin-Mediated Endocytosis

Alessandra Casamento¹ and  Emmanuel Boucrot^{1,2}

¹Institute of Structural and Molecular Biology, University College London, Gower Street, London WC1E 6BT, U.K.; ²Institute of Structural and Molecular Biology, Birkbeck College, Malet Street, London WC1E 7HX, U.K.

Correspondence: Emmanuel Boucrot (e.boucrot@ucl.ac.uk)



Endocytosis mediates the cellular uptake of micronutrients and cell surface proteins. Clathrin-mediated endocytosis (CME) is the housekeeping pathway in resting cells but additional Clathrin-independent endocytic (CIE) routes, including Fast Endophilin-Mediated Endocytosis (FEME), internalize specific cargoes and support diverse cellular functions. FEME is part of the Dynamin-dependent subgroup of CIE pathways. Here, we review our current understanding of the molecular mechanism of FEME. Key steps are: (i) priming, (ii) cargo selection, (iii) membrane curvature and carrier formation, (iv) membrane scission and (v) cytosolic transport. All steps are controlled by regulatory mechanisms mediated by phosphoinositides and by kinases such as Src, LRRK2, Cdk5 and GSK3 β . A key feature of FEME is that it is not constitutively active but triggered upon the stimulation of selected cell surface receptors by their ligands. In resting cells, there is a priming cycle that concentrates Endophilin into clusters on discrete locations of the plasma membrane. In the absence of receptor activation, the patches quickly abort and new cycles are initiated nearby, constantly priming the plasma membrane for FEME. Upon activation, receptors are swiftly sorted into pre-existing Endophilin clusters, which then bud to form FEME carriers within 10 s. We summarize the hallmarks of FEME and the techniques and assays required to identify it. Next, we review similarities and differences with other CIE pathways and proposed cargoes that may use FEME to enter cells. Finally, we submit pending questions and future milestones and discuss the exciting perspectives that targeting FEME may boost treatments against cancer and neurodegenerative diseases.

Introduction

Cell surface transmembrane proteins and extracellular material too large to diffuse through the membrane bilayer or be transported by channels and transporters are internalized through membrane-bound carriers, a process called endocytosis. The various endocytic pathways that exist form parallel portals of entry into cells. Clathrin-mediated endocytosis (hereafter, CME) is the best-characterized and the constitutively active and dominant uptake mechanism to support housekeeping functions in all eukaryotic cells [1–3]. There is evidence of several distinct mechanisms of Clathrin-independent endocytosis (CIE), including the CLIC/GEEC pathway (Clathrin-independent carriers (CLIC), glycosylphosphatidylinositol-anchored proteins (GPI-AP)-enriched early endosomal compartments (GEEC)), IL2R β uptake, EGFR non-clathrin endocytosis (EGFR-NCE), Fast Endophilin-Mediated Endocytosis (FEME), massive endocytosis (MEND), macropinocytosis, as well as neuron-specific activity-dependent bulk endocytosis (ADBE) and ultrafast endocytosis (UFE) [4–9] (Figure 1). Caveolae can in theory contribute to Clathrin-independent uptake. Even though caveolae can bud from the cell surface, few if any cargoes rely on them for their uptake [10,11]. The pathways are defined by characteristic endocytic carrier morphologies, by cytosolic markers, by ligands and receptors (collectively called ‘cargoes’) that use them to enter cells or by the speed of completion [6–9,12].

Received: 10 March 2020
 Revised: 11 May 2020
 Accepted: 18 May 2020

Version of Record published:
 26 June 2020

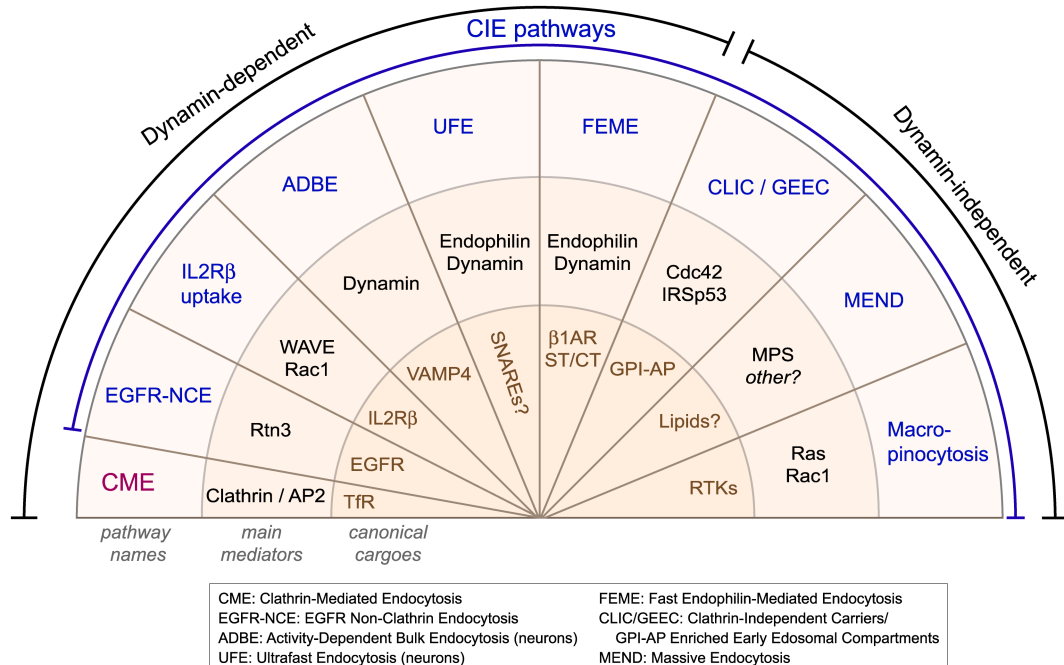


Figure 1. Clathrin-independent endocytic pathways.

Clathrin-mediated endocytosis (CME) is the house-keeping pathway in resting cells. It is mediated by Clathrin and the tetrameric adaptor AP2 and its canonical cargo is Transferrin Receptor (TfR). Clathrin-independent endocytosis (CIE) is composed of Dynamamin-dependent and Dynamamin-independent pathways. EGFR Non Clathrin Pathway (EGFR-NCE) is regulated by Reticulon-3 (Rtn3) and internalizes Epidermal Growth Factor Receptor (EGFR) upon low doses of EGF. IL2Rβ uptake is a constitutive CIE pathway that internalizes IL2Rβ and γ under the control of Rac1 and WAVE. Activity-Dependent Bulk Endocytosis (ADBE) is controlled by Dynamamin and internalizes VAMP4 and large patches of membranes upon high stimuli in neurons. Ultrafast Endocytosis (UFE) mediates the recycling of synaptic vesicle components (SNAREs?) in 50–100 ms following action potential in neurons. It is regulated by Endophilin, Dynamamin and Synaptojanin. Fast Endophilin-Mediated Endocytosis (FEME) internalizes cargoes such as the β 1-adrenergic receptor (β 1AR) in 5–10 s following their stimulation, in an Endophilin- and Dynamamin-dependent manner. Shiga toxin (ST) and cholera toxin (CT) can hijack FEME to enter cells, but can also use other CIE pathways. The Clathrin-Independent Carriers (CLIC)/GPI-anchored proteins (GPI-AP)-Enriched Early Endosomal Compartments (GEEC) pathway is a high capacity, Dynamamin-independent, endocytic route, triggered by the extracellular clustering of GPI-AP, glycosylated proteins or lipids by Galectin-3. It is controlled by Cdc42, GRAF-1 and IRSp53. Massive Endocytosis (MEND) is the significant uptake of membrane induced upon Ca^{2+} and PI3 kinase signaling, mediated by membrane phase separation (MPS). Macropinocytosis is activated by strong and sustained Receptor Tyrosine Kinase (RTKs) signaling and form large (up to 20 μm) vacuoles upon the folding of membrane projections back to the cell surface.

Overall, CIE pathways were observed in a wide variety of *in vitro* cell lines, *ex vivo* primary cells, as well as *in vivo* in mouse, fly, worm, plant and yeast. Several CIE processes are not constitutively active and perform specific or temporally regulated cellular functions. These range from bulk lipid and extracellular protein uptake and removal of activated receptors from the cell surface, to the control of cell polarization, spreading and migration [4–9].

Some CIE pathways such as UFE, FEME and MEND are quicker than CME and function in physiological processes requiring fast (<10 s) internalization from the plasma membrane (Figure 1), such as reaction to stress hormones ('fight-or-flight' response) and receptor hyper-stimulation, chemotaxis or compensatory endocytosis following exocytosis of synaptic or hormone-containing vesicles [6,13]. Many pathogens hijack CIE pathways to infect cells: these include over twenty viruses (including Ebola, HIV, Lassa, Herpes, Dengue and SV40 viruses), some bacteria, prions and bacterial toxins (including cholera and Shiga toxins, Streptolysin O and VacA [14,15]). Finally, deregulations of CIE have been reported during cancer, lysosomal storage disease or atherosclerosis.

FEME was recently added to the CIE family of pathways (Figure 1) and some of its molecular steps are now established [16–18]. The existence of specific cargoes, endocytic carrier attributes and cytoplasmic markers enabled the swift elucidation of key molecular and regulatory mechanisms. In addition, although FEME was only defined few years ago, some of its cargoes have been studied for longer, in particular cholera and Shiga toxins that can hijack FEME to infect cells [17]. These were instrumental in establishing the scission mechanism of FEME [17,19]. In this review, we outlay our current understanding of FEME and compare and contrast it with other CIE pathways. We also discuss potential cargoes that may use FEME and propose future milestones in the understanding of the pathway.

Hallmarks of FEME

FEME is a Dynamin-dependent, Clathrin-independent endocytic route marked and regulated by Endophilin. Part of the BAR domain superfamily, Endophilin consists of five proteins in human: Endophilin A1, A2, A3, B1 and B2. Only A1, A2 and A3 function in FEME (and thus collectively referred to as ‘Endophilin’), while Endophilin B1 and B2 are involved in autophagy and mitochondrial dynamics [20]. Endophilin A2 is ubiquitously expressed whereas Endophilin A1 and A3 are tissue restricted (mostly brain, kidney and testes). However, compensations exist upon genetic ablation of any one of them [21] and the depletion of all three forms is required to block FEME [16]. Endophilin functions in CME by supporting Dynamin-induced scission and Synaptojanin-mediated Clathrin uncoating [21]. However, Endophilin is redundant in CME (its roles can be sustained by Amphiphysin and SNX9 [22,23]), but essential in FEME. Moreover, Endophilin is only detected on a fraction of forming Clathrin-coated vesicles [24], confirming a likely peripheral role in CME.

Endophilin features two functional domains: a BAR domain — banana-shaped concave structure mediating protein dimerization and membrane curvature — and a SRC Homology 3 (SH3) domain that binds to proline-rich regions in target proteins [20,25,26]. The BAR domain contains two amphipathic helices H0 and H1 (H0 is at the N-terminus, making a ‘N-BAR domain’ [25]), which promote membrane curvature sensing, stabilization and formation beyond that of a regular BAR domain. Endophilin is the main marker and regulator of FEME. It performs three functions: (i) membrane curvature promoted by the N-BAR domain; (ii) cargo engagement through its SH3 domain; and (iii) membrane scission, upon recruitment of Dynamin and actin via interactions mediated by the SH3 domain and membrane friction and lipid diffusion barrier by its N-BAR domain.

FEME is defined by the formation of Clathrin-negative, Endophilin-positive endocytic carriers that bud from the plasma membrane upon stimuli. Hallmarks of FEME to date are:

- (1) it is not constitutively active, but is rapidly (few seconds) triggered by the activation of receptors by their cognate ligands [16];
- (2) it requires the pre-enrichment of Endophilin into discrete clusters on the plasma membrane, prior to receptor activation [18];
- (3) it is prominent at the leading edge of cells but also occurs on basal and dorsal cell surfaces [16];
- (4) FEME carriers are small ($\sim 1 \mu\text{m}$), pleiotropic, tubulo-vesicular Endophilin positive assemblies (EPAs) found in the cytosol [16]. Unlike CME, where the Clathrin coat is lost soon after budding, Endophilin remains on FEME carriers until fusion with early endosomes;
- (5) to date, there are 16 confirmed FEME cargoes: $\beta 1$ and $\alpha 2\text{a}$ adrenergic receptors, Dopamine receptors 3 and 4, Muscarinic Acetylcholine receptor 4, EGFR, HGFR, VEGFR, PDGFR, NGFR, IGFR, tetrameric IL2R, PlexinA1 and ROBO1, as well as cholera and Shiga toxins [16,17,27,28]. However, many of these cargoes can also use other endocytic pathways to enter cells;
- (6) membrane scission requires the synergy between Endophilin, Dynamin and actin [17,19];
- (7) cytosolic FEME carriers move retrogradely (towards the perinuclear area) on microtubules, powered by Dynein [17,28];
- (8) it is negatively regulated by Cdk5 and GSK3 β [28]. Acute inhibition of the kinases is sufficient to induce FEME.

Several of these characteristics must be met to establish that FEME is the portal of entry of candidate cargoes. Technical and protocol details are important to preserve and detect FEME carriers.

Mechanism of FEME carrier formation

Priming

The promptness of the pathway to activate upon receptor stimulation is explained by the priming cycle of FEME. In resting cells, Endophilin is pre-enriched into small patches at discrete locations on the plasma membrane. The precession of Endophilin on the membrane before cargo sorting is critical for prompt carrier formation. In resting cells, the vast majority of Endophilin molecules are auto-inhibited in the cytosol, through intra-dimer, inter-monomer interactions between H0 helices and SH3 domains from different subunits within homodimers [29]. As engagement of the SH3 domain with a PRM is necessary, and sufficient, to relieve the auto-inhibition, Endophilin must be targeted to the membrane by other proteins *in vivo* [18,22]. Only then can its N-BAR domain can bind to the membrane to sense, stabilize or induce local membrane curvature.

