Research Article



Significant association between the endothelial lipase gene 584C/T polymorphism and coronary artery disease risk

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Several studies have investigated a potential association between the endothelial lipase gene (LIPG) 584C/T polymorphism and susceptibility to coronary artery disease (CAD), but a uniform conclusion is yet to be reached. To better evaluate the true relationship between the LIPG 584C/T polymorphism and the risk of CAD, a meta-analysis of 14 case-control studies with 9731 subjects was performed. Relevant articles published through August 2020 were searched in the CNKI, PubMed, Embase and Web of Science databases. Thirteen articles, including 14 eligible case-control studies with 4025 cases and 5706 controls, were enrolled in the present meta-analysis. The Newcastle-Ottawa Scale (NOS) scores of the case-control studies ranged from 6 to 8. The pooled results indicated that there is a significant association between the LIPG 584C/T polymorphism and CAD in the homozygote comparison model and the allelic comparison model. Subgroup analyses revealed that the LIPG 584C/T mutation significantly decreased the risk of CAD in the subgroups of African, CAD, hospital-based (HB), and polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) populations in some genetic models. No publication bias was found in our meta-analysis, which certifies the robustness of the current meta-analysis. Trial sequential analysis (TSA) also confirmed the stability of our results. The results of our meta-analysis indicate that the LIPG 584C/T polymorphism plays a protective role in the incidence of CAD. More high-quality case-control studies on various ethnicities are needed to confirm our results.

Introduction

Coronary artery disease (CAD), also called coronary heart disease (CHD), characterized by myocardial hypoxia and ischemia is triggered by coronary atherosclerosis [1]. It is a medical problem of human society and one of the leading causes of disability and deaths in every country [2,3]. The precise mechanism of the onset and development of CAD is still obscure. The pathogenesis of CAD involves many risk factors, including hereditary and environmental factors. Recently, increasing evidence has suggested that genetic factors may play an important role in its pathogenesis. Many genetic variants have been identified to be related to the risk of CAD by previous genetic association studies [4]. The single-nucleotide polymorphism (SNP) is one of the most common genetic variants. Some SNPs may be related to the occurrence and development of CAD, while others are not [5,6].

Endothelial lipase (EL), a new member of the triglyceride (TG) lipase family, was first discovered in 1999. EL consists of 483 amino acids, with a molecular weight of approximately 55 kDa and is mainly secreted from endothelial cells and other cells such as hepatocytes and macrophages [7]. Compared with lipoprotein lipase (LPL) and hepatic lipase (HL), which are also members of the TG lipase family, EL has

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higher phospholipase activity and lower TG lipase. A large number of studies have demonstrated that EL plays a key role in high-density lipoprotein (HDL) metabolism. As a regulator of plasma HDL-C levels, EL can modulate the levels of plasma HDL-C inversely [8]. Elevated plasma HDL levels are associated with a protective effect on CAD and low plasma HDL levels are associated with an elevated risk of CAD [9]. Recently, increasing evidence has indicated that EL plays a pivotal role in the pathogenesis of CAD in part by reducing plasma HDL levels.

The EL gene (*LIPG*), which encodes the EL protein and is located on chromosome 18q21.1, spans approximately 30 kb with ten exons and nine introns [10]. Several genetic polymorphisms have been confirmed in *LIPG*. Among these, the 584C/T gene variant (rs2000813) has been the most frequently investigated variant that can lead the amino acid threonine substitution for isoleucine at codons [11]. Several published studies have assessed the association between *LIPG* 584C/T polymorphism and the risk of CAD; however, the results are still inconclusive and contradictory [12–14]. To testify these results, Cai et al. [15] carried out a meta-analysis to explore the association between the *LIPG* 584C/T variation and CAD risk in 2014. Unfortunately, the result of their meta-analysis was still inconclusive. Thereafter, several new studies have investigated the correlation between the *LIPG* 584C/T polymorphism and CAD risk, but the results of these studies remain inconsistent. Therefore, to elucidate the precise correlation of the *LIPG* 584C/T polymorphism with CAD susceptibility, we performed this updated meta-analysis.

Materials and methods Search strategy

A systematic literature network search without language limitations was performed in the CNKI, PubMed, Embase and Web of Science databases to acquire all eligible case–control studies published before 15 August 2020. The relevant search terms included the following: (endothelial lipase gene OR EL OR *LIPG*) AND (polymorphism OR mutation OR variation OR genotype) AND (cardiovascular disease OR coronary heart disease OR coronary artery disease OR angina pectoris OR acute coronary syndrome OR myocardial infarction). Furthermore, to acquire other relevant published articles, the reference lists of the studies and reviews, which were included from the search parameters illuminated above, were searched.

Inclusion and exclusion criteria

CAD was defined as acute myocardial infarction (AMI), CHD, CAD, myocardial infarction (MI), acute coronary syndrome (ACS), cardiovascular disease. The included articles fulfilled the following criteria: (1) studies assessed the correlation between the *LIPG* 584C/T polymorphism and CAD risk; (2) studies were case–control or cohort; (3) articles reported in Chinese or English; and (4) studies provided precise data for genotype distribution estimation in both groups. Studies were excluded according to the following criteria: (1) not case–control and cohort studies, letters, case reports, reviews or meta-analyses; (2) deficient data; (3) deviating from Hardy–Weinberg equilibrium (HWE) in the control group; and (4) overlapping data or duplicated publications.

