Research Article



Genetic polymorphism and transcriptional regulation of *CREBBP* gene in patient with diffuse large B-cell lymphoma

^(D) Haifeng Zhao^{1,*}, Yutian Kan^{1,*}, Xinyuan Wang¹, Leiyuan Chen¹, Peng Ge² and Zhengzi Qian³

¹Department of Hematology and Oncology, Tianjin Medical University Cancer Institute and Hospital, National Clinical Research Center for Cancer, Key Laboratory of Cancer Prevention and Therapy, Tianjin Clinical Research Center for Cancer, Tianjin, P.R. China; ²Department of Laboratory, Tianjin Medical University Cancer Institute and Hospital, National Clinical Research Center for Cancer, Key Laboratory of Cancer Prevention and Therapy, Tianjin Clinical Research Center for Cancer, Tianjin, P.R. China; ³Department of Lymphoma, Tianjin Medical University Cancer Institute and Hospital, National Clinical Research Center for Cancer, Key Laboratory, Tianjin Clinical Research Center for Cancer, Tianjin, P.R. China

Correspondence: Haifeng Zhao (zhaohaifeng457@163.com) or Zhengzi Qian (qianzhengzi123@163.com)



In the present study, we aim to examine the relationship between genetic polymorphism and transcriptional expression of cyclic AMP response element binding protein (CREBBP) and the risk of diffuse large B-cell lymphoma (DLBCL). Two hundred and fifty healthy individuals and 248 DLBCL patients participated in the present study. The CREBBP rs3025684 polymorphism was detected by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP). The mRNA expression of CREBBP was tested by the real-time quantitative PCR (RT-qPCR). The allele A frequency of CREBBP rs3025684 in DLBCL patients was obviously higher than that of controls (P=0.01). No significant difference was detected between CREBBP rs3025684 polymorphism and clinical characteristics of DLBCL patients when subgrouped according to different parameters. The results demonstrated that the allele A of CREBBP rs3025684 increased the susceptibility to DLBCL (P=0.004), with a worse overall survival (OS) rate (P=0.002), a worse progression-free survival (PFS) rate (P=0.033) and poor prognosis (P=0.003) in DLCBL patients. Furthermore, the expression of CREBBP mRNA was considerably decreased in DLBCL patients as compared with controls (P < 0.001), and the expression in patients with GG genotype was up-regulated in comparison with patients with GA and AA genotype (P=0.016 and P=0.001, respectively). However, no statistical differences were found in OS (P=0.201) and PFS (P=0.353) between the lower CREBBP mRNA level subgroup and higher CREBBP mRNA level subgroup. These data suggested that the CREBBP gene may be an important prognostic factor in DLBCL patients and perform an essential function in the development of DLBCL.

Introduction

Diffuse large B-cell lymphoma (DLBCL) is an aggressive non-Hodgkin lymphoma with extreme heterogeneity, accounting for 30–40% of newly diagnosed lymphomas [1]. Although the standard R-CHOP regimen has extremely good therapeutic effect on DLBCL patients, approximately 30–40% of patients show relapse and 10% have refractory disease [2]. In the past decades, accumulating evidences have shown the genetic, microenvironment, autoimmune diseases and occupational exposure participated in the pathogenesis of DLBCL [3–5]. With gene-expression profiling and next-generation sequencing, some common genetic loci are found to enmesh in the lymphomagenesis of DLBCL [6]. However, the pathogenesis of DLBCL is still not fully understood.

Single nucleotide polymorphism (SNP) is the most abundant type of genetic variation that occurs at a specific position in the human genome. Accumulating evidence indicated that SNPs have shown the

*These authors contributed equally to this work.

Received: 23 April 2019 Revised: 17 July 2019 Accepted: 30 July 2019

Accepted Manuscript Online: 31 July 2019 Version of Record published: 13 August 2019



ability to influence the activity and expression of the genes, and affect the pathogenesis and risk of an extensive range of cancer [7–10]. Previous studies have demonstrated that SNPs in *TP53*, *XRCC1* and *A20* are related to the risk of DLBCL [11–13]. Genome-wide association studies have recognized genetic susceptibility locus for DLBCL [14]. The contribution of SNP in histone-modifying enzymes to DLBCL pathogenesis is a research hotspot.

Somatic mutations in cyclic AMP response element binding protein (*CREBBP*) and *EP300*, and removal or inactivation of the HAT coding domain affect approximately 39% of DLBCL patients [15], and are also associated with Rubinstein Tyabi Syndrome (RTS) [16,17]. CREBBP belongs to the KAT3 family of histone/protein lysine acetyl-transferases, which is a highly conserved and universally expressed nuclear phosphoprotein [18,19]. *CREBBP* inactivation expedites GC-derived pathogenesis of lymphoma [20]. Down-regulation of CREBBP is related to worse overall survival (OS) rate in pediatric acute lymphoblastic leukemia and may affect the response to chemotherapy [21]. *Crebbp*^{+/-} mice had blemish in the development of B-cell lymphoid and an increased incidence of hematopoietic malignancy [22]. The rs3025684 (A/G) is an SNP of *CREBBP* located in intron 21. This SNP was shown to be a possible risk factor for developing autism in the Netherlands, U.K. and Denmark, as well as among Bengali-Hindus [23,24].