Endophilin clusters on the plasma membrane of resting cells are dynamic and last from 5 to 15 s before abortion (dissolution without endocytosis). As they rely on Pi(3,4)P₂ production from Pi(3,4,5)P₃, they are prominent at the leading edge of migrating cells, but are also numerous on the ventral and dorsal surfaces of confluent monolayers [16,18]. The priming cycle is initiated by membrane-bound, active, GTP-loaded Cdc42, which recruits FBP17 and CIP4, via their REM (also called HR1) motifs (Figure 2, Stage I). FBP17 and CIP4 concentrate on the membrane upon binding of their F-BAR domains and heterodimerization [18]. The tissue-restricted TOCA1 might be involved in this step in some cells as it heterodimerizes with both FBP17 and CIP4, and all three are functionally redundant in FEME [18]. Next, the SH3 domains of FBP17 and CIP4 recruit both SHIP1/2 phosphatases and Lamellipodin. SHIP1 and 2 are 5'-phosphatases that locally hydrolyze Pi(3,4,5)P₃ into Pi(3,4)P₂, which is then bound by the PH domain of Lamellipodin, further stabilizing the latter (Figure 2, Stage I). Lamellipodin has multiple PRMs to which Endophilin bind, and thus it concentrates many copies into patches [30]. These events are obligatory steps which must happen in sequential manner. Depletion of any of these proteins stalls the process downstream of the step targeted, which invariably block FEME carrier formation upon receptor activation [18].

Once Endophilin is pre-enriched into clusters, receptor activation, or absence thereof, results in two opposite fates:

- i. Upon ligand-receptor interaction and downstream signaling, Endophilin initiates cargo capture and local membrane curvature (Figure 2, Stage II to V), subsequent mechanisms are described below.
- ii. In the absence of receptor stimulation, local Cdc42 deactivation by GTPase-activating proteins (GAPs) RICH1, SH3BP1 and Oligophrenin induce the disassembly of the priming complex and abortion of the Endophilin patches [18] (Figure 2, Stage I). New clusters of Endophilin reform stochastically nearby, repeatedly priming the membrane for FEME.

Cargo selection

So far, FEME cargoes are a subset of the amine GPCRs (β 1- and α 2a-adrenergic receptors, Dopamine receptors 3 and 4 and Muscarinic Acetylcholine receptor 4), RTKs (EGF, HGF, VEGF, PDGF, NGF and IGF-1 receptors), cytokine receptors (tetrameric Interleukin receptor 2, IL-2R) and axon guidance receptors (Plexin A1 and ROBO1) [16,27,28]. In addition, the pathways is hijacked by cholera and Shiga toxins [17]. Many of these cargoes and toxins can enter cells through other endocytic pathways (CME, CLIC/GEEC, EGFR-NCE or macropinocytosis) [31–33]. To date, only the β 1-adrenergic receptor (β 1-AR) appears to rely exclusively on FEME for cellular entry [16,18,34]. However, inhibition of FEME induces the accumulation of many of these receptors at specific cellular locations (e.g. leading edge), even though their global uptake may not be affected [16].

In all instances studied so far, cargoes are sorted into FEME carriers upon direct or indirect binding to Endophilin (Figure 2, Stage II). The Endophilin SH3 domain binds directly to proline-rich regions present in intercellular loops of the identified GPCRs [16,35]. This contrasts with growth factor and axon guidance receptors, which are sorted through the binding of cytosolic adaptors. Both EGFR and HGFR are sorted upon binding of CIN85 to Endophilin (which are bridged to the receptors by Cbl) [36,37]. Recently, Plexin A1 and ROBO1 were found to bind to Endophilin through Collapsin Response Mediator Protein 4 (CRMP4) and srGAP1, respectively [27,28].

Membrane curvature and carrier formation

How membrane curvature is generated is still unclear. Endophilin and its N-BAR domain are required, as in their absence no FEME carriers are produced [16,18]. However, the transition between the priming patches of

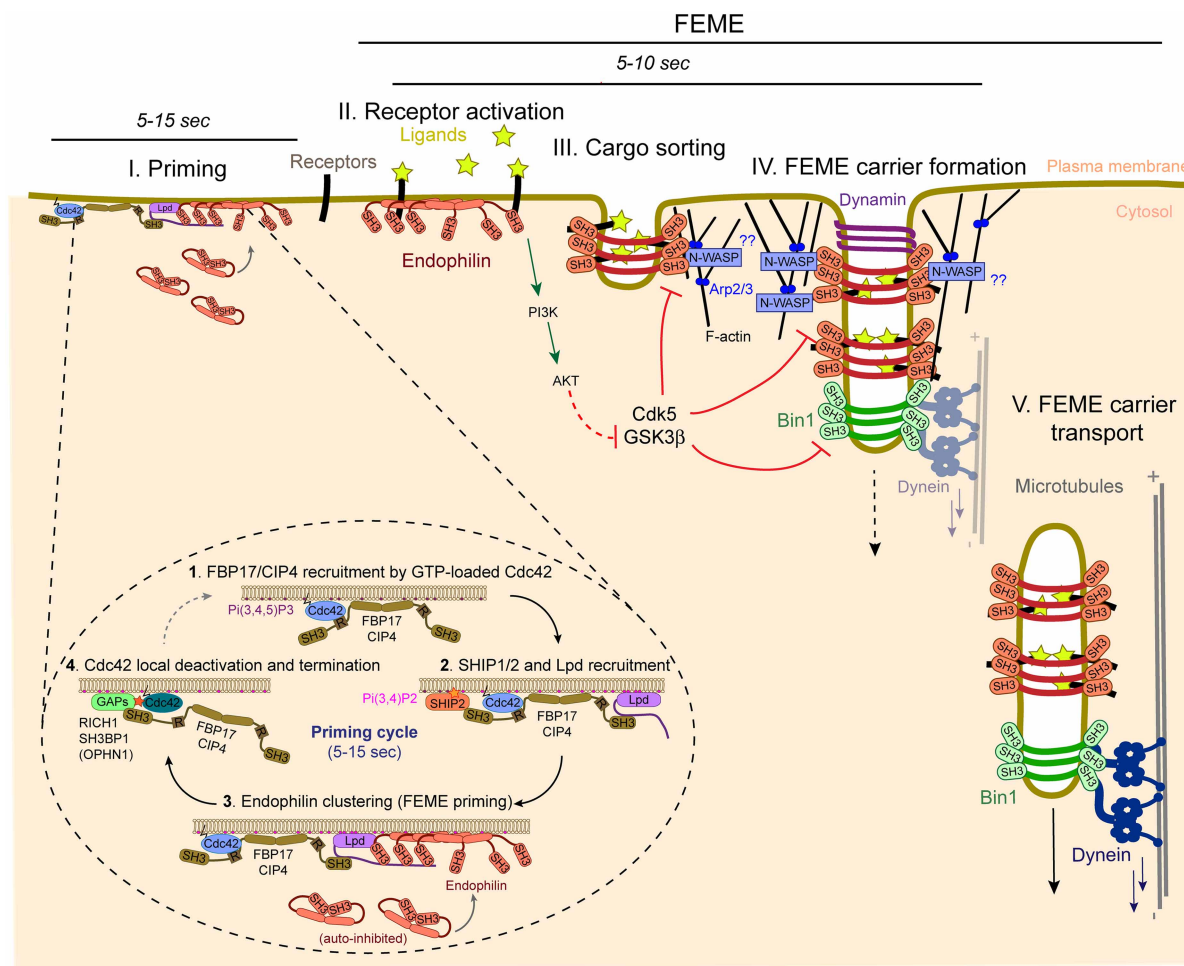


Figure 2. Molecular steps of FEME carrier formation.

Endophilin pre-enrichment on the plasma membrane (Stage I) is a prerequisite for FEME and mediates the promptness of the pathway to activate upon stimulation. FEME in itself only starts upon receptor activation and thus, corresponds to Stages II to V. Stage I. Plasma membrane priming cycle. Step 1, at high $\text{Pi}(3,4,5)\text{P}_3$ patches, active GTP-loaded Cdc42, recruits FBP17 and CIP4 through their REM domains (R). Step 2, FBP17 and CIP4 cluster 5'-phosphatases SHIP1 and 2 as well as Lamellipodin (Lpd) through their SH3 domains. Lpd is further stabilized by $\text{Pi}(3,4)\text{P}_2$ locally produced by SHIP1/2, Step 3, Endophilin is recruited from the cytosol (where it is autoinhibited) and concentrated by Lpd. From there, pre-enriched Endophilin mediates prompt FEME carrier formation upon cargo activation. In absence of cargo activation, the FEME priming complex aborts and disassembles (Step 4), upon local Cdc42 deactivation by the GAPs RICH1, SH3BP1 and Oligophrenin (OPHN1). Stage II. FEME activation upon cargo stimulation. Activation of receptors by their ligands stabilizes pre-enriched Endophilin patches and starts FEME. The exact signals driving this stage are still unclear. However, stages II to IV are inhibited by Cdk5 and GSK3 β , which hold off FEME. Upon receptor activation, PI3K, AKT and ERK signaling inactivate GSK3 β , thereby releasing its inhibitory action and activating FEME. Stage III. Cargo sorting. Activated receptors are recruited to nascent FEME pits either through direct binding to Endophilin (e.g. β 1 adrenergic receptor) or through binding of adaptor proteins (e.g. Cbl–CIN85 complex recruiting EGFR). Stage IV. FEME carrier formation. Local addition of Endophilin molecules by activated cargoes trigger membrane curvature, likely supported by local actin polymerization. Dynamin is recruited by Endophilin but excluded from the main body of the tubules, likely to avoid premature membrane scission. Detachment of FEME carrier is achieved by the synergy of GTP-dependent membrane pinching by Dynamin, membrane tension imposed by local actin cytoskeleton and friction-driven scission (FDS) lipid diffusion barrier by the N-BAR domains of Endophilin. Stage V. FEME carrier transport. Swiftly after budding, FEME carriers are rapidly transported on microtubules by Dynein, recruited onto the tubules by Bin1. The precise timing of Dynein loading on FEME carriers (Stage IV or V) is not yet clear.

Endophilin (which do not appear to generate meaningful curvature) and the shaping of tubules and vesicles following receptor activation is not understood. Either the priming clusters do not reach a critical local concentration of Endophilin required to deform the plasma membrane — and receptor activation boosts the number of Endophilin molecules over the threshold. Or, a so far unidentified mechanism is controlling membrane curvature by the N-BAR domain until activation. The recent finding that Bin1, another N-BAR protein is present on virtually all FEME carriers [28], opens the possibility of a third alternative, whereby the number of Endophilin and Bin1 molecules matter. However, Bin1 is also present on most priming patches [28], thus its recruitment is not the trigger in itself. Actin polymerization, which is required for carrier production [16], may have a role in inducing or stabilizing membrane curvature during tubule formation (Figure 2, Stage III). But isolating its role during priming (potential corralling of the patches), membrane curvature formation, membrane scission and potential short distance cytosolic transport is technically challenging.

Shiga and cholera toxins, which induce inward membrane bending from outside of the cells [38], revealed that if the initial curvature is provided, it is sufficient for Endophilin to take over and mediate the next steps (membrane scission and carrier transport) [17]. Thus, perhaps the number of Endophilin molecules recruited on toxin-induced tubules is lower than that on canonical FEME carriers, and the threshold hypothesis is the main driver.

Membrane scission

Detachment of the carriers requires the co-ordinated action of Dynamin, actin and N-BAR domain of Endophilin (and perhaps Bin1) [17] (Figure 2, Stage IV). Dynamin is a GTPase that mediates membrane scission in a large number of endocytic processes, including CME [39]. It is recruited to the neck of endocytic carriers by the SH3 domains of several BAR domain proteins, including Endophilin and Bin1 [22,40]. There, the co-ordinated hydrolysis of GTP induces conformation changes that drive membrane fission [41]. Because Endophilin can also inhibit Dynamin activity [42], local stoichiometry is likely key in determining the location of the scission event (Figure 2, Stage IV). By locally blocking Dynamin, Endophilin might avoid premature cuts and allow the forming tubules to grow and package enough cargoes before detachment. It is now clear that Dynamin is necessary but not sufficient, and a synergy with local membrane tension provided by local actin scaffold and friction-driven scission (FDS) by N-BAR domains is required [17,19]. The FDS mechanism arises from the frictional barrier for lipid diffusion created by oligomerized BAR domains [19] (Figure 2, Stage IV). One remaining question is whether forces induced by Dynein are required for membrane scission. *In vitro*, typical scission assays use molecular motors (in that case Kinesins) to pull membrane tubes away from giant unilamellar vesicles (GUVs), thereby mimicking the neck of a forming endocytic carrier. However, when exactly is Dynein recruited onto FEME carriers and how much is it required for membrane scission *in vivo* is not clear yet.

Cytosolic transport

Actin polymerization is required for FEME carrier formation and perhaps for the first few microns displacement into the cytosol. Endophilin binds to N-WASP and to the WAVE complex [18,43], but more work is needed to understand the molecular events recruiting actin to budding FEME carriers. As for other CIE carriers, Dynein powers long-range and fast movements of FEME carriers on microtubules [17,28,44] (Figure 2, Stage V). Bin1 recruits Dynein to the carriers [28] but molecular details are lacking to date.

Regulatory mechanisms

Because FEME is not constitutively active, many regulatory mechanisms must exist to hold it off and activate it on demand. This is also apparent in the diversity in FEME activity displayed by different cell types. While ligand binding triggers internalization of cargo receptors, some cell types present robust spontaneous FEME, which is apparent in resting cells that were not activated by any ectopic ligand [28]. This is due to high levels of growth factors in their regular culture media. Because such spontaneous FEME is lowered by serum starvation (removal of growth factors) and, conversely, activated upon stimulation with extra serum [16,28], it suggests a link between some cell signaling events and FEME activity.