Data extraction

Two researchers (Yue-e Wu and Lan Ma) independently extracted data from the qualified articles based on the inclusion criteria. The following information was extracted from the eligible studies: first author, year of publication, ethnicity and country of participants, type of diseases, genotyping method, sources of controls, number of cases and controls for the 584C/T genotypes of *LIPG* and the *P*-value of the HWE test of controls. When we encountered inconsistent evaluations, all researchers were consulted to acquire an agreement regarding the exclusion or inclusion of the study in the present article.

Methodological quality assessment

The Newcastle–Ottawa Scale (NOS) was used to evaluate the quality of each included study [16]. For the selection and exposure categories, each study can be awarded a maximum of one star for each numbered item. A maximum of two stars can be given for the comparability categories. The NOS has a score range of 0–9, and a study with a score higher than 5 could be included in the present meta-analysis.

Statistical analysis

The present meta-analysis was conducted based on the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement [17]. Chi-square test was applied to assess the HWE in the control groups with significance set at *P* less than 0.05. Odds ratios (ORs) with 95% confidence intervals (CIs) were used to evaluate

Table 1 Characteristics of	f eligible case-control s	studies included in this	meta-analysis
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First author	Year	Country	Ethnicity	Type of disease	Source of controls	Genotyping method	Number of cases	Number of controls	HWE (Control)	NOS score
Zhu et al. [12]	2007	China	Asian	CHD	HB	PCR-RFLP	242	196	0.063	7
Ji et al. [23]	2015	China	Asian	ACS	HB	PCR-RFLP	195	159	0.077	6
Jensen et al. [24]	2009	Denmark	Caucasian	ACS	PB	TaqMan	998	1643	0.888	8
Rimm et al. [25]	1992	America	Caucasian	CAD	PB	PCR	262	519	0.063	8
Colditz et al. [26]	1997	America	Caucasian	CAD	PB	PCR	241	477	0.220	8
Tjonneland et al.(1) [27]	2007	Denmark	Caucasian	CHD	PB	TaqMan	235	763	0.838	8
Tjonneland et al.(2) [27]	2007	Denmark	Caucasian	CHD	PB	TaqMan	763	880	0.997	8
Tang et al. [9]	2008	China	Asian	CAD	HB	PCR-RFLP	265	265	0.103	7
Xie et al. [13]	2015	China	Asian	CAD	HB	PCR	287	367	0.065	7
Solim et al. [28]	2018	Turkey	Caucasian	CAD	HB	TaqMan	74	73	0.545	7
Elnaggar et al. [29]	2019	Egypt	African	CAD	HB	PCR-RFLP	84	42	0.492	8
Dalan et al. [30]	2013	Turkey	Caucasian	CAD	HB	PCR-RFLP	104	76	0.286	7
Cai et al. [14]	2014	China	Asian	CAD	HB	PCR-RFLP	135	166	0.146	7
Toosi et al. [31]	2015	Iran	Asian	CAD	HB	PCR-RFLP	140	80	0.092	8

the strength of the relationship between the *LIPG* 584C/T polymorphism and the risk of CAD in five genetic models: heterozygote comparisons (*CT vs. CC*), homozygote comparisons (*TT vs. CC*), recessive model (*TT vs. CT* + *CC*), dominant model (*TT* + *CT vs. CC*), and allelic comparisons (*T vs. C*). Stratified analyses were also performed according to ethnicity (Asian, Caucasian, and African populations), types of diseases (CHD, ACS, and CAD), genotyping method (polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP), Taqman, and PCR), and sources of controls (hospital-based (HB) and population-based (PB)). Differences in terms of a Z-test were considered statistically significant if the *P*-value was less than 0.05. The Cochran's Q statistical test and the I^2 test were used to measure the heterogeneity within the studies, and the significance was set at *P* less than 0.05 [18]. A fixed-effects model or random-effects model was used to calculate the pooled OR according to the heterogeneity [19]. Influence analysis was carried out to examine the effect of an individual study on the pooled OR by omitting a single study each time to assess the stability of the pooled results. Publication bias was examined by Begg's funnel plot test and Egger's test [20,21]. Trial sequential analysis (TSA) was carried out as described in our previous study [22]. STATA 12.0 software (version 12.0; STATA Corp. College Station, TX, U.S.A.) was used to conduct all statistical analyses.

Results Characteristics of eligit

Characteristics of eligible studies

The process of screening eligible articles is shown in Figure 1. One hundred and thirty-seven potential articles were acquired after a systematic literature search. One hundred and twenty-four articles were removed after the comprehensive examining procedures according to the article titles, abstracts, and full-texts. Finally, the remaining 13 articles, including 14 eligible independent case–control studies with 4025 patients and 5706 controls, were identified and included in the meta-analysis based on the inclusion and exclusion criteria [9,12–14,23–31].