However, the status of *CREBBP* rs3025684 SNP and the gene expression in DLBCL in a Chinese Han population is not completely understood. It was hypothesized that *CREBBP* rs3025684 polymorphism and expression may be related to the susceptibility and pathogenesis of DLBCL, and the results found that it may be a prognostic factor for DLBCL patients.

Materials and methods Subjects

The present study recruited 250 healthy individuals and 248 DLBCL patients, diagnosed according to the World Health Organization classification [25] at Tianjin Medical University Cancer Institute and Hospital from 2011 to 2013. Peripheral blood and lymphoid specimens were collected before initial therapy. All participants signed the informed consent, and the approval of the study was obtained from the Tianjin Cancer Institute Institutional Review Board. The clinical and pathological characteristics of the patients and controls are presented in Table 1.

Extraction of DNA and genotyping of CREBBP rs3025684

The TIANamp genomic DNA kit (TIANGEN Biotech, Beijing, China) was used to extract the genomic DNA according to the manufacturer's protocol. Genotypes were analyzed by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP). The reaction system contained 60 ng DNA template, 2 µl of each primer, 25 µl Premix Taq (Takara, Dalian, China) and sterilized water up to 50 µl. The PCR protocol was as follows: initial denaturation at 94°C for 5 min, followed by 30 cycles of denaturation at 94°C for 30 s, annealing at 58°C for 30 s, extension at 72°C for 30 s, followed by 72°C for 7 min. The primers for rs3025684 were: forward 5′-AGGGGAAACAACTCACCCTG-3′ and reverse 5′-CTGGTCTTGTGGTTCCGTGT-3′. The PCR product was digested by MnII (New England Biolabs, Beverly, MA) and analyzed by gel electrophoresis on 2.5% agarose gels. Randomly selected DNA samples were detected by direct sequencing to verify the results (Supplementary Figure S1).

Isolation of RNA and reverse-transcription quantitative PCR

Total RNA was isolated using TRIzol reagent (Invitrogen). We normalized the levels of expression of *CREBBP* relative to glyceraldehyde phosphate dehydrogenase (GAPDH), and $2^{-\Delta\Delta C}_{t}$ indicated the quantification of gene expression. The primers for *CREBBP* were: forward 5'-CGGCTCTAGTATCAACCCAGG-3' and reverse 5'- TTTTGTGCTT GCGGATTCAAGT-3'. The primers for $\langle 0:italic \rangle$ GAPDH $\langle /0:italic \rangle$ were:forward5'-CCACATCGCTCAGACACCAT-3' and reverse 5'- TTTTGTGCTT.

Statistical analysis

A goodness-of-fit Chi-square test was used to analyze the Hardy–Weinberg equilibrium. The Chi-square test was used to detect the allelic frequency and genotypic distribution in all subjects. We estimated the relationship between *CREBBP* rs3025684 and the susceptibility to DLBCL by unconditional logistic regression. The prognostic factors were explored by univariate Cox regression analysis. The *CREBBP* mRNA expression levels were investigated by the independent Student's *t* test. The survival curves were computed by Kaplan–Meier method with log-rank tests. All the statistical analyses mentioned above were executed by IBM SPSS Statistics version 20.0. *P*<0.05 was considered to be statistically significant. The false-positive report probability (FPRP) was calculated as previously described [26], we set an FPRP value of 0.2 and assigned a prior probability of 0.1.



Table 1 Clinical characteristics of DLBCL patients

Characteristics	Number (<i>n</i> =248) <i>n</i> (%)	Control (<i>n</i> =250) <i>n</i> (%)
Gender		
Male	112 (45.16)	109 (43.60)
Female	136 (54.84)	141 (56.40)
Age		
≤60	165 (66.53)	184 (73.60)
>60	83 (33.47)	66 (26.40)
Subtype		
GCB	75 (30.24)	
nGCB	173 (69.76)	
Ann-arbor stage		
I–II	152 (61.29)	
III–IV	96 (38.71)	
IPI score		
0–1	133 (53.63)	
2–5	115 (46.37)	
ECOG		
0–1	230 (92.74)	
2–5	18 (7.26)	
B symptom		
+	79 (31.85)	
	169 (68.15)	
Extra nodal sites	(50,00)	
+	156 (62.90)	
- -	92 (37.10)	
Bone marrow involvement	01 (0.47)	
+	21 (8.47)	
- HBV infection	227 (91.53)	
	74 (29.84)	
+	174 (23.04)	
Bulky tumor (>10 cm)	174 (70.10)	
+	55 (22.18)	
т	193 (77.82)	
Elevated LDH	133 (11.02)	
+	98 (39.52)	
_	150 (60.48)	
Elevated β2-MG	100 (00.+0)	
+	86 (34.68)	
_	162 (65.32)	
KI-67		
≤75%	93 (37.50)	
>75%	155 (62.50)	
Source	(02.00)	
Gastrointestinal	116 (46.77)	
Others	132 (53.23)	
Response rate		
PR+CR	215 (86.69)	

Abbreviations: CR, complete response; ECOG, Eastern Cooperative Oncology Group; IPI, International Prognostic Index; LDH, lactate dehydrogenase; PR, partial response; β2-MG, β2 macroglobulin.