The levels of $\text{Pi}(3,4)\text{P}_2$, and that of the sequential precursors $\text{Pi}(4,5)\text{P}_2$ and $\text{Pi}(3,4,5)\text{P}_3$, are tightly controlled [45,46]. As such, phosphoinositides kinases and phosphatases that balance their production are regulating FEME. By producing the local $\text{Pi}(3,4)\text{P}_2$ required for the priming mechanism, SHIP1 and 2 phosphatases have a direct activating effect on the pathway. Upstream, class I PI3 kinases are also positive regulators because they

boost the levels of $\text{Pi}(3,4,5)\text{P}_3$, which is required for $\text{Pi}(3,5)\text{P}_2$ production by SHIP1/2 [46,47]. At the reverse, PTEN and Synaptojanin are negative regulators because they reduce local levels of $\text{Pi}(3,4,5)\text{P}_3$ and $\text{Pi}(4,5)\text{P}_2$, respectively [46,48]. Beyond phosphoinositides, FEME is also controlled by several phosphorylations imposed on cargoes, Dynamin and Dynein by Cdk5 and GSK3 β [28]. Cdk5 and GSK3 β are well known to play important roles in regulating endocytosis. They do so by phosphorylating Amphiphysin, Dynamin and other endocytic proteins in synapses [49]. Dynamin-1 is phosphorylated at Ser778 by Cdk5, followed by that of Ser774 by GSK3 β [50,51]. The phosphorylation of Ser778 blocks the recruitment of Dynamin-1 by binding partners such as Endophilin, and that of Ser774 inhibits Dynamin-1 activity. Interestingly, these phosphorylations dampen dysregulated CME but activate ADBE [51,52], suggesting they might regulate a balance between the pathways. However, it is not known whether Dynamin-1 directly controls a cross-talk between FEME and CME. Both pathways also use Dynamin-2, which does not have a precise conservation of the Cdk5 and GSK3 β phosphorylation sites and does not appear to be under the same control as Dynamin-1 [52]. But if acute activation of Dynamin-1 upon the inhibition of GSK3 β triggers rapid, dysregulated CME [52], it appears to simply enhance FEME in its regular form [28]. Because Endophilin is phosphorylated by Src, ROCK and LRRK2 and FEME is blocked upon PAK1 and 2 inhibition [16,53–55], additional regulatory mechanisms remain to be understood. Of particular interest are the phosphorylations of Endophilin in its amphipatic Helix 0 (Thr14) by ROCK and Helix 1 (Thr73 and Ser75) by LRRK2, which hamper its membrane-binding ability [54–57]. However, to what extent this mechanism controls FEME remains to be investigated.

Similarities and differences with other CIE pathways

Characteristics and mechanisms of endocytic carrier formation by CIE pathways have been reviewed elsewhere [7–9,12]. We focus here on highlighting analogies and disparities between FEME and other known CIE routes.

Dynamin dependency

Because efficient mutants and excellent small compound inhibitors that block Dynamin exist, the requirement for an endocytic pathway is easy to establish and makes for a clear characteristic. The reliance on Dynamin splits the CIE pathways into two groups (Figure 1). FEME shares this dependency with the IL2R β uptake and EGFR-NCE, as well as ADBE and UFE in neurons. Even though FEME shares some similarities with UFE [58], it is clearly different phenomenologically or mechanistically to the other pathways [6,7] and thus, Clathrin-independence but Dynamin-dependency is not sufficient to identify FEME.

Constitutive or triggered

A cardinal feature of FEME is its non-constitutive activity. It needs a signal to be triggered: receptors to be activated by their cognate ligands [16] (Figure 2, Stage II). This is shared with many other CIE pathways, which also are not constitutively active. CLIC/GEEC appears constitutive because it is detected in resting cells without any external stimuli [59]. However, it is inhibited by increased membrane tension and enhanced by the extracellular clustering of GPI-anchored proteins, or glycoproteins or glycolipids by galectins, toxins and viruses [60]. Although at different signal strengths, macropinocytosis and EGFR-NCE are activated upon growth factor stimulation [33,61], and ADBE and UFE are triggered upon axon depolarization [62,63]. Only the uptake of IL2R β and γ chains, and that of inactive GPCRs occur through constitutive Clathrin-independent routes [64,65].

Role of actin

Actin polymerization is required for all CIE pathways but MEND (which appears to prevent it) [13,66]. However, molecular events controlling actin dynamics vary widely between different pathways. The main discriminators are the small G proteins RhoA, Rac1 and Cdc42, as they control different types of actin polymerization (branched or bundled). For example, the CLIC/GEEC pathway is Cdc42-dependent, whereas IL2R β uptake relies on RhoA and Rac1 (reviewed in [66]). CLIC maturation involves Cdc42- and N-WASP-mediated local actin polymerization, with Cdc42 being spatially and temporally controlled by ARHGAP10 [67]. Local generation of F-actin around clustered IL2R β or γ chains is promoted through the direct binding of the WAVE complex to the receptors, via WIRS motifs present in their cytoplasmic tails [68]. The receptors also induce PI3K-mediated enrichment of $\text{Pi}(3,4,5)\text{P}_3$, followed by the sequential recruitment of RhoA, Rac1, Pak1, WAVE and N-WASP to mediate local cytoskeleton remodeling and membrane protrusions [69,70]. FEME is atypical

in that it needs all three small G proteins, although probably at different time. It is clear that Cdc42 is required for priming, but nothing is known about RhoA and Rac1, other than blocking their activity inhibits FEME [16].

Cargo specificity

Cargoes that enter cells independently of CME have been long used to identify and label CIE pathways [8,9,71]. As such, CLIC/GEEC, IL2R β uptake and EGFR-NCE are cargo specific, and there is a good understanding of the molecular steps leading to cargo selection [7] (Figure 1). Some other CIE processes, such as macropinocytosis, MEND, ADBE or UFE do not have known cargo-sorting mechanisms and internalize any proteins located on the patches of membrane being engulfed [7] (Figure 1). All internalized transmembrane proteins are then sorted in endosomes: recycled back to the cell surface or degraded in lysosomes. FEME is part of the first group (cargo-specific) because of the mechanisms explained above. However, it appears to have the particularity — only shared by IL2R β uptake — of internalizing only one type of cargo per carrier [16]. But studies carefully measuring cargo stoichiometry upon co-stimulation by several ligands are lacking to definitely conclude on that property.

Shapes and speed of carrier formation

Morphologically, FEME carriers most resemble CLICs, which are tubules that often adopt a ‘C’ shape in the cytoplasm [72]. However, FEME tubules are more pleiotropic, as their length can vary from 100 nm to several microns [16]. UFE and IL2R β uptake produce rounded, uncoated vesicles of 60 nm and 500 nm in diameter, respectively [62,68], whereas ADBE, MEND and macropinocytosis generate larger vacuoles (from 500 nm to 10–20 μ m in diameter). The speed of carrier formation places FEME between UFE, which form vesicles within 50 to 100 ms from stimuli, and other, slower, CIE processes.

How to identify FEME

Establishing that a receptor or a ligand enters through FEME requires more than just inhibiting Endophilin. This is because of its additional role in CME [21], as well in potentially unidentified and unrelated processes. High confidence identification requires the combination of: (i) providing direct evidence that the candidate cargo is internalized into FEME carriers, (ii) showing that such uptake is triggered by ligand addition and (iii) that inhibition of Endophilin blocks cargo uptake [16].

Importantly, details of the protocol used are critical. Historically, uptake assays have been performed following a 30 min to 1 h pre-incubation at 4°C, prior to switching to 37°C for various amounts of time. This is to pre-bind ligands to their receptors exposed at the cell surface and to synchronize endocytosis. Other protocols call for uptake at room temperature instead of 37°C. However, these practices reduce membrane fluidity and may induce a phase transition, which will likely not reverse within the time frame of fast endocytic processes. Consistently, both UFE in synapses (which is Endophilin-dependent [58]) and FEME are only observed at physiological temperature (37°C for mammalian cells) [16,62]. An artificial increase in membrane fluidity by increasing the proportion of polyunsaturated lipids boosts Endophilin-mediated uptake [73].

Another step used to boost endocytosis and facilitate measurements is serum starvation for several hours (up to overnight) prior to endocytic assays. This was historically done to artificially increase the levels of receptors at the cell surface for early electron microscopy studies [74]. This practice has been routinely used in many experiments since. However, FEME is inhibited upon serum starvation, even as short as 30 min [16]. This is because growth factor removal increases GSK3 β activity by depressing AKT and ERK-mediated inhibitory phosphorylation on Ser 9 [75]. Together with Cdk5, GSK3 β negatively regulates FEME and thus its activation by serum starvation has lasting depression effects on FEME [28]. In addition, the artificial accumulation of receptors at the cell surface caused by serum starvation can change their mode of internalization and might favour their uptake by pathways that are not normally used [76]. Thus, to identify and measure FEME, it is critical to work in physiological conditions, without any serum starvation or temperature shifts.

To compensate for low and unsynchronized ligand binding to their receptors, direct incubation times at 37°C must be longer than the actual endocytic events. We found that 4 to 5 min incubation times to be sufficient for trigger FEME by all ligands tested to date. This does not mean that individual endocytic events take that long, as most of the time is used by the ligands to diffuse in the medium, to bind to their receptors and to activate them. Indeed, direct observation showed that individual FEME carriers take 5 to 15 s to form [16]. In line with the above considerations, warm fixation (e.g. pre-warmed PFA incubated at 37°C for the first 10 min) best preserves FEME carriers.

A word of caution should be said about ectopic expression of Endophilin and cargoes. Elevated levels will change the stoichiometry between FEME components and may induce artifactual events. Endophilin is auto-inhibited in the cytosol and opens up upon the binding of its SH3 domain to PRMs (of cargoes, Dynamin etc.) [18,22]. Higher levels of expression will induce an excessive membrane tubulation by the BAR domain and destabilize the balance required for FEME carrier formation. Likewise, overexpressing a receptor bears the risk of vastly outnumbering endogenous copies (most receptors are expressed at very low copy numbers [77]) and 'leaking' into non-natural endocytic routes. Endogenous tagging with fluorescent proteins using gene editing is an option and was performed successfully for Endophilin in Hep2 β cells [23], save for the risk of dim signals and potential steric hindrance of the tag. Until now, our technique of choice has been the immunostaining of endogenous Endophilin and cargoes. Ideally, recombinant ligands or antibodies recognizing the ectodomains of endogenous cargo receptors (exposed to the media) should be added to the cells and their enrichment into FEME carriers determined by co-immunostaining of Endophilin post fixation [16]. Cargoes internalized into FEME carriers can be detected by confocal microscopy or electron microscopy.

To date, only genetic means can be used to block FEME with some specificity. Genetic inactivation of Endophilin using triple knock-down (TKD) or knock-out, or overexpression of the inactive mutant Δ H0-BAR domain works well [16], with the caveat mentioned above that CME (and perhaps other cellular processes) might be affected. The Δ H0-BAR mutant is the strict BAR domain (of either Endophilin A1, 2 or 3), lacking the first amphipathic helix, middle region and SH3 domain. It binds very poorly to membranes and should not bind to any interacting partners, but still heterodimerises with endogenous Endophilin [16]. Thus, it imposes a dominant-negative effect upon overexpression as it titrates out endogenous Endophilin and sequesters it in the cytosol. Inhibition of the priming mechanism that pre-enriches Endophilin at the plasma membrane before receptor activation is another strategy. This is achieved by FBP17, CIP4 and TOCA-1 triple knock-down, SHIP1 and 2 double knock-down or Lamellipodin depletion [18]. Overexpression of the Cdc42 GAPs that terminate the priming cycle (RICH1, SH3BP1 or OPHN1) are also effective at depressing FEME [18].

Although many small compound inhibitors block FEME, none, so far, are specific to the pathway. Dynamin inhibitors (e.g. Dynasore, Dyngo 4a, Dynole 34.2, or Chlorpromazine [78–81]) are very effective at stopping FEME, but also block CME and other endocytic pathways. PI3K inhibitors work by inhibiting the production of $\text{Pi}(3,4,5)\text{P}_3$, and thus $\text{Pi}(3,4)\text{P}_2$, thereby abrogating the FEME priming mechanism. But PI3Ks work in many other cellular processes, including actin polymerization and parallel endocytic routes. Consistently, both actin poisons (Cytochalasin D, Latrunculin B, Jaspilakinolide) and inhibitors of actin regulators (RhoA, Rac, Arp2/3 inhibitors) block FEME as well as other pathways [66]. Because a dynamic cycle of activation-inhibition of Cdc42 drives the priming of FEME, Cdc42 inhibitors can either be stimulatory or inhibitory (depending on concentrations and incubation times) [16,18], and thus are not a reliable way of controlling the pathway. Amongst chemicals historically used to inhibit CME, macropinocytosis and other CIE pathways, it is important to realise that many block FEME as well [16]. Thus, the phenotypes observed could be caused by the defect in FEME, instead, or in addition to, the other endocytic pathways studied.

Amiloride, and its more soluble analog 5-(*N*-ethyl-*N*-isopropyl) amiloride (EIPA), are the gold standard for inhibiting macropinocytosis, but they also block FEME [16]. Because they are inhibitors of the Na^+/H^+ exchanger, they depolarize the plasma membrane, thereby destabilizing several small GTPases, including Rac and Cdc42 [82]. PAK1 and 2 inhibitors also perturb macropinocytosis, in addition to IL2R β uptake and FEME [16,69]. High sucrose (hypertonic medium), potassium depletion, MDC, CPZ or PAO stop FEME in addition to CME [16]. Likewise, drugs depleting or sequestering cholesterol (M β CD, filipin, nystatin or simvastatin) have been used in thousands of publications as sole evidence that cargoes (receptors, toxins, viruses or bacteria) enter cells through 'raft endocytosis'. However, because these drugs have pleiotropic effects — including abrogating FEME - some of these papers might need to be revised upon more rigorous investigation. Overall, small compound inhibitors should be used with caution in the study of FEME and cannot be the sole basis for conclusion. They must be complemented by other evidence, as explained above.