Table 1 displays the characteristics of all case–control studies included in our meta-analysis. All studies were reported in English except for two studies that were published in Chinese. Among the 14 studies, 7 studies were conducted in Caucasian populations, 6 in Asian populations, and only 1 in African populations. There were nine HB studies and five PB studies. Seven studies used PCR-RFLP as the genotyping method, four studies used TaqMan, and three studies used PCR. The genotype distributions of controls in all included studies were accordance with HWE. Table 2 shows the distribution of genotypes and allele frequencies of the *LIPG* 584C/T polymorphism in the cases



PRISMA 2009 Flow Diagram Identification Records identified through Additional records identified database searching through other sources (n = 135) (n = 2)Records after duplicates removed (n = 128)Records excluded (n = 88) Screening 61 not relevant studies 24 not relevant to CAD 3 review and meta-analysis Full-text articles assessed for eligibility (n = 40) Records excluded (n = 27)3 not sufficient data 5 not case-control 5 data covered by other Eligibility 10 not relevant to LIPG 584C/T polymorphism Studies included in 4 deviated from HWE qualitative synthesis (n = 13) Included Studies included in quantitative synthesis (meta-analysis) (n = 13)

From: Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group (2009). Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. PLoS Med 6(7): e1000097. doi:10.1371/journal.pmed1000097

For more information, visit www.prisma-statement.org.

Figure 1. The flow diagram of the included and excluded studies



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First author	Year				Geno	type (<i>n</i>)				А	llele fre	quency (/	7)	HWE (Con- trol)
			Ca	ise			Control			Ca	se	Con	trol	
		Total	CC	СТ	TT	Total	CC	СТ	TT	С	Т	С	т	
Zhu et al. [12]	2007	242	186	56	0	196	150	46	0	428	56	346	46	0.063
Ji et al. [23]	2015	195	112	76	7	159	81	71	7	300	90	233	85	0.077
Jensen et al. [24]	2009	998	509	406	83	1643	837	673	133	1424	572	2347	939	0.888
Rimm et al. [25]	1992	262	129	117	16	519	240	239	40	375	149	719	319	0.063
Colditz et al. [26]	1997	241	115	110	16	477	224	214	39	340	142	662	292	0.220
Tjonneland et al.(1) [27]	2007	235	116	102	17	763	391	312	60	334	136	1094	432	0.838
Tjonneland et al.(2) [27]	2007	763	393	304	66	880	446	361	73	1090	436	1253	507	0.997
Tang et al. [9]	2008	265	174	85	6	265	125	122	18	433	97	372	158	0.103
Xie et al. [13]	2015	287	160	116	11	367	216	139	12	436	138	571	163	0.065
Solim et al. [28]	2018	74	40	27	6	73	26	33	14	107	39	85	61	0.545
Elnaggar et al. [29]	2019	84	58	23	2	42	17	21	4	139	27	55	29	0.492
Dalan et al. [30]	2013	104	44	59	1	76	45	29	2	147	61	119	33	0.286
Cai et al. [14]	2014	135	84	45	6	166	97	64	5	213	57	258	74	0.146
Toosi et al. [31]	2015	140	67	70	3	80	28	46	6	204	76	102	58	0.092

Table 2 LIPG 584C/T polymorphism genotype distribution and allele frequency in cases and controls

and controls. Supplementary Table S1 indicated that the studies included in our study were reliable according to the NOS.

Test of heterogeneity

To assess the heterogeneity among the included studies, Q-test and I^2 statistics were applied. High heterogeneity was found across studies in all genetic models except for the recessive comparisons. Thus, random-effects analysis was applied to synthesize the data. Moreover, we explored the heterogeneity of all genetic models based on different ethnicities, type of diseases, genotyping method, and sources of controls. However, the significant heterogeneity could not be entirely explained by diverse ethnicities (Table 3).

Meta-analysis results

The association between the *LIPG* 584C/T polymorphism and the risk of CAD was assessed. The integrated results indicated that the *LIPG* 584C/T variation was significantly associated with CAD risk in the homozygote comparison model (*TT vs. CC:* OR = 0.74, 95% CI = 0.56–0.99, *P*=0.041, *P*_{heterogeneity}=0.029, Table 3 and Figure 2) and allelic comparison model (*T vs. C:* OR = 0.87, 95% CI = 0.76–0.99, *P*=0.037, *P*_{heterogeneity}=0.000, Table 3 and Figure 3). There was no significant association with CAD risk in the other three models, including *CT vs. CC:* OR = 0.89, 95% CI = 0.77–1.03, *P*=0.115, *P*_{heterogeneity}=0.001; *TT + CT vs. CC:* OR = 0.86, 95% CI = 0.73–1.01, *P*=0.066, *P*_{heterogeneity}=0.000; and *TT vs. CT + CC:* OR = 0.88, 95% CI = 0.75–1.04, *P*=0.136, *P*_{heterogeneity}=0.182 (Table 3).