Results Characteristics of study subjects

The genotypic distributions in both patients and controls were under the Hardy–Weinberg equilibrium (P>0.05). The genotypic distribution and allelic frequencies showed significant difference between the DLBCL patients and



	Control (n=250) n (%)	DLBCL patients (n=248) n (%)	Р
Genotype frequency			
GG	153 (61.20)	121 (48.79)	0.021
GA	86 (34.40)	113 (45.56)	
AA	11 (4.40)	14 (5.65)	
Allele frequency			
G	392 (78.40)	355 (71.57)	0.013
А	108 (21.60)	141 (28.43)	

Table 2 Genotype distribution and allele frequencies of CREBBP rs3025684 polymorphism in DLBCL patients and the controls

healthy individuals for *CREBBP* rs3025684 (P=0.021 and P=0.013, respectively, shown in Table 2). Furthermore, the association between the genotype and the clinical parameters of DLBCL patients was explored, which showed no statistical differences as shown in Table 3.

Relationship between the *CREBBP* rs3025684 and the susceptibility to **DLBCL**

The GA genotype was correlated with the risk of DLBCL (odds ratio (OR) = 1.692, 95% confidence interval (95% CI) = 1.170-2.446, P=0.005) (Table 4). However, the AA genotype displayed a slightly increased susceptibility to DLBCL with no statistical significance (OR = 1.620, 95% CI = 0.710-3.696, P=0.252, P=0.252) (Table 4). The combined AA and GA genotype was significantly related to increased susceptibility to DLBCL (OR = 1.684, 95% CI = 1.179-2.404, P=0.004) (Table 4). The results for dominant and recessive models are shown in Table 4. As no statistical significance was shown in the dominant model, we did not calculate the FPRP values and statistical power for the dominant model. Positive association was observed in the recessive model and GA genotype as their FPRP value was less than 0.2 (shown in Table 4).

Survival analysis of DLBCL patients according to the CREBBP rs3025684

The survival analysis of all 248 DLBCL patients showed that the patients with genotype GA/AA (P=0.006, Figure 1A) and the combined GA and AA group (P=0.002, Figure 1B) had a worse OS than patients with GG genotype. Furthermore, the patients with GA/AA genotype had a worse progression-free survival (PFS) in comparison with the GG patients with no statistical significance (P=0.058, Figure 1C), however, the combined GA and AA group showed worse PFS rate compared with the GG group with statistically significant (P=0.033, Figure 1D). Furthermore, it was also indicated that patients with A allele showed poor prognosis (P=0.003, HR = 1.944, 95% CI = 1.247–3.029).

Analysis of the CREBBP mRNA expression levels

CREBBP expression was detected in 63 patients and 32 controls. The results showed that the CREBBP expression was remarkably down-regulated in patients as compared with controls (P<0.001, Figure 2A). The *CREBBP* expression of patients with GG genotype was down-regulated as compared with the patients with GA and AA genotype (P=0.016 and 0.001, respectively, Figure 2B). However, no significant difference was detected between the GA and AA subgroups (P=0.134, Figure 2B). The *CREBBP* expression was down-regulated in the GA/AA subgroup as compared with the GG subgroup (P=0.002, Figure 2C).

Survival analysis based on the CREBBP mRNA levels of DLBCL patients

Based on the median of *CREBBP* mRNA expression value, 63 DLBCL patients were partitioned into two subgroups, the low *CREBBP* expression subgroup and the high *CREBBP* expression subgroup. The results indicated no significant differences between the patients with lower *CREBBP* mRNA level and those with high *CREBBP* mRNA level in OS (P=0.201, Figure 3A) and PFS (P=0.353, Figure 3B).

Discussion

DLBCL is an example of a translocation-based cancer, in which key genes are dysregulated by active lineage-specific promoters or enhancers due to characteristic equilibrium translocations [6]. The evolution of DLBCL was a multi-step procedure that requires the cumulation of multiple genetic pathological changes [14,27]. By the modern genome-wide



Table 3 Characteristics of DLBCL patients and their association with CREBBP rs3025684