Means to boost FEME include hyperactivating the priming cycle by (moderately) overexpressing CIP4 or FBP17, depleting RICH1+SH3BP1 or inhibiting PTEN (RNAi or small compounds) [16,18]. It can also be achieved by relieving the negative regulation by Cdk5 and GSK3 β (RNAi or small compounds) [28]. Overexpression of Endophilin itself does not stimulate well FEME, as explained above.

Finally, the choice of cells is important. Some cell lines such as HeLa or Hek293 display little FEME compared with other cell types such as normal human retinal pigment epithelial cells (RPE1), primary skin

fibroblasts or human umbilical vein endothelial cells (HUVEC) [28]. The differences in activity in different cell types can be 5 to 8 folds [28] and thus influence the ease of observing FEME.

Cargoes that may use FEME

Equipped with recent insight, we can review past literature on cargoes that are known to enter cells in an Endophilin-dependent manner and discuss which ones could be using FEME. Several papers identified the binding of Endophilin to cargoes and concluded that the latter must enter cells through CME, but we now know that this is not necessarily the case. Of particular interest are cargoes that interact with Endophilin in a ligand-dependent manner, as it is a cardinal feature of FEME. For example, both CD36 and scavenger receptor A (SR-A) were immuno-precipitated by Endophilin only upon oxidized low-density lipoprotein (oxLDL) treatment [83]. Depletion of Endophilin increased the cell surface levels of CD36 and SR-A and decreased oxLDL uptake in macrophages [83] (Table 1). CME was proposed to be the pathway on the basis of two inhibitors, but more detailed characterization is required to rule out FEME. Other stimuli-induced binding to cargoes may include the interaction of Endophilin A1 and A2 with voltage-gated Ca^{2+} channels (L, N and P/Q types) [84] (Table 1). They were reported to do so at resting Ca^{2+} concentration (100–300 nM) through their linker regions between the BAR and the SH3 domain [84]. Endophilin was also reported to interact with the Chloride ClC-3 channel [85], the small-conductance calcium activated $\text{K}_{\text{Ca}2.3}$ channel [86] and the gamma-aminobutyric acid GABA receptor B [87], which is a ligand-gated Chloride channel (Table 1), but these interactions have not been followed up to date. The binding of Endophilin to several ion channels is intriguing because it is not believed that channels are actively endocytosed, and even less so rapidly in a ligand- or stimuli-dependent manner.

Vesicular glutamate transporter 1 (VGLUT1) is known to bind directly to the SH3 domain of Endophilin through a PRM in its C-terminus that is absent from VGLUT2 or 3 [88,89] (Table 1). This is reminiscent of

Table 1 Cargoes that may use FEME to enter cells

Putative FEME cargoes	Binds to Endophilin?	Endophilin-dependent uptake?	Ligand-dependent uptake?	References
CD36	Yes	Yes	Yes (oxLDL)	[83]
Scavenger Receptor A	Yes	Yes	Yes (oxLDL)	[83]
voltage-gated Ca^{2+} channels	Yes	?	Yes (Ca^{2+})	[84]
ClC-3 channel	Yes	?	?	[85]
$\text{K}_{\text{Ca}2.3}$ channel	Yes	?	?	[86]
GABA receptor B	Yes	?	?	[87]
VGLUT1	Yes	Yes	?	[88]
AMPA	Yes	Yes	?	[91,93]
ADAM 9	Yes	?	?	[95]
ADAM 15	Yes	?	?	[95]
ADAM 19	Yes	?	?	[95]
MT1-MMP	?	Yes	?	[53]
Enterovirus 71	?	Yes	?	[96]
Cytomegalovirus	Yes	?	?	[97]
Moloney murine leukemia virus	Yes	?	?	[98]

Casamento and Boucrot Table 1. Cargoes that may use FEME to enter cells.

Receptors, channels, and pathogens and viruses that bind to Endophilin enter cells in an Endophilin-dependent manner and/or are internalized in a ligand-dependent manner are all putative FEME cargoes. oxLDL: oxidized low-density lipoprotein; voltage-gated Ca^{2+} channels (L, N and P/Q types); ClC-3: Chloride ClC-3 channel; $\text{K}_{\text{Ca}2.3}$ channel: small-conductance calcium activated $\text{K}_{\text{Ca}2.3}$ channel; GABA receptor B: gamma-aminobutyric acid GABA receptor B (a ligand-gated Chloride channel); VGLUT1: Vesicular glutamate transporter 1; AMPAR: α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor; ADAM9, 15 and 19: disintegrin and metalloproteinase domain-containing proteins 9, 15 and 19; MT1-MMP: membrane-type matrix metalloproteinase 1.

the binding of Endophilin to a PRM within a cytoplasmic loop of β 1- but not β 2- or β 3-adrenergic receptors [35]. The binding of VGLUT1 to Endophilin recruits the transporter to a fast endocytic pathway and synaptic vesicle recycling, thereby regulating neurotransmitter release and short-term plasticity [88,90]. Endophilin also binds to the α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor, either directly to the GluA1, but not GluA2 subunit, or through the adaptor Arc/Arg3.1 [91–93] (Table 1). The absence of Arc or Endophilin reduces AMPAR uptake and increases cell surface levels of the receptor, thereby affecting post-synaptic neuron plasticity and long-term depression. Although UFE — which is Endophilin-dependent as well — could be quickly recycling VGLUT1, this pathway has not been identified on the post post-synaptic side, where AMPA receptors are located. Thus, this opens the possibility that UFE and FEME function on the pre- and post-synaptic sides, respectively. However, before synapse formation, a Clathrin-independent, Endophilin-dependent endocytic pathway reminiscent to FEME controls rapid vesicle production at growth cones [94]. This is consistent with the recent finding that axon guidance receptors PlexinA1 and ROBO1 are FEME cargoes [28].

Endophilin binds to the disintegrin and metalloproteinase domain-containing proteins ADAM 9, 15 and 19, but not ADAM10, 12 or TACE [95] (Table 1). In addition, the Endophilin A2 SH3 domain interacts with a PRM within FAK, which supports the phosphorylation of Endophilin by Scr at Tyr315 [53]. The phosphorylation inhibits the binding of Endophilin to Dynamin as well the uptake of the membrane-type matrix metalloproteinase 1 (MT1-MMP) [53]. Elevated levels of MT1-MMP at the cell surface then increase extracellular matrix degradation and migration. Thus, Endophilin might sort metalloproteinases into FEME carriers. The prominence of FEME at the leading edges of migrating cells is coherent with that hypothesis.

In addition to endogenous channels and receptors, Endophilin is also known to bind to viral proteins or mediate the entry of viruses. Enterovirus 71 entry into cells requires Dynamin and Endophilin A2 (Table 1), in a pathway the authors called Endophilin-Mediated Endocytosis (EME), which may very well be the same as FEME [96]. Endophilin A2 also interacts with mouse cytomegalovirus egress protein pM50 [97] and Moloney murine leukemia virus Gag [98] (Table 1).

Finally, the recent identification of Bin1 as a FEME component opens up possibilities. Its SH3 domain binds to different PRMs to those of Endophilin and may sort different sets of cargoes into FEME carriers. So far, only integrin α 3 has been reported to bind to Bin1 [99], but it is not clear whether the interaction is linked to endocytosis. Because of the poor solubility of transmembrane proteins, they are underrepresented in immunoprecipitation and pull-down experiments performed with classical detergents (e.g. Triton X-100) and many Endophilin or Bin1 interactors may have escaped detection. Targeted efforts are required to identify new FEME cargoes.

Pending questions and future milestones

There are many questions that need to be answered to increase our fundamental understanding of FEME. In this section, we stand on recent literature to ask five questions, answers to which would constitute important milestones in our understanding of the molecular mechanism and physiological functions of FEME.

How are initiation, cargo sorting, curvature and scission co-ordinated?

Several mechanistic details about FEME carrier formation are still missing. First, it is not clear when membrane curvature starts. During initiation, Endophilin is assembled into transient and dynamic patches, but there is no detectable curvature at this stage by immuno-electron microscopy [16]. At this stage, F-BAR and N-BAR domain proteins FBP17 and CIP4, Bin1, SH3BP1 and RICH1 are also present at initiation patches (albeit with different timings of arrival) [18], but they do not appear sufficient to induce significant membrane curvature. It is only after receptor stimulation that membrane bending is observed and tubules form [16]. It could be that cargo sorting brings the local copy number of Endophilin over a threshold triggering membrane curvature. Interestingly, if membrane curvature is induced independently of Endophilin, as it is the case during Shiga and cholera toxin entry where the toxins induce membrane bending upon clustering on the extracellular leaflet of the plasma membrane [38], then the need for receptor activation is bypassed [17]. Endophilin then functions during membrane scission, together with Dynamin and actin [17]. Overexpression of Endophilin (and of other N-BAR domain proteins) has long been known to be sufficient to form membrane tubules [100,101]. However, because Endophilin clustering into the initiation spots is SH3- and not N-BAR-domain dependent [18], it is not clear when the N-BAR domains become engaged with the membrane. It is important to remember that ectopically expressed N-BAR domains heterodimerize with endogenous full-length Endophilin molecules [16].

Therefore, papers claiming that the N-BAR domain is sufficient on its own for membrane targeting and bending *in vivo* must be interpreted with caution. Precise measurements of the number of Endophilin and Bin1 molecules at each stage of FEME carrier formation would likely provide further information on the process.

The mechanism of FEME carrier detachment is better understood but the timing, stoichiometries and individual contributions from (i) the friction imposed by the N-BAR domains of Endophilin and Bin1, (ii) the membrane tension enforced by actin and Dynein, and (iii) the membrane scission activity of Dynamin need to be worked out. For example, the role of Dynamin is not entirely clear. It is obviously required as any genetic or chemical inhibition methods tested so far blocked FEME carrier budding [16,17]. But Dynamin is both recruited onto initiation platforms in resting cells and on forming FEME carrier upon stimulation [18]. It is possible that it has a dual function as it does during CME [102]. Moreover, Endophilin blocks Dynamin-mediated membrane fission at high local concentration [42], suggesting a mechanism to exclude Dynamin from forming tubules and restricts its localization to their extremities. The timing of recruitment and localization of Bin1 and Dynein on FEME carriers (are they localized at the opposite end of the tubules rather than at the plasma membrane?) will be important to figure out. Finally, the mechanism, orientation and timing of actin polymerization around budding FEME carriers will require detailed and careful work, but will complete the picture.

How are inhibitory phosphorylations removed?

FEME is held off by several phosphorylations imposed on cargoes, Dynein and Dynamin by Cdk5, GSK3 β , and on Endophilin by Src, ROCK, DYRK1A and LRRK2 [16,53–55]. The promptness of FEME activation upon stimulation suggests that phosphatases quickly erase inhibitory phosphorylations. However, it is not obvious which phosphatase(s) could be acting downstream of the FEME cargoes known to date, which are as diverse as G α_s - and G α_i -coupled GPCRs, RTKs, cytokine or cell guidance receptors. In addition, kinases other than Cdk5 are likely priming GSK3 β phosphorylation during FEME. In synapses, several endocytic proteins, including Endophilin, Dynamin and Amphiphysin, are targets of Cdk5 and GSK3 β [49,103]. Their acute dephosphorylation upon axon depolarization is mediated by the Calcineurin phosphatase, which is activated by the sudden Ca²⁺ rise. Swift removal of the inhibitory phosphorylations then quickly activates compensatory endocytosis [104,105]. In non-neuronal cells, it is not known whether Ca²⁺ controls FEME. But it does mediate NCE of EGFR through Reticulon-3-dependent ER-PM contact sites [106]. As FEME is very polarized (i.e. it occurs on discrete location on the plasma membrane), it is unlikely that it is activated by a global and diffuse Ca²⁺ rise. Local Ca²⁺ increases happen in confined environments, such as ER-PM or Mitochondria-PM contact sites. But so far, none of these were observed around FEME carriers, nor were their requirements tested.

Another hypothesis is that generic phosphatases constitutively remove the inhibitory phosphorylations and that the regulation is at the level of the kinases — as long as they are active and keep phosphorylating the targets, the pathway is blocked. As soon as one, or several, of the kinases are inhibited and stop adding phosphate groups, the balance shifts under the remaining action of the phosphatases. GSK3 β is an obvious candidate as it is inactivated upon the phosphorylation of Ser9 in its disordered N-terminus. The phosphorylation induces an auto-inhibition of GSK3 β , as it mimics a priming phosphorylated Serine, and occupies the docking site, thereby blocking interactions with substrates [75]. Many kinases that are activated by growth factor receptor signaling, including AKT and ERK, phosphorylate Ser9-GSK3 β [75,107]. In that model, receptor stimulation would trigger the activation of kinase(s) that inactivate GSK3 β , thus relieving the negative pressure on FEME. Constitutive phosphatases would then quickly remove key inhibitory phosphorylations and trigger FEME. This model is supported by the lower level of regulation on phosphatases and the constitutive activity of many of them. Deciphering between these two hypotheses should inform us on a cardinal feature of FEME.

What is the fate of the FEME cargoes?

The current understanding is that, whatever the endocytic pathways used to depart from the plasma membrane, all cargoes end up in early endosomes (defined as Rab5- and EEA1-positive endosomes). Some cargoes transit through APPL1 and 2-positive compartments before reaching early endosomes [108]. From there, cargoes are either sorted into late endosomes and lysosomes or recycled back to the plasma membrane, via few different paths [109]. The two options lead to radically different fates (degradation or recovery, respectively). The generic destination of cargoes that enter cells through FEME is not known yet. Shiga and cholera toxins escape the degrading pathway to reach the Golgi complex and endoplasmic reticulum [110]. Endogenous β 1

adrenergic receptor is found in LAMP-1-positive late endosomes and lysosomes 30 min post-stimulation and its cellular uptake is almost entirely dependent on FEME [16,18]. Endogenous EGFR receptor enters cells through FEME carriers and ends up into lysosomes upon stimulation with high (>50 ng/ml) EGF levels. But inhibition of FEME does not affect EGFR total uptake or its degradation, consistent with the existence of alternative endocytic routes (EGFR-NCE and macropinocytosis). The fate of other FEME cargoes has not been measured precisely to date. The exercise is complicated by the loss of Endophilin staining upon FEME carrier fusion with endosomes, owing to the transformation of Pi(3,4)P₂ into Pi(3)P by PTEN, and INPP4A and B phosphatases [45,111]. Following cargoes that enter cells through several parallel endocytic routes (such as EGFR) carries the risk of accounting for molecules that were endocytosed by other pathways than FEME.