In the subgroup analysis according to ethnicity, the results showed a decreased CAD risk in African populations under four genetic models (*TT vs. CC*: OR = 0.15, 95% CI = 0.03–0.87, *P*=0.035; *CT vs. CC*: OR = 0.32, 95% CI = 0.14–0.72, *P*=0.005; *TT* + *CT vs. CC*: OR = 0.29, 95% CI = 0.14–0.64, *P*=0.002; and *T vs. C*: OR = 0.37, 95% CI = 0.20–0.68, *P*=0.001). However, there was no significant association between the *LIPG* 584C/T polymorphism and the risk of CAD that was identified in Asian and Caucasian populations in all genetic models. In the subgroup analysis stratified by type of diseases, the results indicated that the *LIPG* 584C/T polymorphism was significantly associated with decreased CAD risk in the subgroup of CAD in three genetic models (*TT vs. CC*: OR = 0.55, 95% CI = 0.34–0.88, *P*=0.014; *TT vs. CT* + *CC*: OR = 0.66, 95% CI = 0.49–0.89, *P*=0.006; and *T vs. C*: OR = 0.79, 95% CI = 0.62–0.99, *P*=0.043), and the same results were shown in the HB group (*TT vs. CC*: OR = 0.48, 95% CI = 0.26–0.88, *P*=0.018) and PCR-RFLP (*TT vs. CC*: OR = 0.42, 95% CI = 0.25–0.70, *P*=0.001). Detailed results are shown in Table 3.

Sensitivity analysis

Sensitivity analysis was carried out to examine the influence of each eligible study on the pooled ORs by the sequential removal of each individual study form the analysis. The individual removal procedure affected the pooled ORs in the



