

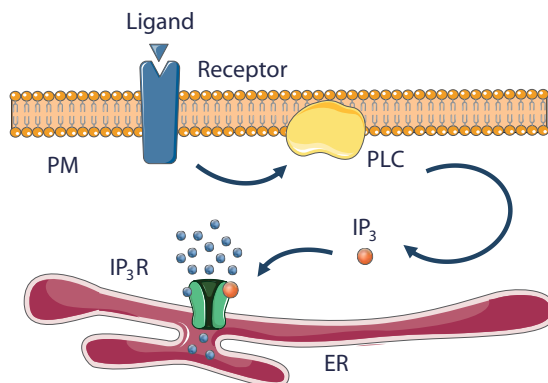
The where and when of intracellular Ca^{2+} signalling

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Ca^{2+} is essential for cellular functions. The ability of this ion to act as an intracellular signal, common to almost all eukaryotic cells, allows it to orchestrate an enormous repertoire of cellular activities, from initiating egg development after fertilization to synaptic transmission between neurons.

Figure 1. The path from extracellular stimulus to intracellular Ca^{2+} release. Many extracellular stimuli activate receptors that signal through phospholipase C (PLC). PLC generates inositol 1,4,5-trisphosphate (IP_3), a diffusible intracellular messenger molecule that opens IP_3 receptors (IP_3Rs), which are Ca^{2+} -permeable channels in the endoplasmic reticulum (ER) membrane. IP_3Rs are the signalling hubs that control the redistribution of Ca^{2+} from internal stores to the cytosol, other organelles and the flow of Ca^{2+} across the plasma membrane through store-operated Ca^{2+} entry. Created with BioRender.com

The first indication that Ca^{2+} has a role as a dynamic physiological signal, beyond its role as a structural component of teeth and bones, was in an experiment performed by British cardiovascular physiologist Sydney Ringer in 1883, just 14 years after Dmitri Mendeleev proposed his periodic table of the elements. In the absence of Ca^{2+} salts in a circulating saline solution, Ringer found that contractions of isolated frog hearts progressively weakened and then ceased. The contractions resumed when Ca^{2+} was restored. We know now that these serendipitous observations reflect the need, during each cycle of contraction and relaxation, for Ca^{2+} release and re-uptake from intracellular stores. The need for extracellular Ca^{2+} arises because the ryanodine receptors (RyR), through which the Ca^{2+} is released, open when they bind Ca^{2+} . This triggers Ca^{2+} is provided via the opening of voltage-gated Ca^{2+} channels in the plasma membrane (PM).



Nearly 150 years later, the role of Ca^{2+} as an intracellular messenger is established in processes as diverse as muscle contraction, neurotransmitter secretion, gene transcription, cell division and apoptosis. Ca^{2+} is unusual among intracellular messengers in that it is an ion, rather than a more complex component that might be made or degraded, and so changes in the free cytosolic Ca^{2+} concentration (abbreviated to $[\text{Ca}^{2+}]_c$) can only occur by moving Ca^{2+} across membranes. Ion channels allow Ca^{2+} to move quickly down its electrochemical gradient, while ATP-dependent pumps or exchange proteins can move Ca^{2+} against its electrochemical gradient to extrude it from the cell or return it to stores. The complex sequence of conformational changes associated with active Ca^{2+} transport means that Ca^{2+} pumps typically transport Ca^{2+} thousands of times more slowly than ion channels. Hence, most acute increases in $[\text{Ca}^{2+}]_c$ are brought about by the regulated opening of Ca^{2+} -permeable channels. One of the most widely expressed of these channels is the inositol 1,4,5-trisphosphate receptor (IP_3R), a cousin of RyR. Similar to RyRs, IP_3Rs are stimulated to open by Ca^{2+} , but for IP_3Rs this only occurs after they have bound IP_3 .