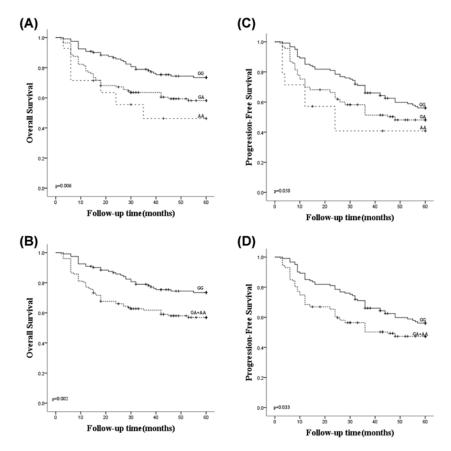
Characteristics	Number	Genotype		Р	
	(n=248) n (%)	GA+AA, n (%)	GG, n (%)		
Gender					
Male	112 (45.16)	61 (24.60)	51 (20.56)	0.352	
Female	136 (54.84)	66 (26.61)	70 (28.23)		
Age					
≤60	165 (66.53)	87 (35.08)	78 (31.45)	0.500	
>60	83 (33.47)	40 (16.13)	43 (17.34)		
Subtype					
GCB	75 (30.24)	38 (15.32)	37 (14.92)	0.910	
nGCB	173 (69.76)	89 (35.89)	84 (33.87)		
Ann-arbor stage					
I–II	152 (61.29)	73 (29.44)	79 (31.85)	0.207	
III–IV	96 (38.71)	54 (21.77)	42 (16.94)		
IPI score					
0–1	133 (53.63)	62 (25.00)	71 (28.63)	0.120	
2–5	115 (46.37)	65 (26.21)	50 (20.16)		
ECOG					
0–1	230 (92.74)	114 (45.97)	116 (46.77)	0.064	
2–5	18 (7.26)	13 (5.24)	5 (2.02)		
B symptom					
+	79 (31.85)	43 (17.34)	36 (14.52)	0.488	
_	169 (68.15)	84 (33.87)	85 (34.27)		
Bone marrow involvement					
+	21 (8.47)	11 (4.44)	10 (4.03)	0.911	
_	227 (91.53)	116 (46.77)	111 (44.76)		
HBV infection					
+	74 (29.84)	39 (15.73)	35 (14.11)	0.759	
_	174 (70.16)	88 (35.48)	86 (34.68)		
Bulky tumor (>10 cm)					
+	55 (22.18)	31 (12.50)	24 (9.68)	0.386	
_	193 (77.82)	96 (38.71)	97 (39.11)		
Elevated LDH					
+	98 (39.52)	57 (22.98)	41 (16.53)	0.077	
-	150 (60.48)	70 (28.23)	80 (32.26)		
Elevated β2-MG					
+	86 (34.68)	41 (16.53)	43 (17.34)	0.588	
-	162 (65.32)	86 (34.68)	78 (31.45)		
KI-67					
≤75%	93 (37.50)	50 (20.16)	43 (17.34)	0.533	
>75%	155 (62.50)	77 (31.05)	78 (31.45)		
Source					
Gastrointestinal	116 (46.77)	55 (22.18)	61 (24.60)	0.262	
Others	132 (53.23)	72 (29.03)	60 (24.19)		

Abbreviations: LDH, lactate dehydrogenase; ECOG, Eastern Cooperative Oncology Group; IPI, International Prognostic Index; β 2-MG, β 2 macroglobulin.

Table 4 Association between CREBBP rs3025684 polymorphism and the risk of DLBCL

Genotype	OR (95% CI)	Р	Statistical power		Prior probability				
				0.25	0.1	0.01	0.001	0.0001	
AA vs. GG	1.620 (0.710–3.696)	0.252							
GA vs.GG	1.692 (1.170-2.446)	0.005	0.500	0.030	0.085	0.505	0.0912	0.990	
GA/AA vs. GG	1.684 (1.179–2.404)	0.004	0.500	0.024	0.069	0.449	0.891	0.988	
GA/GG vs. AA	0.769 (0.342-1.729)	0.526							

© 2019 The Author(s). This is an open access article published by Portland Press Limited on behalf of the Biochemical Society and distributed under the Creative Commons Attribution License 4.0 (CC BY).





(A) OS analysis compared among the GG, GA and AA subgroups (P=0.006). (B) OS analysis compared between the GG subgroup and the combined GA/AA subgroup (P=0.002). (C) PFS analysis compared among the GG, GA and AA subgroups (P=0.058). (D) PFS analysis compared between the GG subgroup and the combined GA/AA subgroup (P=0.033). A check mark indicates that the data is censored.

molecular analysis, an abundance of altered cellular pathways that perform vital functions in the development of DLBCL and the sensitivity of cancer cells to therapy was detected. However, the notable heterogeneity of this disease partly limits the effective treatment. Therefore, it is necessary to seek ameliorated special biomarkers, permitting the devising of more efficient and accurate medical methods to specific cancer-causing addictions.

CREBBP, one of the most frequently mutated genes in DLBCL [15,28–30], acts as a tumor repressor of GC-derived pathogenesis of lymphoma [31]. Most of the *CREBBP*-binding regions in GC B cells displayed characteristics of transcriptionally active (or suspended) enhancers [32], which control the cell type-specific transcription [33]. CREBBP acetylates H3K27 on the promoter or enhancer sequence of the BCL6 target genes, promoting transcription to counteracting the inhibition of BCL6, which leads to the opposition of proto-oncogene activity of BCL6 [20,31]. Therefore, *CREBBP* may be considered as a tumor therapeutic target in DLBCL patients.