Table 3 Meta-analysis results

Genetic model	Category	n	Model	OR (95% CI)	Р	Hetero	geneity	Begg's test	Egger' test
	• •					l ²	P	Р	Р
Homozyaote $(TT vs, CC)$	Overall	14	R	0.740 [0.555; 0.987]	0.041	47.5%	0.020	0.059	0.121
1011102)goto (11 v3. 00)	Asian	6	R	0.599 [0.275; 1.303]					0.727
	Caucasian	7	F	0.920 [0.764; 1.110]					0.115
	African	1	F	0.147 [0.025; 0.870]					-
	CHD	3	F	1.006 [0.741; 1.364]					-
	ACS	2	F	1.002 [0.753; 1.333]					-
	CAD	9	R	0.545 [0.336; 0.884]					0.651
	HB	9	R	0.474 [0.255; 0.880]					0.814
	PB	9 5	F						0.014
	PCR-RFLP	7	F	0.963 [0.796; 1.165]					0.080
				0.420 [0.253; 0.698]					
	Taqman	4	F	0.963 [0.783; 1.186]					0.151
	PCR	3	F	0.850 [0.577; 1.252]					0.077
eterozygote (C1 vs. CC)	Overall	14	R	0.883 [0.765; 1.031]					0.441
	Asian	6	R	0.786 [0.599; 1.031]		Heterogeneity test p l^2 p p 41 47.5% 0.029 0.059 66 61.2% 0.035 0.806 64 7.7% 0.370 0.072 55 - - - 60 0.0% 0.836 1.000 61 49.6% 0.044 0.602 68 50.6% 0.048 0.711 66 0.0% 0.860 0.086 61 39.1% 0.145 1.000 55 61.1% 0.001 0.663 52 59.5% 0.030 0.707 0 37.2% 0.145 1.000 56 - - - 66 0.0% 0.747 1.000 57 - - - 56 - - - 61.1% 0.000 0.251 88 71.5% 0.000 72.6% <	0.856		
	Caucasian	7	F	0.994 [0.898; 1.101]				test P 229 0.059 035 0.806 370 0.072 - - 336 1.000 542 1.000 542 1.000 543 0.086 336 1.000 542 1.000 543 0.086 145 0.089 513 0.296 001 0.663 030 0.707 145 1.000 293 1.000 293 1.000 000 0.251 000 0.251 000 0.764 320 0.707 021 0.700 0201 0.764 320 0.707 031 0.000 0201 0.764 320 0.707 033 1.000 043 1.000 0258 1.000 026	0.790
	African	1	F	0.321 [0.144; 0.715]					-
	CHD	3	F	0.997 [0.850; 1.168]					0.707
	ACS	2	F	0.961 [0.824; 0.715]	0.614			1.000	-
	CAD	9	R	0.804 [0.605; 1.121]	0.131			0.251	0.675
	HB	9	R	0.779 [0.574; 1.056]	0.108	71.5%	0.000	0.466	0.848
	PB	5	F	0.986 [0.888; 1.094]	0.788	0.0%	0.929	1.000 0.74 1.000 0.74 1.000 - 0.251 0.6 0.466 0.84 1.000 0.75 0.764 0.55 0.734 0.44 1.000 0.43 0.051 0.34 0.764 0.59 0.777 0.88 0.764 0.73 1.000 0.73 1.000 0.73 1.000 - 0.348 0.6	0.725
	PCR-RFLP	7	R	0.760 [0.529; 1.092]	0.138	72.5%	0.001	0.764	0.541
		0.320	0.734	0.444					
	PCR	3	F	1.006 [0.838; 1.208]	0.947	0.0%	0.643	1.000	0.421
Dominant (<i>TT</i> + <i>CT vs.</i> <i>CC</i>)	Overall	14	R	0.856 [0.725; 1.010]	0.066	68.3%	0.000	0.051	00 0.42 51 0.36 07 0.89
	Asian	6	R	0.772 [0.570; 1.044]	0.093	68.6%	0.007	0.707	0.898
	Caucasian	7	F	0.984 [0.893;1.085]	0.752	46.7%	0.081	0.764	0.996
	African	1	F	0.293 [0.135; 0.636]	0.002	-	-		
	CHD	3	F	0.998 [0.857; 1.162]	0.979	0.0%	0.830	1.000	0.739
	ACS	2	F	0.966 [0.834; 1.120]	0.649	22.0%	0.258	1.000	-
Homozygote (TT vs. CC) Heterozygote (CT vs. CC) Dominant (TT + CT vs. CC)	CAD	9	R	0.764 [0.566; 1.032]	0.079	77.1%	0.000	0.348	0.649
	HB	9	R	0.745 [0.538; 1.031]	0.076	76.4%	0.000	0.466	0.802
	PB	5	F	0.983 [0.889; 1.086]	0.730	0.0%	0.923	0.806	0.922
	PCR-RFLP	7	R	0.734 [0.503; 1.072]	0.109	75.9%	0.000	0.764	0.529
	Taqman	4	F	0.977 [0.874; 1.091]	0.677				0.326
	PCR	3	F	0.988 [0.828; 1.180]	0.898				0.346
	Overall	14	F	0.881 [0.745; 1.041]	0.136				0.103
	Asian	6	F	0.682 [0.435; 1.068]	0.094	47.1%	0.109	0.806	0.781
	Caucasian	7	F	0.932 [0.778; 1.117]	0.448				0.031
	African	1	F	0.235 [0.041; 1.337]	0.103				-
	CHD	3	F	1.007 [0.750; 1.352]		0.0%	0.686	1.000	-
	ACS	2	F	1.013 [0.768; 1.336]					-
	CAD	9	F	0.658 [0.487; 0.889]					0.659
	HB	9	F	0.578 [0.390; 0.856]					0.726
	PB	5	F	0.969 [0.806; 1.165]					0.035
	PCR-RFLP	7	F	0.499 [0.302; 0.825]					0.881
	Taqman	4	F	0.976 [0.798; 1.193]					0.001
	PCR	4	F	0.855 [0.586; 1.246]					
Allolio $(T_{1/2}, C)$									0.028
Allelic (7 VS. C)	Overall	14	R	0.869 [0.761; 0.992]					0.288
	Asian	6	R	0.813 [0.630; 1.048]	l^2 l^2 0.041 47.5% 0.1 0.196 61.2% 0.1 0.384 7.7% 0.2 0.972 0.0% 0.2 0.990 0.0% 0.2 0.990 0.0% 0.2 0.014 49.6% 0.2 0.018 50.6% 0.2 0.696 0.0% 0.2 0.011 39.1% 0. 0.725 44.4% 0. 0.411 0.0% 0.2 0.082 59.5% 0.2 0.910 37.2% 0. 0.005 - 0. 0.966 0.0% 0.2 0.131 72.6% 0.2 0.132 72.5% 0.2 0.133 72.5% 0.2 0.712 14.4% 0.2 0.752 46.7% 0. 0.076 76.4% 0. 0.752 46.7% 0.2			0.912	
Homozygote (TT vs. CC) Heterozygote (CT vs. CC) Dominant (TT + CT vs. CC) Recessive (TT vs. CT + CC)	Caucasian	7	F	0.978 [0.907; 1.055]			0.096	0.174	0.484
	African	1	F	0.368 [0.200; 0.678]	0.001	-	-	-	-

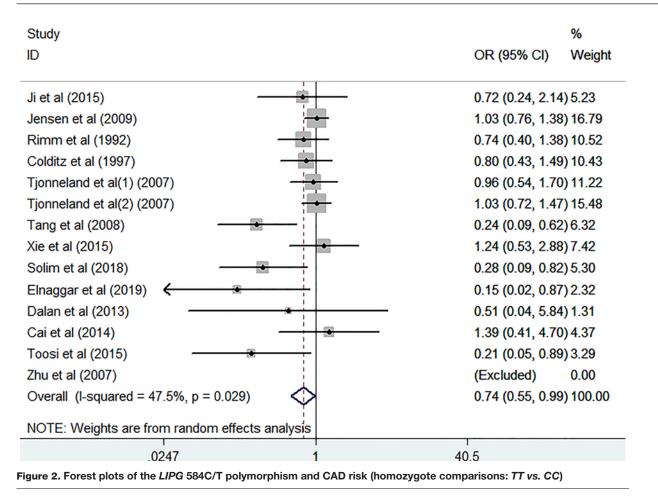
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Table 3 Meta-analysis results (Continued)