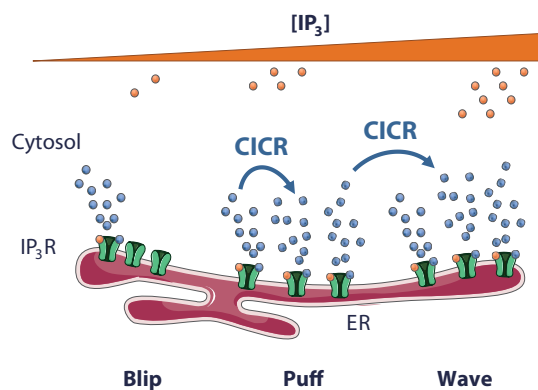
The ER is the major intracellular Ca^{2+} store, and since most IP_3Rs are expressed within ER membranes, IP_3Rs allow Ca^{2+} to be redistributed from the ER to the cytosol. The link with extracellular signals, like hormones or growth factors, is provided by receptors in the PM that can stimulate phospholipase C (PLC). The IP_3 produced by PLC binds to IP_3Rs causing them to open and release Ca^{2+} from the ER, with several important consequences (Figure 1). This action increases $[\text{Ca}^{2+}]_c$, which can

then regulate cellular activities often through the ubiquitous Ca^{2+} -binding protein, calmodulin. As IP_3Rs are stimulated by Ca^{2+} once they have bound IP_3 , they can propagate cytosolic Ca^{2+} signals regeneratively. IP_3Rs held close to other organelles, mitochondria for example, can deliver Ca^{2+} to them and regulate their activities. Finally, loss of Ca^{2+} from the ER lumen stimulates store-operated Ca^{2+} entry (SOCE) across the PM. SOCE occurs when the luminal Ca^{2+} -binding site of another ER protein, STIM1, senses the loss of Ca^{2+} , causing it to unfold the cytosolic domain through which it binds to a Ca^{2+} channel in the PM, Orai1. This direct interaction between STIM1 and Orai1 opens the Ca^{2+} channel allowing Ca^{2+} to flow into the cell across the PM. Hence, it is clear that IP_3Rs serve as hubs that control the redistribution of Ca^{2+} from the ER to the cytosol or other organelles, and ultimately the flow of Ca^{2+} across the PM.

Cells are bombarded with extracellular stimuli that control every cellular activity, and much of this communication passes through Ca^{2+} signals; so how is specificity maintained? The answer seems to lie largely with the spatial and temporal organization of Ca^{2+} signals. Where and when Ca^{2+} signals are generated is just as important as how they are generated.

Location, location, location

Cytosolic Ca^{2+} buffers are so abundant that only one of every 100 Ca^{2+} ions to flow into the cytosol remains unbound. This buffering slows Ca^{2+} diffusion and allows local Ca^{2+} signals to persist around the mouths of open Ca^{2+} channels. These microdomains of increased $[\text{Ca}^{2+}]_c$ can selectively regulate Ca^{2+} -binding proteins situated within them. Many of these Ca^{2+} -binding proteins can ignore small changes in $[\text{Ca}^{2+}]_c$ and tune-in to the much larger changes within a microdomain because they have a relatively low affinity for Ca^{2+} . Within neurons, for example, such microdomains allow Ca^{2+} signals to selectively regulate neurotransmitter release or gene transcription, and in many cells they allow the Ca^{2+} signals evoked by SOCE or IP_3Rs to control different events. A now classic example of the importance of spatial organization is provided by the smooth muscle of blood vessels, where global increases in $[\text{Ca}^{2+}]_c$ trigger contraction by activating, via calmodulin, myosin light-chain kinase. But local Ca^{2+} signals provided by RyRs beneath the PM deliver their Ca^{2+} to Ca^{2+} -activated K^+ channels, causing them to open and allow K^+ to leave the cell. The resulting hyperpolarization of the PM prevents activation of voltage-gated Ca^{2+} channels, and by attenuating global increases in $[\text{Ca}^{2+}]_c$, causes relaxation. Hence, Ca^{2+} signals can, according to their location, cause relaxation or contraction.