Gene expression-based categorization of DLBCL has been established [34,35], and the relevance to prognosis has been illustrated [36]. It has been demonstrated that certain genetic defects and distinct signal transduction pathways occur in specific subtypes [37,38]. In this study, the association between *CREBBP* rs3025684 polymorphism and its expression with the susceptibility and survival of DLBCL patients was investigated. The results showed that the A allele carriers were related to increased risk of DLBCL and had worse OS rate and PFS rate. Hence, *CREBBP* rs3025684 polymorphism may be used as a hallmark for the prediction of risk and prognosis of DLBCL.

In East Asians, the frequency of A allele of CREBBP rs3025684 was reported to be 19.3% in the HapMap Project. However, the present study showed that the frequency of allele A in controls and DLBCL patients was 21.60 and 28.43%, respectively. This difference in allele frequency may owe to the risk effects of allele A and the small sample



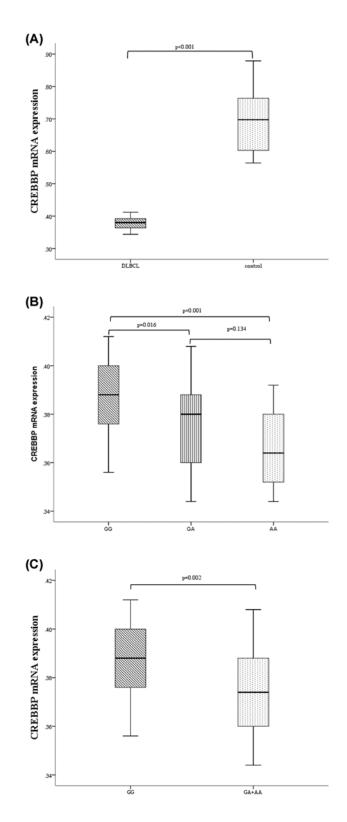


Figure 2. The expression of CREBBP mRNA in DLBCL patients

(A) The expression of *CREBBP* mRNA in DLBCL patients was down-regulated as compared with the controls (P<0.001). (B) The expression of *CREBBP* mRNA in DLBCL patients with GG genotype was significantly up-regulated than those with GA and AA genotypes (P=0.016 and 0.001, respectively). (C) The expression of *CREBBP* mRNA in DLBCL patients with GA/AA genotype was significantly down-regulated than those with GG genotype (P=0.002).

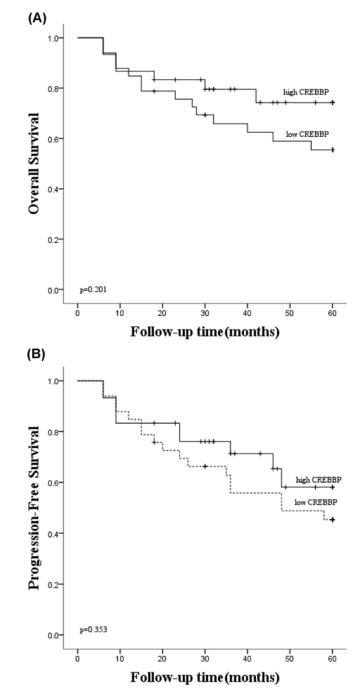


Figure 3. Survival analyses of DLBCL patients according to *CREBBP* mRNA expression
(A) The OS analysis indicated no significant differences between lower *CREBBP* subgroup and high *CREBBP* subgroup (*P*=0.201).
(B) The PFS analysis indicated no significant differences between the lower *CREBBP* subgroup and high *CREBBP* subgroup (*P*=0.353). A check mark indicates that the data is censored.

size. In the recessive model, the positive association between the risk of DLBCL and CREBBP rs3025684 was observed. However, the statistical power was only 0.5, which may be a result of the small simple size.

The CREBBP/EP300 complex participates in numerous life events, such as cell growth, proliferation, apoptosis, metabolism and oncogenesis [18,39,40]. CREBBP/EP300 complex also targets many transcriptional factors significantly related to the development of B-cell lymphoma and immune response, such as p53 and c-MYC [41–43]. HAT activity is related to the survival rate of patients with B-cell lymphoma [31,44]. Furthermore, HAT mutations likely



predict treatment efficiency in epigenetically targeted therapy, such as the HDAC3 inhibitors [20,45]. The present study indicated that the expression of *CREBBP* was notably down-regulated in patients as compared with the controls, and was especially down-regulated in the GA/AA genotype subgroup, indicating that CREBBP may be used as a therapeutic target for DLBCL.

In conclusion, the polymorphism and expression of *CREBBP* gene may play a vital role as a genetic risk factor and poor prognostic factor among DLBCL Chinese Han patients, indicating that CREBBP could be used for the prognosis and treatment of DLBCL. Additional studies with larger sample sizes are necessary to verify the results and further functional analyses are warranted to explore the lymphoma biology.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Funding

This work was supported by the National Natural Science Foundation of China [grant numbers 81100337, 81470283].