Genetic model	Category	ory <i>n</i> Mode		del OR (95% CI)		Heterogeneity		Begg's test	Egger's test	
						l ²	Р	Р	P	
	CHD	3	F	1.000 [0.886; 1.128]	0.999	0.0%	0.953	1.000	0.871	
	ACS	2	F	0.981 [0.874; 1.102]	0.750	13.5%	0.282	1.000	-	
	CAD	9	R	0.785 [0.622; 0.992]	0.043	76.3%	0.000	0.348	0.620	
	HB	9	R	0.772 [0.595; 1.001]	0.051	75.3%	0.000	0.466	0.830	
	PB	5	F	0.984 [0.910; 1.064]	0.682	0.0%	0.909	0.462	0.471	
	PCR-RFLP	7	R	0.765 [0.571; 1.026]	0.074	73.0%	0.001	0.548	0.434	
	Taqman	4	F	0.981 [0.900; 1.070]	0.669	58.0%	0.066	0.308	0.231	
	PCR	3	F	0.971 [0.845; 1.117]	0.682	0.0%	0.467	0.296	0.028	

Abbreviations: F, fixed-effects model; n, number of studies; R, random-effects model.

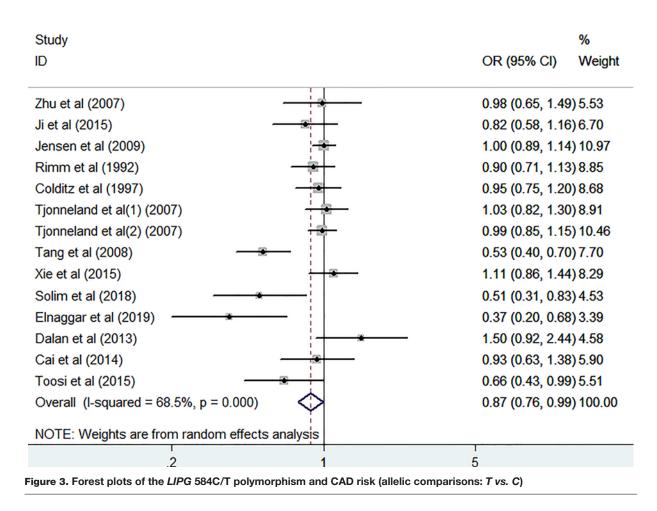


homozygote and allelic comparisons model, indicating the instability and unreliability of our findings. Sensitivity analyses of the other three genetic models showed similar results (Figure 4).

Publication bias

To assess the potential for publication bias among the studies on the the *LIPG* 584C/T polymorphism included in the present meta-analysis, Begg's and Egger's tests were performed for the five genetic models. No evidence of asymmetry was observed by the appearance of the shape in the Begg's funnel plots, and neither Egger's regression nor Begg's rank correlation indicated publication bias among the studies in the five genetic models. The publication bias tests of





subgroups were also performed, and no significant publication bias were found, except in few subgroups (Table 3 and Figure 5).

TSA

The results of the TSA using the homozygote and allelic comparisons model are shown in Figures 6 and 7. The cumulative z-curve obviously crossed the traditional boundary, and it further confirmed the results of our meta-analysis that the *LIPG* 584C/T polymorphism was significantly associated with decreased CAD risk. Nevertheless, the cumulative z-curve had crossed the TSA monitoring boundary without success before reaching the required information size and demonstrated that the cumulative sample size is insufficient and the more eligible case–control studies are necessary.

Discussion

The present meta-analysis indicated that the *LIPG* 584C/T variation was significantly associated with CAD risk in the homozygote comparison model (*TT vs. CC*: OR = 0.74, 95% CI = 0.56-0.99, *P*=0.041) and allelic comparison model (*T vs. C*: OR = 0.87, 95% CI = 0.76-0.99, *P*=0.037). According to the integrated analyses of 14 eligible case-control studies, our study revealed that the *LIPG* 584C/T polymorphism was significantly associated with a decreased risk of CAD, which suggested that the *LIPG* 584C/T polymorphism plays a protective role in the incidence of CAD in individuals and that the T allele may be a protective factor against CAD.

It has been proven that elevated plasma HDL levels are associated with a protective effect on CAD, and low plasma HDL levels are associated with an elevated risk of CAD [9]. The role of EL in the pathogenesis of CAD is regulated by its effect on the metabolism of HDL by its increasing clearance. Therefore, it is speculated that *LIPG* is closely related to the incidence of CAD by affecting the expression of EL. In recent years, more and more attention has been paid to genetic research on CAD, which is not surprising considering the impact of CAD on the health of people around the world [13]. To date, many molecular epidemiological studies have been performed to investigate the



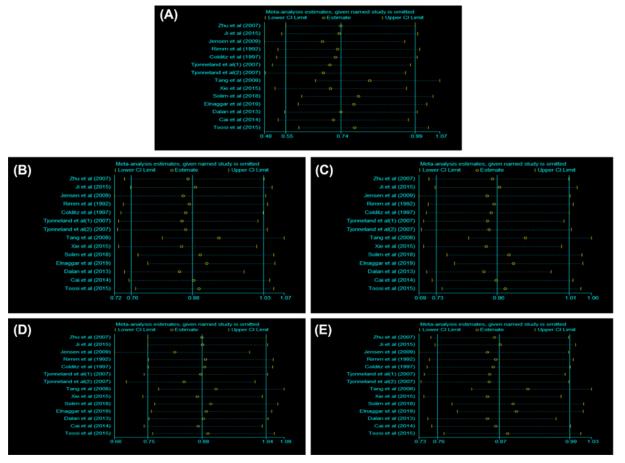


Figure 4. The results of sensitivity analysis between the *LIPG* 584C/T polymorphism and susceptibility to CAD (A) homozygous model; (B) heterozygous model; (C) dominant model; (D) recessive model; (E) allele model.