For IP_3Rs , the local Ca^{2+} signals evoked by the opening of single channels can grow into larger ones because IP_3Rs are stimulated by both IP_3 and Ca^{2+} , so that Ca^{2+} released by one IP_3R can ignite the activity of its neighbours to generate a ' Ca^{2+} puff' as a small cluster of IP_3Rs open together (Figure 2). The 'gain' on this regenerative Ca^{2+} -induced Ca^{2+} release (CICR) mechanism is set by the concentration of IP_3 (which primes IP_3Rs to respond to Ca^{2+}) and by the distance between clustered IP_3Rs . Ca^{2+} puffs become more frequent as the IP_3 concentration increases and the Ca^{2+} signal can then spread, like a bushfire, across the entire cell as a regenerative wave. Recent work suggests that although Ca^{2+} puffs may be the basic building blocks of IP_3 -evoked Ca^{2+} signals, only a tiny fraction of the IP_3Rs in a cell are responsible for generating them. These 'licensed' IP_3Rs are strategically placed alongside the sites where STIM1 regulates SOCE, suggesting that the spatial organization of IP_3Rs may also contribute to regulating Ca^{2+} entry across the PM.

In textbooks, you might see organelles—such as the ER, mitochondria and lysosomes—represented as separate compartments. In reality, organelles form intimate contacts with each other, where their membranes are held just a few nanometres apart by scaffold proteins. These membrane contact sites mediate many exchanges between organelles, including the transfer of Ca^{2+} from

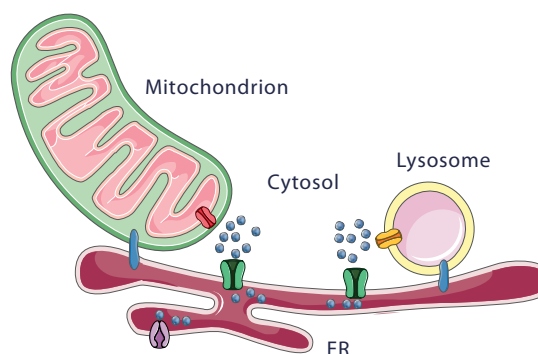


Figure 2. Local Ca^{2+} signals are the building blocks of global Ca^{2+} signals. IP_3Rs require the binding of both IP_3 and Ca^{2+} to open. This Ca^{2+} -induced Ca^{2+} release (CICR) is the basis for a hierarchical model of how local Ca^{2+} release events, such as blips (the result of a single IP_3R opening) and puffs (the concerted opening of IP_3R clusters), contribute to global signals, such as waves and spikes in response to increasing IP_3 concentration.

Figure 3. Intimate contact between organelles. Organelles form membrane contact sites that are held together by scaffolding proteins (blue). Ca^{2+} -uptake systems in mitochondria and lysosomes (red and orange) have low affinity for Ca^{2+} . The high-affinity uptake system of the ER, the sarco/endoplasmic reticulum Ca^{2+} -ATPase (SERCA, purple), actively accumulates Ca^{2+} within the ER lumen. IP_3Rs (green) sit next to membrane contact sites can then selectively deliver high concentrations of Ca^{2+} directly to organelles, allowing Ca^{2+} to mediate organelle-specific functions such as ATP synthesis by oxidative phosphorylation in mitochondria. Created with BioRender.com

one organelle to another, notably between the ER and mitochondria. Ca^{2+} uptake into mitochondria occurs via a channel, the mitochondrial uniporter (MCU), that only opens when it is exposed to much higher $[\text{Ca}^{2+}]_c$ than typically prevails in a healthy cell. Hence, mitochondria accumulate Ca^{2+} only when it is delivered to them by closely apposed Ca^{2+} channels—for example, IP_3Rs in the ER (Figure 3). The link with IP_3Rs is apt because IP_3 -evoked cytosolic Ca^{2+} signals will trigger cellular activity, for example smooth muscle contraction, while local delivery of Ca^{2+} to the outer mitochondrial membrane both parks mitochondria in regions where $[\text{Ca}^{2+}]_c$ is elevated and by driving Ca^{2+} uptake through MCU, it stimulates oxidative phosphorylation. IP_3 -evoked release of Ca^{2+} from the ER thereby both stimulates cellular activity and ensures delivery of the ATP required to sustain it.