Author Contribution

Z.Q. and H.Z. designed the research. Y.K. performed the experiment, analyzed the experimental data and wrote the manuscript. H.Z., P.G. and X.W. helped to analyze the experimental data and collected the samples. L.C. gathered the clinical information. All authors read and approved the final manuscript.

Abbreviations

CREBBP, cyclic AMP response element binding protein; DLBCL, diffuse large B-cell lymphoma; FPRP, false-positive report probability; GAPDH, glyceraldehyde phosphate dehydrogenase; HAT, histone acetyltransferase; HDAC3, histone deacetylase 3; HR, hazard ratio; OR, odds ratio; OS, overall survival; PFS, progression-free survival; SNP, single nucleotide polymorphism; 95% CI, 95% confidence interval.

References

- 1 Lenz, G. and Staudt, L.M. (2010) Aggressive lymphomas. N. Engl. J. Med. 362, 1417–1429, https://doi.org/10.1056/NEJMra0807082
- 2 Coiffier, B., Thieblemont, C., Van Den Neste, E., Lepeu, G., Plantier, I., Castaigne, S. et al. (2010) Long-term outcome of patients in the LNH-98.5 trial, the first randomized study comparing rituximab-CHOP to standard CHOP chemotherapy in DLBCL patients: a study by the Groupe d'Etudes des Lymphomes de l'Adulte. *Blood* **116**, 2040–2045, https://doi.org/10.1182/blood-2010-03-276246
- 3 Morton, L.M., Slager, S.L., Cerhan, J.R., Wang, S.S., Vajdic, C.M., Skibola, C.F. et al. (2014) Etiologic heterogeneity among non-Hodgkin lymphoma subtypes: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. *J. Natl. Cancer Inst. Monogr.* 2014, 130–144, https://doi.org/10.1093/jncimonographs/lgu013
- 4 Cerhan, J.R. and Slager, S.L. (2015) Familial predisposition and genetic risk factors for lymphoma. *Blood* **126**, 2265–2273, https://doi.org/10.1182/blood-2015-04-537498
- 5 Scott, D.W. and Gascoyne, R.D. (2014) The tumour microenvironment in B cell lymphomas. *Nat. Rev. Cancer* **14**, 517–534, https://doi.org/10.1038/nrc3774
- 6 Armitage, J.O., Gascoyne, R.D., Lunning, M.A. and Cavalli, F. (2017) Non-Hodgkin lymphoma. Lancet **390**, 298–310, https://doi.org/10.1016/S0140-6736(16)32407-2
- 7 Abd El-Fattah, A.A., Sadik, N.A.H., Shaker, O.G. and Mohamed Kamal, A. (2018) Single nucleotide polymorphism in SMAD7 and CHI3L1 and colorectal cancer risk. *Mediators Inflamm.* 2018, 9853192, https://doi.org/10.1155/2018/9853192
- 8 Wen, J., Lv, Z., Ding, H., Fang, X. and Sun, M. (2018) Association of miRNA biosynthesis genes DROSHA and DGCR8 polymorphisms with cancer susceptibility: a systematic review and meta-analysis. *Biosci. Rep.* 38, https://doi.org/10.1042/BSR20180072
- 9 Liu, C., Cui, H., Gu, D., Zhang, M., Fang, Y., Chen, S. et al. (2017) Genetic polymorphisms and lung cancer risk: Evidence from meta-analyses and genome-wide association studies. *Lung Cancer* **113**, 18–29, https://doi.org/10.1016/j.lungcan.2017.08.026
- 10 Liu, G.C., Zhou, Y.F., Su, X.C. and Zhang, J. (2019) Interaction between TP53 and XRCC1 increases susceptibility to cervical cancer development: a case control study. *BMC Cancer* **19**, 24, https://doi.org/10.1186/s12885-018-5149-0
- 11 Li, Y., Bai, O., Cui, J. and Li, W. (2016) Genetic polymorphisms in the DNA repair gene, XRCC1 associate with non-Hodgkin lymphoma susceptibility: a systematic review and meta-analysis. *Eur. J. Med. Genet.* **59**, 91–103, https://doi.org/10.1016/j.ejmg.2015.12.011
- 12 Wenzl, K., Hofer, S., Troppan, K., Lassnig, M., Steinbauer, E., Wiltgen, M. et al. (2016) Higher incidence of the SNP Met 788 lle in the coding region of A20 in diffuse large B cell lymphomas. *Tumour Biol.* **37**, 4785–4789, https://doi.org/10.1007/s13277-015-4322-1
- 13 Liu, Y., Wang, X., Ding, N., Mi, L., Ping, L., Jin, X. et al. (2017) TP53 Arg72 as a favorable prognostic factor for Chinese diffuse large B-cell lymphoma patients treated with CHOP. *BMC Cancer* **17**, 743, https://doi.org/10.1186/s12885-017-3760-0
- 14 Cerhan, J.R., Berndt, S.I., Vijai, J., Ghesquieres, H., McKay, J., Wang, S.S. et al. (2014) Genome-wide association study identifies multiple susceptibility loci for diffuse large B cell lymphoma. *Nat. Genet.* **46**, 1233–1238, https://doi.org/10.1038/ng.3105