association between the *LIPG* 584C/T variant and CAD risk. However, the results of these studies are inconsistent or even contradictory. The results found by Ji et al. [23], Zhu et al. [12], and Jensen et al. [24] revealed that there was no significant association between the *LIPG* 584C/T polymorphism and risk of CAD; in contrast, Tang et al. [9], Solim et al. [28], and Elnaggar et al. [29] found similar results as ours that the *LIPG* 584C/T polymorphism was significantly associated with a decreased risk of CAD. Cai et al. conducted a meta-analysis to investigate the true association between the *LIPG* 584C/T variant and CAD risk in 2014. However, their analysis included only 9 case–control studies, including 3036 cases and 4777 controls, and concluded that there was no significant association between the *LIPG* 584C/T polymorphism and the susceptibility to CAD. Their results were inconsistent with our study, but three case–control studies of their meta-analysis did not conform to HWE [32–34]. A latest meta-analysis of 13 published case–control studies was performed by Zhao et al. (2020) [35], and also significant results were found. However, in addition to also including three case–control studies that did not conform to HWE, some qualified studies were not included in their meta-analysis. Compared with their meta-analyses, the results of our meta-analysis, which included 14 case–control studies that all were in line with HWE, had more statistical power and reliability.

We noticed significant heterogeneities among the studies and carried out subgroup analyses according to ethnicity. According to previous studies, the *LIPG* 584C/T variant had different effects on CAD in different ethnicities. Even results from studies on the same ethnicity were inconsistent. For example, Tang et al. conducted a study including 530 Chinese subjects to assess the relationship between the common variant and CAD risk [9]. They demonstrated that the T allele could significantly reduce the CAD risk. The same results were found in studies conducted by Solim et al. [28] on the Turkish population, by Shimizu et al. [32] on the Japanese population, and by Toosi et al. [31] on the Iranian population. However, Jensen et al. [24] and Zhu et al. [12] showed no relationship between the *LIPG* 584C/T variant and CAD susceptibility in Caucasian and Asian populations. Our present study showed that the *LIPG* 584C/T



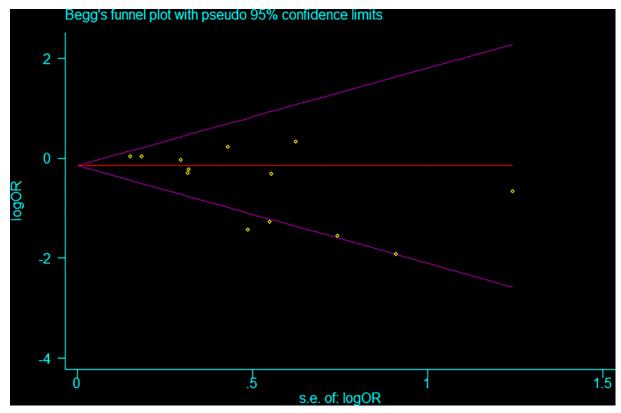
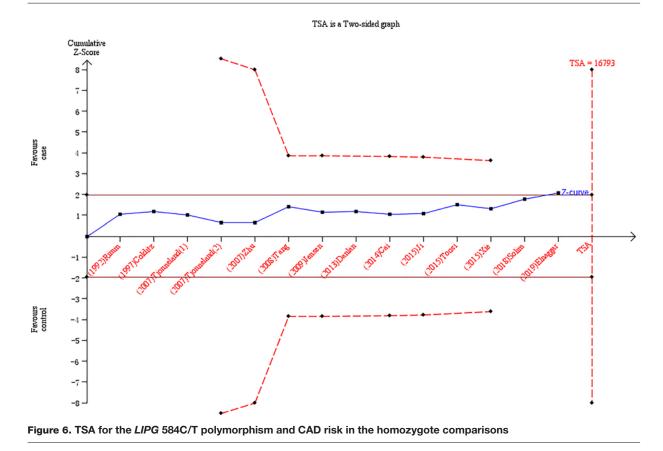


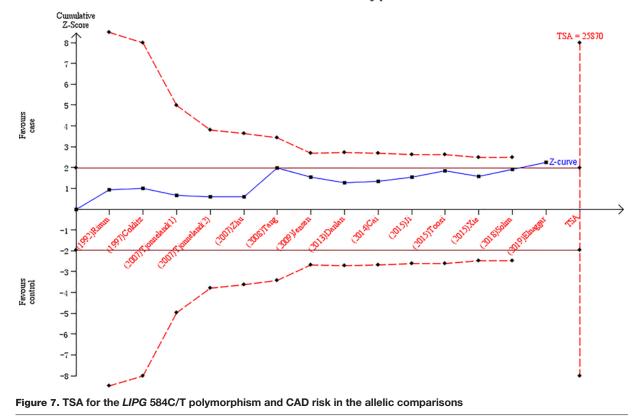
Figure 5. Funnel plot assessing evidence of publication bias (homozygote comparisons: TT vs. CC)