But should all FEME cargoes end up being sent for degradation, one could ask about the need for a fast endocytic pathway for the very first step, as early endosome to lysosome maturation lasts at least 20 min [112,113]. An exciting hypothesis is whether FEME could deliver cargoes directly to late endosomes for expedited degradation, bypassing early endosomes. Indeed, in INPP4A and B depleted cells, Endophilin staining is found on LAMP1-positive late, not EEA1-positive early endosomes [16] and Endophilin binds to ESCRT-I components ALIX and Tsg101 [18,114]. Endophilin also recruits the ubiquitin ligase Cbl (through CIN85) to EGFR and HGFR [36,37], thereby potentially linking mono-ubiquitination (the ESCRT sorting signal) and FEME carrier formation. Alternatively, as Endophilin (both A and B subtypes) has been involved in autophagosome formation and protein degradation [115–117], it is tempting to speculate that FEME carriers mature into atypical autophagosomes for accelerated delivery into lysosomes. However, molecular details and direct evidence to support either hypothesis are lacking so far. Whatever the mechanism, future work uncovering the fate of FEME cargoes will set a milestone in our understanding of the pathway.

Is FEME involved in neurodegenerative diseases?

There is evidence that the two main FEME components Endophilin and Bin1 are involved in neurodegeneration, in particular Alzheimer's and Parkinson's diseases. Endophilin A1 levels are elevated in Alzheimer's patients and transgenic mouse models [118]. Consistently, EndoA1 overexpression augments cerebral amyloid- β and increases neuronal death [119] and its depletion prevents synaptic dysfunction induced by oligomeric amyloid- β [120]. In addition, LRRK2 is one of the small numbers of proteins shown to cause autosomal Parkinson's disease, mostly upon Gly2019Ser mutation that enhances its kinase activity [121]. LRRK2 phosphorylates Endophilin at Thr73 and Ser75, which are located in the N-BAR domain, and thus decreases membrane binding and affects synaptic vesicle endocytosis [55,56]. Thus, one could speculate that impairment of UFE and/or FEME, which are both Endophilin-dependent, play a role in Parkinson's disease. However, considering the implication of Endophilin proteins in autophagosome formation and the strong link between autophagy and neurodegeneration, additional defects might induce the diseases.

The recent identification of Bin1 as a FEME component [28] broadens the possibilities for a link between FEME and neurodegeneration. Bin1 is strongly linked to Alzheimer's disease, but through Tau and amyloid- β pathology [122,123]. As for Endophilin, Bin1 levels are increased in Alzheimer's patients and its depletion suppressed Tau-mediated neurotoxicity [124]. However, lower Bin1 levels promote propagation of Tau pathology by decreasing aggregate endocytosis and lysosomal degradation [125]. In addition, Bin1 depletion increased BACE1 (because of decreased lysosomal targeting and degradation), resulting in elevated amyloid- β production [126]. Although a link between FEME and neurodegenerative diseases is yet to be established, there are several indications that the pathway may be involved. Its study holds promises for deepening our understanding of the pathologies and for potential new therapeutic avenues.

Can FEME be targeted to potentiate anti-cancer treatments?

Endophilin is involved in several cancers. Its role appears complex, however, as both increased and decreased levels were measured in tumors. Moreover, Endophilin A2 (also known as extra eleven nineteen, EEN) is a fusion partner of the mixed-lineage leukemia protein (MLL) in human acute leukemia [127]. Endophilin loss of expression in bladder cancer correlates with tumor progression [128]. Consistent with a role for FEME in EGFR signaling [16], the stable silencing of Endophilin increases phosphorylation of AKT, GSK3 β , SFK and STAT3 after EGF stimulation, mimicking the signaling pattern in urothelial carcinoma [128]. Interestingly, Endophilin stimulates cell migration by binding to RacGEF TIAM1 and potentiates colon cancer metastasis [129]. Conversely, Endophilin depletion blunts cell migration [16,27], and may limit tumor spreading. In

addition, FEME priming proteins SHIP2, FBP17, CIP4, Lamellipodin and SH3BP1 all promote tumor metastasis in several types of cancer, including lung and breast cancer [130–135].

In other cancers, such as Human epidermal growth factor receptor 2 (HER2)-positive tumors, Endophilin levels are elevated [136]. Endophilin knock-down reduced HER2 uptake and elevated the levels of HER2 at the plasma membrane. This decreased AKT and ERK downstream signaling as well as migration and metastasis of HER2+ cancer cells. Interestingly, elevated HER2 levels at the cell surface upon Endophilin depletion made cells more sensitive to trastuzumab and trastuzumab-emtansine (T-DM1) antibody therapies [136]. As several receptors, including EGFR, accumulate at the cell surface upon FEME inhibition [16], the possibility that blocking the pathway may potentiate anti-cancer therapies is exciting. Recently, stopping endocytosis using Dynamin but not Clathrin inhibitors was shown to improve antibody-dependent cellular cytotoxicity (ADCC) to the anti-EGFR, anti-Her2 and anti-PD-L1 monoclonal antibodies (cetuximab, trastuzumab and avelumab, respectively) [137]. ADCC is mediated by the recognition of opsonized targets by FcγRs and natural killer (NK) cells, which kill their target using perforin and granzymes [138]. For ADCC to occur, the antigen-mAb complex must remain on the cell surface long enough to engage the interaction of the mAb Fc region and cytotoxic cells [139]. Interestingly, FEME was identified as the main endocytic pathway inhibited alongside CME in Dynamin-inhibited cells [137]. One of the Dynamin drugs, Prochlorperazine (PCZ) [80], has been used clinically in humans for many years and the efficacy and safety of the combination of PCZ with monoclonal antibody therapy was demonstrated in Phase 1B clinical trials [137]. PCZ blocks FEME [16] and was used to control epilepsy [140], chronic kidney disease [141], *Candida* and viral infections [142–144], opening the possibility that dampening FEME may be beneficial in treating cancers as well as other diseases.

Conclusion and perspectives

FEME may be a major Clathrin-independent, Dynamin-dependent endocytic pathway and may emerge as an important therapeutic target. FEME was not characterized until few years ago, perhaps because it is not constitutively active and because it is only observed under physiological conditions (i.e. at 37°C and full serum media). Thus, the molecular mechanism is still being worked out and many aspects — some of which were discussed in this review — remain to be understood. Breakthrough in its understanding would arise from the discovery of ubiquitously expressed cargo receptor(s) that can easily be assayed, as transferrin receptor is for CME. Considerable advances may also come from the identification of specific cytosolic markers, if such proteins exist. Endophilin A1 to A3 being simple proteins binding to membranes via their N-BAR domains and to PRM-containing proteins through their SH3 domains, they are unlikely to be only functioning in FEME. Consistently, they have been involved in other cellular processes such as CME, some type of autophagy [115–117], as well as exocytosis of neurosecretory vesicles [146–148]. There are also probably important functional differences between the A1, A2 and A3 subtypes, as suggested by the recent finding that Endophilin A3, but not A2, controls the Clathrin-independent and Dynamin-independent uptake of CD166, induced upon extracellular clustering by Galectin-8 [145]. Finally, further investigation of the physiological functions supported by FEME should inform on new strategies to treat some of the corresponding diseases. The non-constitutive and regulated nature of the pathway is likely to be a precious asset to activate or inhibit on demand certain specific cellular processes, for the benefit of novel medical treatments.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Open Access

Open access for this article was enabled by the participation of University College London in an all-inclusive *Read & Publish* pilot with Portland Press and the Biochemical Society under a transformative agreement with JISC.

Funding

A.C. was supported by a Biotechnology and Biological Sciences Research Council (BBSRC) LIDo PhD scholarship. E.B. was a Lister Institute Research Fellow and a recipient of a BBSRC Pathfinder grant [BB/R01552X] and a Birkbeck/Wellcome Trust Institutional Strategic Support Fund (ISSF2) Career Development Award.

Abbreviations

ADBE, activity-dependent bulk endocytosis; ADCC, antibody-dependent cellular cytotoxicity; AMPA, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; AR, adrenergic receptor; CIE, clathrin-independent endocytosis; CLIC, clathrin-independent carriers; CME, clathrin-mediated endocytosis; EGFR, epidermal growth factor receptor; EGFR-NCE, EGFR non-clathrin endocytosis; FDS, friction-driven scission; FEME, Fast Endophilin-Mediated Endocytosis; GABA, gamma-aminobutyric acid; GAPs, GTPase-activating proteins; HER2, human epidermal growth factor receptor 2; MEND, massive endocytosis; MT1-MMP, membrane-type matrix metalloproteinase 1; PCZ, prochlorperazine; RTK, receptor tyrosine kinase; SH3, SRC Homology 3; UFE, ultrafast endocytosis; VGLUT1, vesicular glutamate transporter 1.

References

- 1 McMahon, H.T. and Boucrot, E. (2011) Molecular mechanism and physiological functions of clathrin-mediated endocytosis. *Nat. Rev. Mol. Cell Biol.* **12**, 517–533 <https://doi.org/10.1038/nrm3151>
- 2 Kaksonen, M. and Roux, A. (2018) Mechanisms of clathrin-mediated endocytosis. *Nat. Rev. Mol. Cell Biol.* **19**, 313–326 <https://doi.org/10.1038/nrm.2017.132>
- 3 Bitsikas, V., Correa, Jr, I.R. and Nichols, B.J. (2014) Clathrin-independent pathways do not contribute significantly to endocytic flux. *eLife* **3**, e03970 <https://doi.org/10.7554/eLife.03970>
- 4 Mayor, S., Parton, R.G. and Donaldson, J.G. (2014) Clathrin-independent pathways of endocytosis. *Cold Spring Harb. Perspect. Biol.* **6**, a016758 <https://doi.org/10.1101/cshperspect.a016758>
- 5 Johannes, L., Wunder, C. and Shafaq-Zadah, M. (2016) Glycolipids and lectins in endocytic uptake processes. *J. Mol. Biol.* **S0022-2836**, 30453-3 <https://doi.org/10.1016/j.jmb.2016.10.027>
- 6 Watanabe, S. and Boucrot, E. (2017) Fast and ultrafast endocytosis. *Curr. Opin. Cell Biol.* **47**, 64–71 <https://doi.org/10.1016/j.ccb.2017.02.013>
- 7 Ferreira, A.P.A. and Boucrot, E. (2018) Mechanisms of carrier formation during clathrin-independent endocytosis. *Trends Cell Biol.* **28**, 188–200 <https://doi.org/10.1016/j.tcb.2017.11.004>
- 8 Sandvig, K., Kavaliuskiene, S. and Skotland, T. (2018) Clathrin-independent endocytosis: an increasing degree of complexity. *Histochem. Cell Biol.* **150**, 107–118 <https://doi.org/10.1007/s00418-018-1678-5>
- 9 Thottacherry, J.J., Sathe, M., Prabhakara, C. and Mayor, S. (2019) Spoiled for choice: diverse endocytic pathways function at the cell surface. *Annu. Rev. Cell Dev. Biol.* **35**, 55–84 <https://doi.org/10.1146/annurev-cellbio-100617-062710>
- 10 Parton, R.G. (2018) Caveolae: structure, function, and relationship to disease. *Annu. Rev. Cell Dev. Biol.* **34**, 111–136 <https://doi.org/10.1146/annurev-cellbio-100617-062737>
- 11 Rewatkar, P.V., Parton, R.G., Parekh, H.S. and Parat, M.O. (2015) Are caveolae a cellular entry route for non-viral therapeutic delivery systems? *Adv. Drug Deliv. Rev.* **91**, 92–108 <https://doi.org/10.1016/j.addr.2015.01.003>
- 12 Mathew, M.P. and Donaldson, J.G. (2019) Glycosylation and glycan interactions can serve as extracellular machinery facilitating clathrin-independent endocytosis. *Traffic* **20**, 295–300 <https://doi.org/10.1111/tra.12636>
- 13 Hilgemann, D.W., Lin, M.J., Fine, M. and Deisl, C. (2020) On the existence of endocytosis driven by membrane phase separations. *Biochim. Biophys. Acta* **1862**, 183007 <https://doi.org/10.1016/j.bbame.2019.06.006>
- 14 Johannes, L. (2017) Shiga toxin-A model for glycolipid-dependent and lectin-driven endocytosis. *Toxins (Basel)* **9**, 340 <https://doi.org/10.3390/toxins9110340>
- 15 Yamauchi, Y. and Helenius, A. (2013) Virus entry at a glance. *J. Cell Sci.* **126**, 1289–1295 <https://doi.org/10.1242/jcs.119685>
- 16 Boucrot, E., Ferreira, A.P., Almeida-Souza, L., Debard, S., Vallis, Y., Howard, G. et al. (2015) Endophilin marks and controls a clathrin-independent endocytic pathway. *Nature* **517**, 460–465 <https://doi.org/10.1038/nature14067>
- 17 Renard, H.F., Simunovic, M., Lemiere, J., Boucrot, E., Garcia-Castillo, M.D., Arumugam, S. et al. (2015) Endophilin-A2 functions in membrane scission in clathrin-independent endocytosis. *Nature* **517**, 493–496 <https://doi.org/10.1038/nature14064>
- 18 Chan Wah Hak, L., Khan, S., Di Meglio, I., Law, A.L., Lucken-Ardjomande Hasler, S., Quintaneiro, L.M. et al. (2018) FBP17 and CIP4 recruit SHIP2 and lamellipodin to prime the plasma membrane for fast endophilin-mediated endocytosis. *Nat. Cell Biol.* **20**, 1023–1031 <https://doi.org/10.1038/s41556-018-0146-8>
- 19 Simunovic, M., Manneville, J.B., Renard, H.F., Evergren, E., Raghunathan, K., Bhatia, D. et al. (2017) Friction mediates scission of tubular membranes scaffolded by BAR proteins. *Cell* **170**, 172–184 e111 <https://doi.org/10.1016/j.cell.2017.05.047>
- 20 Kjaerulff, O., Brodin, L. and Jung, A. (2011) The structure and function of endophilin proteins. *Cell Biochem. Biophys.* **60**, 137–154 <https://doi.org/10.1007/s12013-010-9137-5>
- 21 Milosevic, I., Giovedi, S., Lou, X., Raimondi, A., Collesi, C., Shen, H. et al. (2011) Recruitment of endophilin to clathrin-coated pit necks is required for efficient vesicle uncoating after fission. *Neuron* **72**, 587–601 <https://doi.org/10.1016/j.neuron.2011.08.029>
- 22 Meinecke, M., Boucrot, E., Camdere, G., Hon, W.C., Mittal, R. and McMahon, H.T. (2013) Cooperative recruitment of dynamin and BIN/amphiphysin/Rvs (BAR) domain-containing proteins leads to GTP-dependent membrane scission. *J. Biol. Chem.* **288**, 6651–6661 <https://doi.org/10.1074/jbc.M112.444869>
- 23 Bertot, L., Grassart, A., Lagache, T., Nardi, G., Basquin, C., Olivo-Marin, J.C. et al. (2018) Quantitative and statistical study of the dynamics of clathrin-dependent and -independent endocytosis reveal a differential role of endophilinA2. *Cell Rep.* **22**, 1574–1588 <https://doi.org/10.1016/j.celrep.2018.01.039>
- 24 Taylor, M.J., Perrais, D. and Merrifield, C.J. (2011) A high precision survey of the molecular dynamics of mammalian clathrin-mediated endocytosis. *PLoS Biol.* **9**, e1000604 <https://doi.org/10.1371/journal.pbio.1000604>
- 25 Gallop, J.L., Jao, C.C., Kent, H.M., Butler, P.J., Evans, P.R., Langen, R. et al. (2006) Mechanism of endophilin N-BAR domain-mediated membrane curvature. *EMBO J.* **25**, 2898–2910 <https://doi.org/10.1038/sj.emboj.7601174>