- 15 Pasqualucci, L., Dominguez-Sola, D., Chiarenza, A., Fabbri, G., Grunn, A., Trifonov, V. et al. (2011) Inactivating mutations of acetyltransferase genes in B-cell lymphoma. *Nature* **471**, 189–195, https://doi.org/10.1038/nature09730
- 16 Bartsch, O., Schmidt, S., Richter, M., Morlot, S., Seemanova, E., Wiebe, G. et al. (2005) DNA sequencing of CREBBP demonstrates mutations in 56% of patients with Rubinstein-Taybi syndrome (RSTS) and in another patient with incomplete RSTS. *Hum. Genet.* **117**, 485–493, https://doi.org/10.1007/s00439-005-1331-y
- 17 Stef, M., Simon, D., Mardirossian, B., Delrue, M.A., Burgelin, I., Hubert, C. et al. (2007) Spectrum of CREBBP gene dosage anomalies in Rubinstein-Taybi syndrome patients. *Eur. J. Hum. Genet.* **15**, 843–847, https://doi.org/10.1038/sj.ejhg.5201847
- 18 Goodman, R.H. and Smolik, S. (2000) CBP/p300 in cell growth, transformation, and development. *Genes Dev.* 14, 1553–1577
- 19 Kalkhoven, E. (2004) CBP and p300: HATs for different occasions. Biochem. Pharmacol. 68, 1145–1155, https://doi.org/10.1016/j.bcp.2004.03.045
- 20 Jiang, Y., Ortega-Molina, A., Geng, H., Ying, H.Y., Hatzi, K., Parsa, S. et al. (2017) CREBBP inactivation promotes the development of HDAC3-dependent lymphomas. *Cancer Discov.* 7, 38–53, https://doi.org/10.1158/2159-8290.CD-16-0975
- 21 Gao, C., Zhang, R.D., Liu, S.G., Zhao, X.X., Cui, L., Yue, Z.X. et al. (2017) Low CREBBP expression is associated with adverse long-term outcomes in paediatric acute lymphoblastic leukaemia. *Eur. J. Haematol.* **99**, 150–159, https://doi.org/10.1111/ejh.12897
- 22 Kung, A.L., Rebel, V.I., Bronson, R.T., Ch'ng, L.E., Sieff, C.A., Livingston, D.M. et al. (2000) Gene dose-dependent control of hematopoiesis and hematologic tumor suppression by CBP. Genes Dev. 14, 272–277
- 23 Barnby, G., Abbott, A., Sykes, N., Morris, A., Weeks, D.E., Mott, R. et al. (2005) Candidate-gene screening and association analysis at the autism-susceptibility locus on chromosome 16p: evidence of association at GRIN2A and ABAT. Am. J. Hum. Genet. 76, 950–966, https://doi.org/10.1086/430454
- 24 Kumar, D., Deb, I., Chakraborty, J., Mukhopadhyay, S. and Das, S. (2011) A polymorphism of the CREB binding protein (CREBBP) gene is a risk factor for addiction. *Brain Res.* **1406**, 59–64, https://doi.org/10.1016/j.brainres.2011.05.048
- 25 Turner, J.J., Hughes, A.M., Kricker, A., Milliken, S., Grulich, A., Kaldor, J. et al. (2005) WHO non-Hodgkin's lymphoma classification by criterion-based report review followed by targeted pathology review: an effective strategy for epidemiology studies. *Cancer Epidemiol. Biomarkers Prev.* 14, 2213–2219, https://doi.org/10.1158/1055-9965.EPI-05-0358
- 26 Wacholder, S., Chanock, S., Garcia-Closas, M., El Ghormli, L. and Rothman, N. (2004) Assessing the probability that a positive report is false: an approach for molecular epidemiology studies. *J. Natl. Cancer Inst.* **96**, 434–442, https://doi.org/10.1093/jnci/djh075
- 27 Schmitz, R., Wright, G.W., Huang, D.W., Johnson, C.A., Phelan, J.D., Wang, J.Q. et al. (2018) Genetics and pathogenesis of diffuse large B-cell lymphoma. N. Engl. J. Med. 378, 1396–1407, https://doi.org/10.1056/NEJMoa1801445
- 28 Lohr, J.G., Stojanov, P., Lawrence, M.S., Auclair, D., Chapuy, B., Sougnez, C. et al. (2012) Discovery and prioritization of somatic mutations in diffuse large B-cell lymphoma (DLBCL) by whole-exome sequencing. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 3879–3884, https://doi.org/10.1073/pnas.1121343109
- 29 Morin, R.D., Mendez-Lago, M., Mungall, A.J., Goya, R., Mungall, K.L., Corbett, R.D. et al. (2011) Frequent mutation of histone-modifying genes in non-Hodgkin lymphoma. *Nature* 476, 298–303, https://doi.org/10.1038/nature10351