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TSA is a Two-sided graph



variant be connected with a decreased CAD risk only in African populations. However, there was no significant association between the *LIPG* 584C/T polymorphism and the risk of CAD that was identified in Asian and Caucasian populations. This result was not convincing enough, because only one case–control study was included in the African subgroup; therefore, this result needs to be confirmed by more high-quality case–control studies in various ethnicities. When carrying out subgroup analysis by type of diseases, we found that the *LIPG* 584C/T polymorphism was significantly associated with a reduced risk in the CAD group, but we failed to find a significant risk association in other types. This result may be explained by the inherent heterogeneity of development in diverse diseases types.

Some inherent drawbacks of our meta-analysis should be illustrated when interpreting our results. First, only unadjusted estimates were applied to evaluate the strength of the relationship between the LIPG 584C/T variant and CAD risk. Due to a lack of raw data, such as exposing factors, life habits, gene-environment interactions, interactions between gene-gene and even diverse mutation loci in the same gene factors, a further exact adjustment analysis adjusting for confounding factors could not be carried out. Second, there is still high heterogeneity in our meta-analysis, although we used strict inclusion criteria, accurate data extraction, and rigorous data analysis to perform the present meta-analysis. The persuasiveness of the significant association between the LIPG 584C/T polymorphism and CAD susceptibility was affected. Third, the sample sizes of our study were still small, especially in subgroup analysis. Only one eligible study was included in the analysis of African populations. TSA identified that the cumulative sample size was insufficient. There were not enough eligible case-control studies, weakening the statistical power to detect the real correlation between the LIPG 584C/T polymorphism and susceptibility to CAD. Fourth, the result of our meta-analysis should be interpreted with caution and needs to be confirmed by more case-control studies, because the sensitivity analyses indicated that deletion of certain individual study had an impact on the reliability of our results. Fifth, although Begg's and Egger's tests proved that there was no publication bias, it may inevitably exist in our meta-analysis. Only published articles reported in English or Chinese were included in the present study. Some candidate studies may not be included because they were not published, discovered, or because they were not published in English or Chinese.



In conclusion, our meta-analysis suggested that the *LIPG* 584C/T polymorphism plays a protective role in the incidence of CAD in individuals. The T allele may be a protective factor against CAD. Because of the limitations mention above, more high-quality case–control studies in various ethnicities are needed to confirm our results.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

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Author Contribution

Y.W. and Z.H. conceived the study. Y.W., L.M. and H.Z. searched the databases and extracted the data. X.C. and X.X. analyzed the data. Y.W. wrote the draft of the paper. Z.H. and L.M. reviewed the manuscript.

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Abbreviations

ACS, acute coronary syndrome; CAD, coronary artery disease; CHD, coronary heart disease; CI, confidence interval; CNKI, China National Knowledge Infrastructure; EL, endothelial lipase; HB, hospital-based; HDL, high-density lipoprotein; HDL-C, high-density lipoprotein cholesterol; HWE, Hardy–Weinberg equilibrium; LIPG, endothelial lipase gene; NOS, Newcastle–Ottawa Scale; OR, odds ratio; PB, population-based; PCR-RFLP, polymerase chain reaction-restriction fragment length polymorphism; SNP, single-nucleotide polymorphism; TG, triglyceride; TSA, trial sequential analysis.

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(score Study Adequa definition of patients	Selection (score)	Representativeness	Selection of	Definition	Comparability (score)	Exposure (score)	Same method of	Non-response	Total
	Adequate definition of patient case	of patients cases	controls	of control	Control for important factor or additional factor	Ascertainment of exposure	ascertainment for participants	Rate	Score
Zhu et al 2007 [12]	*	*	NA	*	**	*	*	NA	7
Cai et al 2014 [14]	*	*	NA	*	**	*	*	NA	7
Ji et al 2015 [23]	*	*	NA	*	*	*	*	NA	6
Jensen et al 2009 [24]	*	*	*	*	**	*	*	NA	8
Toosi et al 2015 [31]	*	*	NA	*	**	*	*	NA	7
Rimm et al 1992 [25]	*	*	*	*	**	*	*	NA	8
Colditz et al 1997 [26]	*	*	*	*	**	*	*	NA	8
Tjonneland et al 2007 [27]	*	*	*	*	**	*	*	NA	8
Tang et al 2008 [9]	*	*	NA	*	**	*	*	NA	7
Xie et al 2015 [13]	*	*	NA	*	**	*	*	NA	7
Solim et al 2018 [28]	*	*	NA	*	**	*	*	NA	7
Elnaggar et al 2019 [29]	*	*	NA	*	**	*	*	*	8
Dalan et al 2013 [30]	*	*	NA	*	**	*	*	NA	7

Supplemental Table 1 Methodological quality of the included studies according to the Newcastle-Ottawa Scale.