- 26 Loll, P.J., Swain, E., Chen, Y., Turner, B.T. and Zhang, J.F. (2008) Structure of the SH3 domain of rat endophilin A2. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.* **64**, 243–246 <https://doi.org/10.1107/S1744309108007574>
- 27 Genet, G., Boye, K., Mathivet, T., Ola, R., Zhang, F., Dubrac, A. et al. (2019) Endophilin-A2 dependent VEGFR2 endocytosis promotes sprouting angiogenesis. *Nat. Commun.* **10**, 2350 <https://doi.org/10.1038/s41467-019-10359-x>
- 28 Ferreira, A.P.A., Casamento, A., Roas, S.C., Panambalana, J., Subramaniam, S., Schützenhofer, K. et al. (2020) Cdk5 and GSK3 β inhibit Fast Endophilin-Mediated Endocytosis. *bioRxiv* 2020.2004.20111.036863 <https://doi.org/10.1101/2020.04.11.036863>
- 29 Chen, Z., Chang, K., Capraro, B.R., Zhu, C., Hsu, C.J. and Baumgart, T. (2014) Intradimer/Intermolecular interactions suggest autoinhibition mechanism in endophilin A1. *J. Am. Chem. Soc.* **136**, 4557–4564 <https://doi.org/10.1021/ja411607b>
- 30 Vehlow, A., Soong, D., Vizcay-Barrena, G., Bodo, C., Law, A.L., Perera, U. et al. (2013) Endophilin, Lamellipodin, and Mena cooperate to regulate F-actin-dependent EGF-receptor endocytosis. *EMBO J.* **32**, 2722–2734 <https://doi.org/10.1038/emboj.2013.212>
- 31 Massol, R.H., Larsen, J.E., Fujinaga, Y., Lencer, W.I. and Kirchhausen, T. (2004) Cholera toxin toxicity does not require functional Arf6- and dynamin-dependent endocytic pathways. *Mol. Biol. Cell.* **15**, 3631–3641 <https://doi.org/10.1091/mbc.e04-04-0283>
- 32 Lundmark, R., Doherty, G.J., Howes, M.T., Cortese, K., Vallis, Y., Parton, R.G. et al. (2008) The GTPase-activating protein GRAF1 regulates the CLIC/GEEC endocytic pathway. *Curr. Biol.* **18**, 1802–1808 <https://doi.org/10.1016/j.cub.2008.10.044>
- 33 Sigismund, S., Argenzio, E., Tosoni, D., Cavallaro, E., Polo, S. and Di Fiore, P.P. (2008) Clathrin-mediated internalization is essential for sustained EGFR signaling but dispensable for degradation. *Dev. Cell* **15**, 209–219 <https://doi.org/10.1016/j.devcel.2008.06.012>
- 34 Eichel, K., Jullie, D. and von Zastrow, M. (2016) beta-Arrestin drives MAP kinase signalling from clathrin-coated structures after GPCR dissociation. *Nat. Cell Biol.* **18**, 303–310 <https://doi.org/10.1038/ncb3307>
- 35 Tang, Y., Hu, L.A., Miller, W.E., Ringstad, N., Hall, R.A., Pitcher, J.A. et al. (1999) Identification of the endophilins (SH3p4/p8/p13) as novel binding partners for the beta1-adrenergic receptor. *Proc. Natl Acad. Sci. U.S.A.* **96**, 12559–12564 <https://doi.org/10.1073/pnas.96.22.12559>
- 36 Soubeyran, P., Kowanetz, K., Szymkiewicz, I., Langdon, W.Y. and Dikic, I. (2002) Cbl-CIN85-endophilin complex mediates ligand-induced downregulation of EGF receptors. *Nature* **416**, 183–187 <https://doi.org/10.1038/416183a>
- 37 Petrelli, A., Gilestro, G.F., Lanzardo, S., Comoglio, P.M., Migone, N. and Giordano, S. (2002) The endophilin-CIN85-Cbl complex mediates ligand-dependent downregulation of c-Met. *Nature* **416**, 187–190 <https://doi.org/10.1038/416187a>
- 38 Romer, W., Berland, L., Chambon, V., Gaus, K., Windschiagl, B., Tenza, D. et al. (2007) Shiga toxin induces tubular membrane invaginations for its uptake into cells. *Nature* **450**, 670–675 <https://doi.org/10.1038/nature05996>
- 39 Ramachandran, R. and Schmid, S.L. (2018) The dynamin superfamily. *Curr. Biol.* **28**, R411–R416 <https://doi.org/10.1016/j.cub.2017.12.013>
- 40 Sundborger, A., Soderblom, C., Vorontsova, O., Evergren, E., Hinshaw, J.E. and Shupliakov, O. (2011) An endophilin-dynamin complex promotes budding of clathrin-coated vesicles during synaptic vesicle recycling. *J. Cell Sci.* **124**, 133–143 <https://doi.org/10.1242/jcs.072686>
- 41 Ford, M.G.J. and Chappie, J.S. (2019) The structural biology of the dynamin-related proteins: new insights into a diverse, multitasking family. *Traffic* **20**, 717–740 <https://doi.org/10.1111/tra.12676>
- 42 Hohendahl, A., Talledge, N., Galli, V., Shen, P.S., Humbert, F., De Camilli, P. et al. (2017) Structural inhibition of dynamin-mediated membrane fission by endophilin. *eLife* **6**, e26856 <https://doi.org/10.7554/eLife.26856>
- 43 Otsuki, M., Itoh, T. and Takenawa, T. (2003) Neural Wiskott-Aldrich syndrome protein is recruited to rafts and associates with endophilin A in response to epidermal growth factor. *J. Biol. Chem.* **278**, 6461–6469 <https://doi.org/10.1074/jbc.M207433200>
- 44 Day, C.A., Baetz, N.W., Copeland, C.A., Kraft, L.J., Han, B., Tiwari, A. et al. (2015) Microtubule motors power plasma membrane tubulation in clathrin-independent endocytosis. *Traffic* **16**, 572–590 <https://doi.org/10.1111/tra.12269>
- 45 Malek, M., Kielkowska, A., Chessa, T., Anderson, K.E., Barneda, D., Pir, P. et al. (2017) PTEN regulates PI(3,4)P2 signaling downstream of class I PI3K. *Mol. Cell* **68**, 566–580 e510 <https://doi.org/10.1016/j.molcel.2017.09.024>
- 46 Goulden, B.D., Pacheco, J., Dull, A., Zewe, J.P., Deiters, A. and Hammond, G.R.V. (2019) A high-avidity biosensor reveals plasma membrane PI(3,4)P2 is predominantly a class I PI3K signaling product. *J. Cell Biol.* **218**, 1066–1079 <https://doi.org/10.1083/jcb.201809026>
- 47 Batty, I.H., van der Kaay, J., Gray, A., Telfer, J.F., Dixon, M.J. and Downes, C.P. (2007) The control of phosphatidylinositol 3,4-bisphosphate concentrations by activation of the Src homology 2 domain containing inositol polyphosphate 5-phosphatase 2, SHP2. *Biochem. J.* **407**, 255–266 <https://doi.org/10.1042/BJ20070558>
- 48 McPherson, P.S., Garcia, E.P., Slepnev, V.I., David, C., Zhang, X., Grabs, D. et al. (1996) A presynaptic inositol-5-phosphatase. *Nature* **379**, 353–357 <https://doi.org/10.1038/379353a0>
- 49 Tomizawa, K., Sunada, S., Lu, Y.F., Oda, Y., Kinuta, M., Ohshima, T. et al. (2003) Cophosphorylation of amphiphysin I and dynamin I by Cdk5 regulates clathrin-mediated endocytosis of synaptic vesicles. *J. Cell Biol.* **163**, 813–824 <https://doi.org/10.1083/jcb.200308110>
- 50 Tan, T.C., Valova, V.A., Malladi, C.S., Graham, M.E., Berven, L.A., Jupp, O.J. et al. (2003) Cdk5 is essential for synaptic vesicle endocytosis. *Nat. Cell Biol.* **5**, 701–710 <https://doi.org/10.1038/ncb1020>
- 51 Clayton, E.L., Sue, N., Smillie, K.J., O’Leary, T., Bache, N., Cheung, G. et al. (2010) Dynamin I phosphorylation by GSK3 controls activity-dependent bulk endocytosis of synaptic vesicles. *Nat. Neurosci.* **13**, 845–851 <https://doi.org/10.1038/nn.2571>
- 52 Reis, C.R., Chen, P.H., Srinivasan, S., Aguet, F., Mettlen, M. and Schmid, S.L. (2015) Crosstalk between Akt/GSK3 β signaling and dynamin-1 regulates clathrin-mediated endocytosis. *EMBO J.* **34**, 2132–2146 <https://doi.org/10.15252/emboj.201591518>
- 53 Wu, X., Gan, B., Yoo, Y. and Guan, J.L. (2005) FAK-mediated src phosphorylation of endophilin A2 inhibits endocytosis of MT1-MMP and promotes ECM degradation. *Dev. Cell* **9**, 185–196 <https://doi.org/10.1016/j.devcel.2005.06.006>
- 54 Kaneko, T., Maeda, A., Takefuji, M., Aoyama, H., Nakayama, M., Kawabata, S. et al. (2005) Rho mediates endocytosis of epidermal growth factor receptor through phosphorylation of endophilin A1 by Rho-kinase. *Genes Cells* **10**, 973–987 <https://doi.org/10.1111/j.1365-2443.2005.00895.x>
- 55 Matta, S., Van Kolen, K., da Cunha, R., van den Bogaart, G., Mandemakers, W., Miskiewicz, K. et al. (2012) LRRK2 controls an endoA phosphorylation cycle in synaptic endocytosis. *Neuron* **75**, 1008–1021 <https://doi.org/10.1016/j.neuron.2012.08.022>
- 56 Arranz, A.M., Delbroek, L., Van Kolen, K., Guimaraes, M.R., Mandemakers, W., Daneels, G. et al. (2015) LRRK2 functions in synaptic vesicle endocytosis through a kinase-dependent mechanism. *J. Cell Sci.* **128**, 541–552 <https://doi.org/10.1242/jcs.158196>
- 57 Ambroso, M.R., Hegde, B.G. and Langen, R. (2014) Endophilin A1 induces different membrane shapes using a conformational switch that is regulated by phosphorylation. *Proc. Natl Acad. Sci. U.S.A.* **111**, 6982–6987 <https://doi.org/10.1073/pnas.1402233111>