Frequency codes

There is, however, a problem with the toxicity of Ca^{2+} ; sustained or excessive increase in $[\text{Ca}^{2+}]_c$ kill cells. However, natural selection has provided defences: most Ca^{2+} channels are inhibited by increases in $[\text{Ca}^{2+}]_c$, constraining their ability to poison the cell; and many Ca^{2+} extrusion mechanisms are stimulated by increased $[\text{Ca}^{2+}]_c$. So how can Ca^{2+} evoke sustained responses when powerful homeostatic mechanisms are engaged to prevent sustained increases in $[\text{Ca}^{2+}]_c$? Ca^{2+} oscillations or spikes provide an answer and, by encoding stimulus intensity in the frequency of Ca^{2+} spikes—just as nerves encode activity in the frequency of action potentials—they provide a robust means of communication. It is easier to distinguish a doubling of signal frequency than it is to distinguish a doubling of its amplitude, as the switch from AM (amplitude-modulated) to digital FM (frequency-modulated) radio attests.

To make the most of these digital Ca^{2+} signals, cells must decode them into graded and sustained responses. Ca^{2+} -calmodulin-dependent protein kinase II (CaMKII) provides a classic example of how a digital frequency-modulated signal can be transduced into an amplitude-modulated signal. CaMKII forms a complex structure with 12 subunits arranged around the spokes of 2 wheels with a shared axle. Binding of Ca^{2+} -calmodulin to a subunit activates its protein kinase activity, but it also primes phosphorylation of the subunit. Phosphorylation is important because it prolongs the kinase activity beyond the duration of the Ca^{2+} signal (it creates a memory of a preceding Ca^{2+} signal), but phosphorylation only occurs when two adjacent subunits have both bound Ca^{2+} -calmodulin: one subunit to provide the enzyme, and the other the substrate. Hence, appreciable

autophosphorylation occurs only when several of the 12 subunits are occupied. This tunes the initiation of the molecular memory to large Ca^{2+} signals, such as occur at the peak of Ca^{2+} spikes. The greater the frequency of the Ca^{2+} spikes, the less opportunity there is to reverse the phosphorylation between spikes and the greater the amplitude of the sustained CaMKII activity. Hence, CaMKII reads the frequency of Ca^{2+} spikes—indeed different forms of CaMKII are tuned to read different frequencies—and transduces them into graded increases in kinase activity.

However, as with any biological system that relies on relatively small numbers of molecules, there is an element of stochasticity to Ca^{2+} spikes. The interval between one spike and the next can vary randomly, and spiking frequencies show heterogeneity even between cells within the same population. So how can frequency reliably encode changes in stimulus intensity, given this randomness? In response to Ca^{2+} -mobilizing agonists, cells with variable spiking frequencies show a consistent *fold* change in the average interspike interval that scales with increasing stimulus strength. Drawing a musical analogy, although there are variations in ‘pitch’ (the absolute frequency of spikes), the change from one ‘note’ to another (the fold-change in frequency) is conserved. In this way, information about the initial stimulus intensity can be encoded in the temporal profile of Ca^{2+} transients.

Concluding remarks

Unlike most intracellular messengers, which can be made or degraded, Ca^{2+} is an element, and can only be redistributed from one cellular compartment to another. IP_3Rs are often responsible for this redistribution, delivering Ca^{2+} from the ER to the cytosol or other organelles, and initiating the events that stimulate Ca^{2+} entry across the PM. Hence, IP_3Rs are the hubs that coordinate the spatially and temporally organized Ca^{2+} signals that regulate so many cellular activities. ■

Further reading


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