- 30 Pasqualucci, L., Trifonov, V., Fabbri, G., Ma, J., Rossi, D., Chiarenza, A. et al. (2011) Analysis of the coding genome of diffuse large B-cell lymphoma. *Nat. Genet.* **43**, 830–837, https://doi.org/10.1038/ng.892
- 31 Zhang, J., Vlasevska, S., Wells, V.A., Nataraj, S., Holmes, A.B., Duval, R. et al. (2017) The CREBBP acetyltransferase is a haploinsufficient tumor suppressor in B-cell lymphoma. *Cancer Discov.* 7, 322–337, https://doi.org/10.1158/2159-8290.CD-16-1417
- 32 Heinz, S., Romanoski, C.E., Benner, C. and Glass, C.K. (2015) The selection and function of cell type-specific enhancers. *Nat. Rev. Mol. Cell Biol.* **16**, 144–154, https://doi.org/10.1038/nrm3949
- 33 Hnisz, D., Abraham, B.J., Lee, T.I., Lau, A., Saint-Andre, V., Sigova, A.A. et al. (2013) Super-enhancers in the control of cell identity and disease. *Cell* **155**, 934–947, https://doi.org/10.1016/j.cell.2013.09.053
- 34 Alizadeh, A.A., Eisen, M.B., Davis, R.E., Ma, C., Lossos, I.S., Rosenwald, A. et al. (2000) Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. *Nature* **403**, 503–511, https://doi.org/10.1038/35000501
- 35 Monti, S., Savage, K.J., Kutok, J.L., Feuerhake, F., Kurtin, P., Mihm, M. et al. (2005) Molecular profiling of diffuse large B-cell lymphoma identifies robust subtypes including one characterized by host inflammatory response. *Blood* **105**, 1851–1861, https://doi.org/10.1182/blood-2004-07-2947
- 36 Lenz, G., Wright, G., Dave, S.S., Xiao, W., Powell, J., Zhao, H. et al. (2008) Stromal gene signatures in large-B-cell lymphomas. *N. Engl. J. Med.* **359**, 2313–2323, https://doi.org/10.1056/NEJMoa0802885
- 37 Davis, R.E., Ngo, V.N., Lenz, G., Tolar, P., Young, R.M., Romesser, P.B. et al. (2010) Chronic active B-cell-receptor signalling in diffuse large B-cell lymphoma. *Nature* 463, 88–92, https://doi.org/10.1038/nature08638
- 38 Ngo, V.N., Young, R.M., Schmitz, R., Jhavar, S., Xiao, W., Lim, K.H. et al. (2011) Oncogenically active MYD88 mutations in human lymphoma. *Nature* **470**, 115–119, https://doi.org/10.1038/nature09671
- 39 Giordano, A. and Avantaggiati, M.L. (1999) p300 and CBP: partners for life and death. *J. Cell. Physiol.* **181**, 218–230, https://doi.org/10.1002/(SICI)1097-4652(199911)181:2%3c218::AID-JCP4%3e3.0.C0;2-5
- 40 Giles, R.H., Peters, D.J. and Breuning, M.H. (1998) Conjunction dysfunction: CBP/p300 in human disease. *Trends Genet.* 14, 178–183, https://doi.org/10.1016/S0168-9525(98)01438-3
- 41 Gu, W. and Roeder, R.G. (1997) Activation of p53 sequence-specific DNA binding by acetylation of the p53 C-terminal domain. *Cell* **90**, 595–606, https://doi.org/10.1016/S0092-8674(00)80521-8
- 42 Vervoorts, J., Luscher-Firzlaff, J.M., Rottmann, S., Lilischkis, R., Walsemann, G., Dohmann, K. et al. (2003) Stimulation of c-MYC transcriptional activity and acetylation by recruitment of the cofactor CBP. *EMBO Rep.* **4**, 484–490, https://doi.org/10.1038/sj.embor.embor821
- 43 Weaver, B.K., Kumar, K.P. and Reich, N.C. (1998) Interferon regulatory factor 3 and CREB-binding protein/p300 are subunits of double-stranded RNA-activated transcription factor DRAF1. *Mol. Cell. Biol.* **18**, 1359–1368, https://doi.org/10.1128/MCB.18.3.1359



- 44 Juskevicius, D., Jucker, D., Klingbiel, D., Mamot, C., Dirnhofer, S. and Tzankov, A. (2017) Mutations of CREBBP and SOCS1 are independent prognostic factors in diffuse large B cell lymphoma: mutational analysis of the SAKK 38/07 prospective clinical trial cohort. *J. Hematol. Oncol.* **10**, 70, https://doi.org/10.1186/s13045-017-0438-7
- 45 Andersen, C.L., Asmar, F., Klausen, T., Hasselbalch, H. and Gronbaek, K. (2012) Somatic mutations of the CREBBP and EP300 genes affect response to histone deacetylase inhibition in malignant DLBCL clones. *Leukemia Res. Rep.* **2**, 1–3