- 58 Watanabe, S., Mamer, L.E., Raychaudhuri, S., Luvsanjav, D., Eisen, J., Trimbuch, T. et al. (2018) Synaptotagmin and endophilin mediate neck formation during ultrafast endocytosis. *Neuron* **98**, 1184–1197 e1186 <https://doi.org/10.1016/j.neuron.2018.06.005>
- 59 Howes, M.T., Kirkham, M., Riches, J., Cortese, K., Walsler, P.J., Simpson, F. et al. (2010) Clathrin-independent carriers form a high capacity endocytic sorting system at the leading edge of migrating cells. *J. Cell Biol.* **190**, 675–691 <https://doi.org/10.1083/jcb.201002119>
- 60 Lakshminarayan, R., Wunder, C., Becken, U., Howes, M.T., Benzing, C., Arumugam, S. et al. (2014) Galectin-3 drives glycosphingolipid-dependent biogenesis of clathrin-independent carriers. *Nat. Cell Biol.* **16**, 595–606 <https://doi.org/10.1038/ncb2970>
- 61 Yoshida, S., Pacitto, R., Sesi, C., Kotula, L. and Swanson, J.A. (2018) Dorsal ruffles enhance activation of Akt by growth factors. *J. Cell Sci.* **131**, jcs220517 <https://doi.org/10.1242/jcs.220517>
- 62 Watanabe, S., Rost, B.R., Camacho-Perez, M., Davis, M.W., Sohl-Kielczynski, B., Rosenmund, C. et al. (2013) Ultrafast endocytosis at mouse hippocampal synapses. *Nature* **504**, 242–247 <https://doi.org/10.1038/nature12809>
- 63 Clayton, E.L., Evans, G.J. and Cousin, M.A. (2008) Bulk synaptic vesicle endocytosis is rapidly triggered during strong stimulation. *J. Neurosci.* **28**, 6627–6632 <https://doi.org/10.1523/JNEUROSCI.1445-08.2008>
- 64 Lamaze, C., Dujancourt, A., Baba, T., Lo, C.G., Benmerah, A. and Dautry-Varsat, A. (2001) Interleukin 2 receptors and detergent-resistant membrane domains define a clathrin-independent endocytic pathway. *Mol. Cell* **7**, 661–671 [https://doi.org/10.1016/S1097-2765\(01\)00212-X](https://doi.org/10.1016/S1097-2765(01)00212-X)
- 65 Scarselli, M. and Donaldson, J.G. (2009) Constitutive internalization of G protein-coupled receptors and G proteins via clathrin-independent endocytosis. *J. Biol. Chem.* **284**, 3577–3585 <https://doi.org/10.1074/jbc.M806819200>
- 66 Hinze, C. and Boucrot, E. (2018) Local actin polymerization during endocytic carrier formation. *Biochem. Soc. Trans.* **46**, 565–576 <https://doi.org/10.1042/BST20170355>
- 67 Kumari, S. and Mayor, S. (2008) ARF1 is directly involved in dynamin-independent endocytosis. *Nat. Cell Biol.* **10**, 30–41 <https://doi.org/10.1038/ncb1666>
- 68 Basquin, C., Trichet, M., Vihinen, H., Malarde, V., Lagache, T., Ripoll, L. et al. (2015) Membrane protrusion powers clathrin-independent endocytosis of interleukin-2 receptor. *EMBO J.* **34**, 2147–2161 <https://doi.org/10.15252/embj.201490788>
- 69 Grassart, A., Dujancourt, A., Lazarow, P.B., Dautry-Varsat, A. and Sauvonnnet, N. (2008) Clathrin-independent endocytosis used by the IL-2 receptor is regulated by Rac1, Pak1 and Pak2. *EMBO Rep.* **9**, 356–362 <https://doi.org/10.1038/embor.2008.28>
- 70 Basquin, C., Malarde, V., Mellor, P., Anderson, D.H., Meas-Yedid, V., Olivo-Marin, J.C. et al. (2013) The signalling factor PI3K is a specific regulator of the clathrin-independent dynamin-dependent endocytosis of IL-2 receptors. *J. Cell Sci.* **126**, 1099–1108 <https://doi.org/10.1242/jcs.110932>
- 71 Maldonado-Baez, L., Williamson, C. and Donaldson, J.G. (2013) Clathrin-independent endocytosis: a cargo-centric view. *Exp. Cell Res.* **319**, 2759–2769 <https://doi.org/10.1016/j.yexcr.2013.08.008>
- 72 Kirkham, M., Fujita, A., Chadda, R., Nixon, S.J., Kurzchalia, T.V., Sharma, D.K. et al. (2005) Ultrastructural identification of uncoated caveolin-independent early endocytic vesicles. *J. Cell Biol.* **168**, 465–476 <https://doi.org/10.1083/jcb.200407078>
- 73 Pinot, M., Vanni, S., Pagnotta, S., Lacas-Gervais, S., Payet, L.A., Ferreira, T. et al. (2014) Lipid cell biology. Polyunsaturated phospholipids facilitate membrane deformation and fission by endocytic proteins. *Science* **345**, 693–697 <https://doi.org/10.1126/science.1255288>
- 74 Brown, M.S. and Goldstein, J.L. (1986) A receptor-mediated pathway for cholesterol homeostasis. *Science* **232**, 34–47 <https://doi.org/10.1126/science.3513311>
- 75 Cohen, P. and Frame, S. (2001) The renaissance of GSK3. *Nat. Rev. Mol. Cell Biol.* **2**, 769–776 <https://doi.org/10.1038/35096075>
- 76 Boucrot, E., Saffarian, S., Zhang, R. and Kirchhausen, T. (2010) Roles of AP-2 in clathrin-mediated endocytosis. *PLoS ONE* **5**, e10597 <https://doi.org/10.1371/journal.pone.0010597>
- 77 Hein, M.Y., Hubner, N.C., Poser, I., Cox, J., Nagaraj, N., Toyoda, Y. et al. (2015) A human interactome in three quantitative dimensions organized by stoichiometries and abundances. *Cell* **163**, 712–723 <https://doi.org/10.1016/j.cell.2015.09.053>
- 78 Macia, E., Ehrlich, M., Massol, R., Boucrot, E., Brunner, C. and Kirchhausen, T. (2006) Dynasore, a cell-permeable inhibitor of dynamin. *Dev. Cell* **10**, 839–850 <https://doi.org/10.1016/j.devcel.2006.04.002>
- 79 McCluskey, A., Daniel, J.A., Hadzic, G., Chau, N., Clayton, E.L., Mariana, A. et al. (2013) Building a better dynasore: the dyngo compounds potently inhibit dynamin and endocytosis. *Traffic* **14**, 1272–1289 <https://doi.org/10.1111/tra.12119>
- 80 Daniel, J.A., Chau, N., Abdel-Hamid, M.K., Hu, L., von Kleist, L., Whiting, A. et al. (2015) Phenothiazine-derived antipsychotic drugs inhibit dynamin and clathrin-mediated endocytosis. *Traffic* **16**, 635–654 <https://doi.org/10.1111/tra.12272>
- 81 Hill, T.A., Gordon, C.P., McGeachie, A.B., Venn-Brown, B., Odell, L.R., Chau, N. et al. (2009) Inhibition of dynamin mediated endocytosis by the dynoles—synthesis and functional activity of a family of indoles. *J. Med. Chem.* **52**, 3762–3773 <https://doi.org/10.1021/jm900036m>
- 82 Koivusalo, M., Welch, C., Hayashi, H., Scott, C.C., Kim, M., Alexander, T. et al. (2010) Amiloride inhibits macropinocytosis by lowering submembranous pH and preventing Rac1 and Cdc42 signaling. *J. Cell Biol.* **188**, 547–563 <https://doi.org/10.1083/jcb.200908086>
- 83 Huang, E.W., Liu, C.Z., Liang, S.J., Zhang, Z., Lv, X.F., Liu, J. et al. (2016) Endophilin-A2-mediated increase in scavenger receptor expression contributes to macrophage-derived foam cell formation. *Atherosclerosis* **254**, 133–141 <https://doi.org/10.1016/j.atherosclerosis.2016.10.009>
- 84 Chen, Y., Deng, L., Maeno-Hikichi, Y., Lai, M., Chang, S., Chen, G. et al. (2003) Formation of an endophilin-Ca²⁺ channel complex is critical for clathrin-mediated synaptic vesicle endocytosis. *Cell* **115**, 37–48 [https://doi.org/10.1016/S0092-8674\(03\)00726-8](https://doi.org/10.1016/S0092-8674(03)00726-8)
- 85 Liu, C.Z., Li, X.Y., Du, R.H., Gao, M., Ma, M.M., Li, F.Y. et al. (2016) Endophilin A2 influences volume-regulated chloride current by mediating ClC-3 trafficking in vascular smooth muscle cells. *Circ. J.* **80**, 2397–2406 <https://doi.org/10.1253/circj.CJ-16-0793>
- 86 Janbein, M., Quader, M.A., Hoppner, A.C., Gruner, I., Wanker, E., Walter, S. et al. (2014) Evidence for the interaction of endophilin a3 with endogenous Kca2.3 channels in PC12 cells. *Cell. Physiol. Biochem.* **34**, 474–490 <https://doi.org/10.1159/000363016>
- 87 Stelzl, U., Worm, U., Lalowski, M., Haenig, C., Brembeck, F.H., Goehler, H. et al. (2005) A human protein-protein interaction network: a resource for annotating the proteome. *Cell* **122**, 957–968 <https://doi.org/10.1016/j.cell.2005.08.029>
- 88 Voglmaier, S.M., Kam, K., Yang, H., Fortin, D.L., Hua, Z., Nicoll, R.A. et al. (2006) Distinct endocytic pathways control the rate and extent of synaptic vesicle protein recycling. *Neuron* **51**, 71–84 <https://doi.org/10.1016/j.neuron.2006.05.027>
- 89 De Gois, S., Jeanclos, E., Morris, M., Grewal, S., Varoqui, H. and Erickson, J.D. (2006) Identification of endophilins 1 and 3 as selective binding partners for VGLUT1 and their co-localization in neocortical glutamatergic synapses: implications for vesicular glutamate transporter trafficking and excitatory vesicle formation. *Cell. Mol. Neurobiol.* **26**, 679–693 <https://doi.org/10.1007/s10571-006-9054-8>

- 90 Weston, M.C., Nehring, R.B., Wojcik, S.M. and Rosenmund, C. (2011) Interplay between VGLUT isoforms and endophilin A1 regulates neurotransmitter release and short-term plasticity. *Neuron* **69**, 1147–1159 <https://doi.org/10.1016/j.neuron.2011.02.002>
- 91 Chowdhury, S., Shepherd, J.D., Okuno, H., Lyford, G., Petralia, R.S., Plath, N. et al. (2006) Arc/Arg3.1 interacts with the endocytic machinery to regulate AMPA receptor trafficking. *Neuron* **52**, 445–459 <https://doi.org/10.1016/j.neuron.2006.08.033>
- 92 Rial Verde, E. M., Lee-Osbourne, J., Worley, P. F., Malinow, R. and Cline, H. T. (2006) Increased expression of the immediate-early gene arc/arg3.1 reduces AMPA receptor-mediated synaptic transmission. *Neuron* **52**, 461–474 <https://doi.org/10.1016/j.neuron.2006.09.031>
- 93 Zhang, J., Yin, Y., Ji, Z., Cai, Z., Zhao, B., Li, J. et al. (2017) Endophilin2 interacts with GluA1 to mediate AMPA receptor endocytosis induced by oligomeric amyloid-beta. *Neural Plast.* **2017**, 8197085 <https://doi.org/10.1155/2017/8197085>
- 94 Nozumi, M., Nakatsu, F., Katoh, K. and Igarashi, M. (2017) Coordinated movement of vesicles and actin bundles during nerve growth revealed by superresolution microscopy. *Cell Rep.* **18**, 2203–2216 <https://doi.org/10.1016/j.celrep.2017.02.008>
- 95 Howard, L., Nelson, K.K., Maciewicz, R.A. and Blobel, C.P. (1999) Interaction of the metalloprotease disintegrins MDC9 and MDC15 with two SH3 domain-containing proteins, endophilin I and SH3PX1. *J. Biol. Chem.* **274**, 31693–31699 <https://doi.org/10.1074/jbc.274.44.31693>
- 96 Chen, S.L., Liu, Y.G., Zhou, Y.T., Zhao, P., Ren, H., Xiao, M. et al. (2019) Endophilin-A2-mediated endocytic pathway is critical for enterovirus 71 entry into caco-2 cells. *Emerg. Microbes Infect.* **8**, 773–786 <https://doi.org/10.1080/22221751.2019.1618686>
- 97 Lemnitzer, F., Raschbichler, V., Kolodziejczak, D., Israel, L., Imhof, A., Bailer, S.M. et al. (2013) Mouse cytomegalovirus egress protein pM50 interacts with cellular endophilin-A2. *Cell Microbiol.* **15**, 335–351 <https://doi.org/10.1111/cmi.12080>
- 98 Wang, M.Q., Kim, W., Gao, G., Torrey, T.A., Morse, III, H.C., De Camilli, P. et al. (2003) Endophilins interact with Moloney murine leukemia virus Gag and modulate virion production. *J. Biol.* **3**, 4 <https://doi.org/10.1186/1475-4924-3-4>
- 99 Wixler, V., Laplantine, E., Geerts, D., Sonnenberg, A., Petersohn, D., Eckes, B. et al. (1999) Identification of novel interaction partners for the conserved membrane proximal region of alpha-integrin cytoplasmic domains. *FEBS Lett.* **445**, 351–355 [https://doi.org/10.1016/S0014-5793\(99\)00151-9](https://doi.org/10.1016/S0014-5793(99)00151-9)
- 100 Peter, B.J., Kent, H.M., Mills, I.G., Vallis, Y., Butler, P.J., Evans, P.R. et al. (2004) BAR domains as sensors of membrane curvature: the amphiphysin BAR structure. *Science* **303**, 495–499 <https://doi.org/10.1126/science.1092586>
- 101 Itoh, T., Erdmann, K.S., Roux, A., Habermann, B., Werner, H. and De Camilli, P. (2005) Dynamin and the actin cytoskeleton cooperatively regulate plasma membrane invagination by BAR and F-BAR proteins. *Dev. Cell* **9**, 791–804 <https://doi.org/10.1016/j.devcel.2005.11.005>
- 102 Loerke, D., Mettlen, M., Yarar, D., Jaqaman, K., Jaqaman, H., Danuser, G. et al. (2009) Cargo and dynamin regulate clathrin-coated pit maturation. *PLoS Biol.* **7**, e57 <https://doi.org/10.1371/journal.pbio.1000057>
- 103 Liang, S., Wei, F.Y., Wu, Y.M., Tanabe, K., Abe, T., Oda, Y. et al. (2007) Major Cdk5-dependent phosphorylation sites of amphiphysin 1 are implicated in the regulation of the membrane binding and endocytosis. *J. Neurochem.* **102**, 1466–1476 <https://doi.org/10.1111/j.1471-4159.2007.04507.x>
- 104 Marks, B. and McMahon, H.T. (1998) Calcium triggers calcineurin-dependent synaptic vesicle recycling in mammalian nerve terminals. *Curr. Biol.* **8**, 740–749 [https://doi.org/10.1016/S0960-9822\(98\)70297-0](https://doi.org/10.1016/S0960-9822(98)70297-0)
- 105 Lai, M.M., Hong, J.J., Ruggiero, A.M., Burnett, P.E., Slepnev, V.I., De Camilli, P. et al. (1999) The calcineurin-dynamin 1 complex as a calcium sensor for synaptic vesicle endocytosis. *J. Biol. Chem.* **274**, 25963–25966 <https://doi.org/10.1074/jbc.274.37.25963>
- 106 Caldieri, G., Barbieri, E., Nappo, G., Raimondi, A., Bonora, M., Conte, A. et al. (2017) Reticulon 3-dependent ER-PM contact sites control EGFR nonclathrin endocytosis. *Science* **356**, 617–624 <https://doi.org/10.1126/science.aah6152>
- 107 Patel, P. and Woodgett, J.R. (2017) Glycogen synthase kinase 3: a kinase for all pathways? *Curr. Top. Dev. Biol.* **123**, 277–302 <https://doi.org/10.1016/bs.ctdb.2016.11.011>
- 108 Zoncu, R., Perera, R.M., Balkin, D.M., Pirruccello, M., Toomre, D. and De Camilli, P. (2009) A phosphoinositide switch controls the maturation and signaling properties of APPL endosomes. *Cell* **136**, 1110–1121 <https://doi.org/10.1016/j.cell.2009.01.032>
- 109 Cullen, P.J. and Steinberg, F. (2018) To degrade or not to degrade: mechanisms and significance of endocytic recycling. *Nat. Rev. Mol. Cell Biol.* **19**, 679–696 <https://doi.org/10.1038/s41580-018-0053-7>
- 110 Sandvig, K., Garred, O., Prydz, K., Kozlov, J.V., Hansen, S.H. and van Deurs, B. (1992) Retrograde transport of endocytosed Shiga toxin to the endoplasmic reticulum. *Nature* **358**, 510–512 <https://doi.org/10.1038/358510a0>
- 111 Gewinner, C., Wang, Z.C., Richardson, A., Teruya-Feldstein, J., Etemadmoghadam, D., Bowtell, D. et al. (2009) Evidence that inositol polyphosphate 4-phosphatase type II is a tumor suppressor that inhibits PI3K signaling. *Cancer Cell* **16**, 115–125 <https://doi.org/10.1016/j.ccr.2009.06.006>
- 112 Huotari, J. and Helenius, A. (2011) Endosome maturation. *EMBO J.* **30**, 3481–3500 <https://doi.org/10.1038/emboj.2011.286>
- 113 Scott, C.C., Vacca, F. and Gruenberg, J. (2014) Endosome maturation, transport and functions. *Semin. Cell Dev. Biol.* **31**, 2–10 <https://doi.org/10.1016/j.semcdb.2014.03.034>
- 114 Mercier, V., Laporte, M.H., Destaing, O., Blot, B., Blouin, C.M., Pernet-Gallay, K. et al. (2016) ALG-2 interacting protein-X (Alix) is essential for clathrin-independent endocytosis and signaling. *Sci. Rep.* **6**, 26986 <https://doi.org/10.1038/srep26986>
- 115 Murdoch, J.D., Rostovsky, C.M., Gowrisankaran, S., Arora, A.S., Soukup, S.F., Vidal, R. et al. (2016) Endophilin-A deficiency induces the Foxo3a-Fbxo32 network in the brain and causes dysregulation of autophagy and the ubiquitin-proteasome system. *Cell Rep.* **17**, 1071–1086 <https://doi.org/10.1016/j.celrep.2016.09.058>
- 116 Soukup, S.F. and Verstreken, P. (2017) Endoa/Endophilin-A creates docking stations for autophagic proteins at synapses. *Autophagy* **13**, 971–972 <https://doi.org/10.1080/15548627.2017.1286440>
- 117 Wang, X.Q., Xu, Z.T., Zhang, G.P., Hou, N., Mo, Q.X., Wei, J. et al. (2019) Endophilin A2 attenuates cardiac hypertrophy induced by isoproterenol through the activation of autophagy. *Am. J. Transl. Res.* **11**, 5065–5075 PMID:31497222
- 118 Ren, Y., Xu, H.W., Davey, F., Taylor, M., Aiton, J., Coote, P. et al. (2008) Endophilin I expression is increased in the brains of Alzheimer disease patients. *J. Biol. Chem.* **283**, 5685–5691 <https://doi.org/10.1074/jbc.M707932200>
- 119 Yu, Q., Wang, Y., Du, F., Yan, S., Hu, G., Origlia, N. et al. (2018) Overexpression of endophilin A1 exacerbates synaptic alterations in a mouse model of Alzheimer's disease. *Nat. Commun.* **9**, 2968 <https://doi.org/10.1038/s41467-018-04389-0>
- 120 Yin, Y., Cha, C., Wu, F., Li, J., Li, S., Zhu, X., et al. (2019) Endophilin 1 knockdown prevents synaptic dysfunction induced by oligomeric amyloid β . *Mol. Med. Rep.* **19**, 4897–4905 <https://doi.org/10.3892/mmr.2019.10158>
- 121 Gilks, W.P., Abou-Sleiman, P.M., Gandhi, S., Jain, S., Singleton, A., Lees, A.J. et al. (2005) A common LRRK2 mutation in idiopathic Parkinson's disease. *Lancet* **365**, 415–416 [https://doi.org/10.1016/S0140-6736\(05\)17830-1](https://doi.org/10.1016/S0140-6736(05)17830-1)

- 122 Seshadri, S., Fitzpatrick, A.L., Ikram, M.A., DeStefano, A.L., Gudnason, V., Boada, M. et al. (2010) Genome-wide analysis of genetic loci associated with Alzheimer disease. *JAMA* **303**, 1832–1840 <https://doi.org/10.1001/jama.2010.574>
- 123 Wang, H.F., Wan, Y., Hao, X.K., Cao, L., Zhu, X.C., Jiang, T. et al. (2016) Bridging integrator 1 (BIN1) genotypes mediate Alzheimer's disease risk by altering neuronal degeneration. *J. Alzheimers Dis.* **52**, 179–190 <https://doi.org/10.3233/JAD-150972>
- 124 Chapuis, J., Hansmannel, F., Gistelnic, M., Mounier, A., Van Cauwenbergh, C., Kolen, K.V. et al. (2013) Increased expression of BIN1 mediates Alzheimer genetic risk by modulating tau pathology. *Mol. Psychiatry* **18**, 1225–1234 <https://doi.org/10.1038/mp.2013.1>
- 125 Calafate, S., Flavin, W., Verstreken, P. and Moechars, D. (2016) Loss of Bin1 promotes the propagation of tau pathology. *Cell Rep.* **17**, 931–940 <https://doi.org/10.1016/j.celrep.2016.09.063>
- 126 Wagoner, N.E. and Bridwell, S.D. (1989) High school students' motivations for a career as a physician. *Acad. Med.* **64**, 325–327 <https://doi.org/10.1097/00001888-198906000-00009>
- 127 So, C.W., Caldas, C., Liu, M.M., Chen, S.J., Huang, Q.H., Gu, L.J. et al. (1997) EEN encodes for a member of a new family of proteins containing an Src homology 3 domain and is the third gene located on chromosome 19p13 that fuses to MLL in human leukemia. *Proc. Natl Acad. Sci. U.S.A.* **94**, 2563–2568 <https://doi.org/10.1073/pnas.94.6.2563>
- 128 Majumdar, S., Gong, E.M., Di Vizio, D., Dreyfuss, J., Degraff, D.J., Hager, M.H. et al. (2013) Loss of Sh3gl2/endophilin A1 is a common event in urothelial carcinoma that promotes malignant behavior. *Neoplasia* **15**, 749–760 <https://doi.org/10.1593/neo.121956>
- 129 Poudel, K.R., Roh-Johnson, M., Su, A., Ho, T., Mathysaraja, H., Anderson, S. et al. (2018) Competition between TIAM1 and membranes balances endophilin A3 activity in cancer metastasis. *Dev. Cell* **45**, 738–752 e736 <https://doi.org/10.1016/j.devcel.2018.05.021>
- 130 Carmona, G., Perera, U., Gillett, C., Naba, A., Law, A.L., Sharma, V.P. et al. (2016) Lamellipodin promotes invasive 3D cancer cell migration via regulated interactions with Ena/VASP and SCAR/WAVE. *Oncogene* **35**, 5155–5169 <https://doi.org/10.1038/nc.2016.47>
- 131 Cerqueira, O.L., Truesdell, P., Baldassarre, T., Vilella-Arias, S.A., Watt, K., Meens, J. et al. (2015) CIP4 promotes metastasis in triple-negative breast cancer and is associated with poor patient prognosis. *Oncotarget* **6**, 9397–9408 <https://doi.org/10.18632/oncotarget.3351>
- 132 Truesdell, P., Ahn, J., Chander, H., Meens, J., Watt, K., Yang, X. et al. (2015) CIP4 promotes lung adenocarcinoma metastasis and is associated with poor prognosis. *Oncogene* **34**, 3527–3535 <https://doi.org/10.1038/nc.2014.280>
- 133 Suman, P., Mishra, S. and Chander, H. (2018) High expression of FBP17 in invasive breast cancer cells promotes invadopodia formation. *Med. Oncol.* **35**, 71 <https://doi.org/10.1007/s12032-018-1132-5>
- 134 Ghosh, S., Scozzaro, S., Ramos, A.R., Delcambre, S., Chevalier, C., Krejci, P. et al. (2018) Inhibition of SHIP2 activity inhibits cell migration and could prevent metastasis in breast cancer cells. *J. Cell Sci.* **131**, jcs216408 <https://doi.org/10.1242/jcs.216408>
- 135 Tao, Y., Hu, K., Tan, F., Zhang, S., Zhou, M., Luo, J. et al. (2016) SH3-domain binding protein 1 in the tumor microenvironment promotes hepatocellular carcinoma metastasis through WAVE2 pathway. *Oncotarget* **7**, 18356–18370 <https://doi.org/10.18632/oncotarget.7786>
- 136 Baldassarre, T., Truesdell, P. and Craig, A.W. (2017) Endophilin A2 promotes HER2 internalization and sensitivity to trastuzumab-based therapy in HER2-positive breast cancers. *Breast Cancer Res.* **19**, 110 <https://doi.org/10.1186/s13058-017-0900-z>
- 137 Chew, H.Y., De Lima, P.O., Gonzalez Cruz, J.L., Banushi, B., Echejoh, G., Hu, L. et al. (2020) Endocytosis inhibition in humans to improve responses to ADCC-mediating antibodies. *Cell* **180**, 895–914 e827 <https://doi.org/10.1016/j.cell.2020.02.019>
- 138 Wang, W., Erbe, A.K., Hank, J.A., Morris, Z.S. and Sondel, P.M. (2015) NK cell-mediated antibody-dependent cellular cytotoxicity in cancer immunotherapy. *Front. Immunol.* **6**, 368 <https://doi.org/10.3389/fimmu.2015.00368>
- 139 Scott, A.M., Wolchok, J.D. and Old, L.J. (2012) Antibody therapy of cancer. *Nat. Rev. Cancer* **12**, 278–287 <https://doi.org/10.1038/nrc3236>
- 140 Li, Y.Y., Chen, X.N., Fan, X.X., Zhang, Y.J., Gu, J., Fu, X.W. et al. (2015) Upregulated dynamin 1 in an acute seizure model and in epileptic patients. *Synapse* **69**, 67–77 <https://doi.org/10.1002/syn.21788>
- 141 Schiffer, M., Teng, B., Gu, C., Shchedrina, V.A., Kasaikina, M., Pham, V.A. et al. (2015) Pharmacological targeting of actin-dependent dynamin oligomerization ameliorates chronic kidney disease in diverse animal models. *Nat. Med.* **21**, 601–609 <https://doi.org/10.1038/nm.3843>
- 142 Bar-Yosef, H., Vivanco Gonzalez, N., Ben-Aroya, S., Kron, S.J. and Kornitzer, D. (2017) Chemical inhibitors of *candida albicans* hyphal morphogenesis target endocytosis. *Sci. Rep.* **7**, 5692 <https://doi.org/10.1038/s41598-017-05741-y>
- 143 Hernaez, B. and Alonso, C. (2010) Dynamin- and clathrin-dependent endocytosis in African swine fever virus entry. *J. Virol.* **84**, 2100–2109 <https://doi.org/10.1128/JVI.01557-09>
- 144 Simanjuntak, Y., Liang, J.J., Lee, Y.L. and Lin, Y.L. (2015) Repurposing of prochlorperazine for use against dengue virus infection. *J. Infect. Dis.* **211**, 394–404 <https://doi.org/10.1093/infdis/jiu377>
- 145 Renard, H.F., Tyckaert, F., Lo Giudice, C., Hirsch, T., Valades-Cruz, C.A., Lemaigre, C. et al. (2020) Endophilin-A3 and galectin-8 control the clathrin-independent endocytosis of CD166. *Nat. Commun.* **11**, 1457 <https://doi.org/10.1038/s41467-020-15303-y>
- 146 Gowrisankaran, S., Houy, S., Del Castillo, J.G.P., Steubler, V., Gelker, M., Kroll, J. et al. (2020) Endophilin-A coordinates priming and fusion of neurosecretory vesicles via intersectin. *Nat. Commun.* **11**, 1266 <https://doi.org/10.1038/s41467-020-14993-8>
- 147 Pechstein, A., Gerth, F., Milosevic, I., Japel, M., Eichhorn-Grunig, M., Vorontsova, O. et al. (2015) Vesicle uncoating regulated by SH3-SH3 domain-mediated complex formation between endophilin and intersectin at synapses. *EMBO Rep.* **16**, 232–239 <https://doi.org/10.15252/embr.201439260>
- 148 Bai, J., Hu, Z., Dittman, J.S., Pym, E.C. and Kaplan, J.M. (2010) Endophilin functions as a membrane-bending molecule and is delivered to endocytic zones by exocytosis. *Cell* **143**, 430–441 <https://doi.org/10.1016/j.cell.2010.09.024>